

GLOBAL JOURNAL OF MEDICAL RESEARCH: C MICROBIOLOGY AND PATHOLOGY Volume 15 Issue 4 Version 1.0 Year 2015 Type: Double Blind Peer Reviewed International Research Journal Publisher: Global Journals Inc. (USA) Online ISSN: 2249-4618 & Print ISSN: 0975-5888

### Comparative Study of Techniques in the Diagnosis of Mycoplasma Pneumonia among the Patients of Respiratory Tract Infections in Northern Indian Population

By Dr. Abhineet Mehrotra, Dr. S. K. Mehra, Dr. M E Siddque & Dr. Sushil Suri

Geetanjali Medical College, Udaipur

*Abstract- Background:* In the present study an attempt was made to compare the different diagnosis of mycoplasma pneumonia among the patients of respiratory tract infections in northern Indian population. Commonly used techniques in the diagnosis of mycoplasma pneumonia are culture, serology and PCR and different studies assure the different sensitivity and specificity. In the present study author also aim to find the common risk factors of mycoplasma pneumonia.

*Material & Methods:* The present study was undertaken at Career Institute of Medical Science, Lucknow. The total number of study subjects were 193. The total number of male subjects were 115 (59.6 %) while 78 (40.4 %) were females. Samples were Collected for culture, serology and PCR. All samples of culture & serology were undergone for PCR.

*Results:* Out of 193 patients 58 (30 %) samples were found positive in culture and 73 (38 %) samples were found to be positive in serology test. 58 patients were found to be positive by both culture and serology which are further tested in PCR and only 10 (17 %) samples were found positive.

Keywords: mycoplasma pneumonia, culture, serology, PCR & risk factor.

GJMR-C Classification : NLMC Code: WC 246, WC 202

## COMPARATIVESTUDY OF TECHNIQUES IN THE DIAGNOSIS OF MYCOPLASMAPNEUMONIA AMONG THE PATIENTS OF RESPIRATORY TRACTINE ECTIONS IN NORTHERN INDIAN POPULATION

Strictly as per the compliance and regulations of:



© 2015. Dr. Abhineet Mehrotra, Dr. S. K. Mehra, Dr. M E Siddque & Dr. Sushil Suri. This is a research/review paper, distributed under the terms of the Creative Commons Attribution-Noncommercial 3.0 Unported License http://creativecommons.org/ licenses/by-nc/3.0/), permitting all non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

# Comparative Study of Techniques in the Diagnosis of Mycoplasma Pneumonia among the Patients of Respiratory Tract Infections in Northern Indian Population

Dr. Abhineet Mehrotra <sup>a</sup>, Dr. S. K. Mehra <sup>a</sup>, Dr. M E Siddque <sup>e</sup> & Dr. Sushil Suri <sup>a</sup>

Abstract- Background: In the present study an attempt was made to compare the different diagnosis of mycoplasma pneumonia among the patients of respiratory tract infections in northern Indian population. Commonly used techniques in the diagnosis of mycoplasma pneumonia are culture, serology and PCR and different studies assure the different sensitivity and specificity. In the present study author also aim to find the common risk factors of mycoplasma pneumonia.

*Material & Methods:* The present study was undertaken at Career Institute of Medical Science, Lucknow. The total number of study subjects were 193. The total number of male subjects were 115 (59.6 %) while 78 (40.4 %) were females. Samples were Collected for culture, serology and PCR. All samples of culture & serology were undergone for PCR.

*Results:* Out of 193 patients 58 (30 %) samples were found positive in culture and 73 (38 %) samples were found to be positive in serology test. 58 patients were found to be positive by both culture and serology which are further tested in PCR and only 10 (17 %) samples were found positive. The chi square test shows that respiratory & renal diseases may be considered as significant risk factors of Mycoplasma pneumonia (p<0.05).

*Keywords:* mycoplasma pneumonia, culture, serology, PCR & risk factor.

#### I. INTRODUCTION

ycoplasmas (mushroom form) are eubacteria included within the class Mollicutes (from latin mollis = "soft," cutis = "skin"), which comprises the smallest and simplest self-replicating, pleotrophic bacteria that lack cells wall thus, betalactam antibiotics. such as penicillin and cephalosporins, are ineffective. The name Mycoplasma, from the Greek μυκής, mykes (fungus) and πλάσμα, plasma (formed), was first used by Albert Bernhard Frank in 1889<sup>1</sup>. Mycoplasmas are primarily mucosal pathogens, living a parasitic existence in close

association with epithelial cells of their host, usually in the respiratory or urogenital tracts. M. pneumoniae exclusively parasitizes humans, whereas some of the other human mycoplasmas have also been recovered from nonhuman primates<sup>2</sup>. Mycoplasma pneumoniae is an important cause of respiratory tract infection and is increasingly being associated with other diseases such asthma and extra-pulmonary complications. as Considerable cross-reactivity is known to exist between the whole cell antigens used in the commercial serological testing assays. Usually, mycoplasma infection is a mild illness characterized by fever, cough, bronchitis, sore throat, and headache. In very rare cases, mycoplasma can cause serious illness such as encephalitis (an inflammation of the brain) or meningitis (inflammation of the lining of the brain and spinal cord).

As the diagnosis the clinical symptommatology of a mycoplasma infection does not show pathogenspecific characteristics, the diagnostic differentiation from other pathogens such as viruses and gram-positive bacteria is decisive for appropriate therapy<sup>3</sup>. The diagnosis can be based on direct detection and serology. Detection of the pathogen has been regarded as efficient diagnosis at acute, early stages of disease. Isolation of the pathogen by culture has been considered the reference method. However, it is too insensitive and time-consuming (6-14 days). A good guality pathogen DNA detection system (PCR) is not yet commercially available. To date, serology has been considered the method of choice for diagnosis of infections. The complement fixation test (CFT) represents the classic antibody detection. The CFT cannot discriminate between antibody isotypes. Agalutination tests cannot discriminate between antibody classes either. Both test systems detect mainly the IgM antibody response. In reinfections both CFT and agglutination tests provide predominantly negative results. Nevertheless these test systems currently dominate the ELISA technology<sup>4</sup>. By using ELISA, IgG, IgA, and IgM, antibodies can be differentiated. The crucial factor for a specific and sensitive ELISA is the antigen.<sup>5</sup>

Author α: Research Scholar, Department of Microbiology, Geetanjali Medical College, Udaipur. e-mail: abhineetmehrotra@gmail.com

Author o: Prof. & HoD, Department of Microbiology, Geetanjali Medical College, Udaipur.

Author p: Prof. & HoD, Department of Microbiology Career Institute of Medical Sciences & Research, Lucknow.

Author G2: Associate Professor, Department of Medicine, Career Institute of Medical Sciences & Research, Lucknow.

In the present study we try to evaluate the effective method in the diagnosis of mycoplasma pneumonia among three techniques i.e. Serology, Culture & PCR and an attempt was made to explore the socio demographic and clinical characteristics.

#### II. MATERIAL & METHODS

The present study was undertaken at Career Institute of Medical Science, Lucknow. The total number of study subjects were 286. The total number of male subjects were 115 (59.6 %) while 78 (40.4 %) were females. The present study protocol was approved by institutional ethical committee.

#### a) Culture, Serology and PCR

The expectorated sputum (throat swabs) and blood was collected from all patients. The M. pneumoniae standard strain was revived according to ATCC (American Type Culture Collection) 15531™ guidelines (www.atcc.org) which is commercially available in market. In brief, the lyophilized culture was resuspended in 6 ml pleropnemonia like organism (PPLO) broth. A single drop was used to inoculate PPLO agar; 3 ml suspension was used to prepare glycerol stocks and stored at -70°C. The remaining 3ml suspension was incubated at 37°C and 5 per cent CO2 incubator till growth was observed with change in color from red to yellow. Mycoplasma pneumoniae strains are very slow growing and produce a very light turbidity. Growth in broth is best observed after 10 to 14 days of incubation. Usually it takes at least seven days for the first tubes to start showing growth. Growth is easily recognized by an indicator change from red to orange to yellow. The cells are best transferred when the medium is orange. After medium changes to yellow, cells have started to die. All the patients whose culture was found to be positive were taken for further consideration. The patients having positive culture were marked and further followed for serology. Only serum part which was separated from whole blood of those whose culture was positive. Serum was stored at -20°C till assayed. ELISA (Enzyme-Linked Immunosorbent Assay) kit is used for the accurate qualitative measurement of IgM class antibodies against Mycoplasma pneumoniae in Human serum and plasma. The clinical measurements were recorded. After serological test those patients were again used for follow up (for the purpose of PCR) whose culture as well as serology was found to be positive.as well as all other samples of culture and serology negative were also processed for PCR.

The sputum samples were collected in PBS were centrifuged at 1957 x g for 10 min. Supernatant was decanted and the pellet was resuspended in 0.5 ml PBS and stored at -700C till further processing. The positive control consisted of relevant purified mycoplasma DNA. The extraction of DNA was done

using the organic methods described by Das et al [5]. A 543 bp section of the P1 protein gene of M.pneumoniae was selected for amplification. The primers (Bangalore Genei Pvt. Ltd., Banglore, India) were:

*Primer1:* 5'CAAGCCAAACACGAGCTCCGGCC-3', which is complimentary to the P1 gene negative strand residues 3666-3688, and

*Primer 2 :* 5'CCAGTGTCAGCTGTTTGTCCTTCCCC- 3', which is complimentary to the P1 gene positive strand residues 4208-4183.

Amplification was done according to the guidelines of Taq DNA polymerase (Bangalore Genei Pvt. Ltd., Banglore, India). Amplified PCR products were subjected to electrophoresis on 3 ethidium bromide stained agarose gel, along with a molecular weight marker. A mixture of  $2 \mu$ I genomic DNA,  $2 \mu$ I of 6 X BPB and 8  $\mu$ I of distilled water was loaded into the 1 per cent agarose gel. The electrophoresis was carried out at a constant voltage of 50 V for 1 h, and a band at 543 bp was taken to be a positive result. The PCR of those patients whose culture and serology was positive was carried out.

#### III. Results

Table 1 represents the age wise distribution of study subjects. The total number of study subjects was 286. The total number of male subjects were 207 (72.4 %) while 79 (27.6 %) were females. Highest number of subject's i.e. 32.9 % subjects were less than 30 years while second highest number i.e. 30.4 % of study subjects was more than 45 years. Table 2 Gender wise distribution of study subjects. Table 3 shows the test result by different diagnosis technique of mycoplasma pneumonia. Out of 286 patients 168 (58.7 %) samples were found positive in culture and 98 (34.3%) samples were found to be positive in serology test. PCR testing shows only 80 (28 %) samples were found to be positive.

Table 1: Age and sex distribution of the study population (n = 286)

Age Group (years)	Sample	N (%)
<3	15	5.2%
3-5	21	7.3%
6-10	33	11.5%
11-15	19	6.6 %
16-30	94	32.9 %
31-45	87	30.4%
46-60	9	3.1%
>60	8	28 %

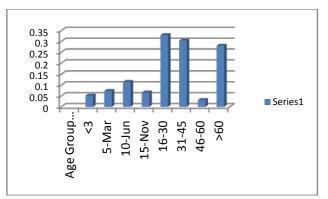


Figure 1

Table 2 : Gender wise distribution

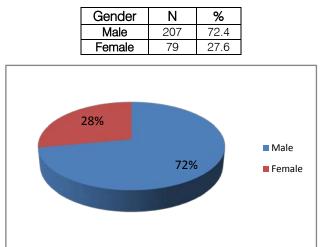


Figure 2

Table 3 : Test results of sample

Test Result	Culture (n=286)	Serology (n=286)	PCR (n=286)
Positive Result	168(58.7%)	98 (34.3%)	80(28%)
Negative result	118(41.3%)	188 (65.7%)	206(72%)

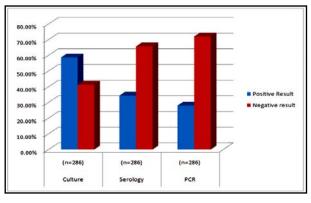


Figure 3

#### a) Study design

This prospective cross-sectional study design will be used for the present study was conducted on community-acquired pneumonia in the Department of Microbiology, at Career Institute of Medical Science, Lucknow.

#### b) Sample size calculation

Sample size was calculated by the sample size calculation formula for sing proportion by  $n=4pq/d^2$ , Where n: sample size, p: prevalence q: 1-p and d is specified absolute precision by assuming prevalence 21 %, with specified absolute precision 0.06 and 10 % data loss, be. Calculated sample size is 260. Adding 10% non-response, the final sample size is 286.

Sample size is 260. Adding 10% non-response, the final sample size is 286.

#### i. Inclusion criteria

Patients clinically diagnosed clinical and radiological proven of atypical pneumonia will be included in the study with,

- 1. Community acquired pneumonia (CAP):
- 2. Presence of a new pulmonary infiltrate/ shadow on chest X-ray suggestive of pneumonia at/ within 24 h of hospitalization.
- 3. Patient residing in community.
- ii. Exclusion criteria
- 1. Hospital acquired pneumonia i.e. pneumonia that developed 72 h after hospitalization or within 7 days of discharge.
- 2. Pulmonary shadow due to a cause other than pneumonia.
- 3. The serious patients having other disease with atypical pneumonia will be excluded

A written informed consent was taken from the parent/legal guardian of the children before them being enrolled in the study.

Clinical data from the patients were collected using a questionnaire developed and validated and a detailed examination was performed. Routine laboratory investigations were done in all subjects. In the present study culture, serology and PCR technique is used to diagnose the mycoplasma pneumonia.

#### IV. DISCUSSION

A variety of techniques are used to assess *M.* pneumoniae, each with different advantages and disadvantages. In this study, PCR, ELISA, and culture methods were employed for the detection of *M.* pneumoniae infection in 286 suspected patients. The culture method for the isolation of *M. pneumoniae* requires 2–4 weeks, which limits itsclinical usefulness. Moreover, culturing of *M. pneumoniae* is expensive and time-consuming (7-10). Serological methods are more extensively used than culture, because they are easier to carry out and more affordable (11-13).However, they are generally nonspecific, retrospective, and need 2 samples for titration<sup>7</sup>. It should be noted that in our study, the most reliable result was obtained by the PCR method, which proved to be highly sensitive, specific,

and faster than other methods. The specificity of the culture method was 100%, while its sensitivity barely reached 33%, relative to the results of the PCR method as a gold standard. All of the culture and IgM ELISA test positive patients were PCR-positive, too. The PCR method is more sensitive and is the gold standard currently being used for diagnosis of this organism in some laboratories<sup>14</sup>

#### V. Conclusion

In the present study it is concluded that only one technique may not be sufficient to diagnose the mycoplasma pneumonia as these techniques do not show higher sensitivity and specificity as many results suggest the same [6]. However, serology and culture is done commonly.

#### VI. Acknowledgment

The authors acknowledge the help provided by various faculties, technicians and paramedical staffs of Career Institute of Medical Sciences, Lucknow, for providing patient samples. Authors would like to acknowledge the Dean/Director of the institute to provide great support.

#### References Références Referencias

- 1. Shin DH, Kim J-S, Yokota H, Kim R, kim S-H. Crystal structure of the DUF16 domain of MPN010 from Mycoplasma pneumoniae. Protein Science : A Publication of the Protein Society. 2006; 15 (4): 921-928. doi:10.1110/ps.051993506.
- Ken B. Waites and Deborah F. Talkington Clin. Microbiol. Rev. October 2004 vol. 17 no. 4 697-7281 October 200 doi: 10.1128/CMR.17.4.697-728.2004
- 3. http://www.medac.de/medac\_international/data/dia gnostics Specific and sensitive diagnosis Mycoplasma pneumoniae IgG-, IgA-, IgM-ELISA(nb)
- Talkington DF, Shott S, Fallon MT, Schwartz SB, Thacker WL. Analysis of Eight Commercial Enzyme Immunoassay Tests for Detection of Antibodies to Mycoplasma pneumoniae in Human Serum. Clinical and Diagnostic Laboratory Immunology. 2004; 11 (5): 862-867. doi: 10. 1128/CDLI.11.5.862-867.2004.
- Bineeta Kashyap, Surinder Kumar, G.R. Sethi\*, B.C.Das\*\* & S.R. Saigal Comparison of PCR, culture & serological tests for the diagnosis of Mycoplasma pneumoniae in community-acquired lower respiratory tract infections in children Indian J Med Res 128, August 2008, pp 134-139
- Zhang L, Zong Z-Y, Liu Y-B, Ye H, Lv X-J. PCR versus serology for diagnosing *Mycoplasma pneumoniae* infection: A systematic review & metaanalysis. *The Indian Journal of Medical Research*. 2011; 134 (3): 270-280.
- 7. Atkinson TP, Balish MF, Waites KB. Epidemiology, clinical manifestations, pathogenesis and laboratory

detection of Mycoplasma pneumoniae infections. FEMS Microbiol Rev 2008; 32: 956–73.

- Ma LD, Chen M, Dong Y, Fan J, Xia L, Wang S et al. Rapid *Mycoplasma* culture for the early diagnosis of *Mycoplasma pneumoniae* infection. J Clin Lab Anal 2010; 24: 224–9.
- Sasaki T, Kenri T, Okazaki N, Iseki M, Yamashita R, Shintani M et al. Epidemiological study of *Mycoplasma pneumoniae* infections in Japan based on PCR-restriction fragment length polymorphism of the P1 cytadhesin gene. J Clin Microbiol 1996; 34: 447–9.
- 10. Waites KB, Talkington DF. *Mycoplasma pneumoniae* and its role as a human pathogen. Clin Microbiol Rev 2004; 17: 697–728.
- 11. Liu FC, Chen PY, Huang FL, Tasai CR, Lwee CY, Wang LC. Rapid diagnosis of *Mycoplasma pneumoniae* infection in children by polymerase chain reaction. J Microbiol Immunol Infect 2007; 40: 507–12.
- 12. Dunn JJ, Malan AK, Evans J, Litwin CM. Rapid detection of *Mycoplasma pneumoniae* IgM antibodies in pediatric patients using ImmunoCard Mycoplasma compared to conventional enzyme immunoassays. Eur J Clin Microbiol Infect Dis 2004; 23: 412–4.
- Daxboeck F, Krause R, Wenissch C. Laboratory diagnosis of *Mycoplasma pneumoniae* infection. Clin Microbiol Infect 2003; 9: 263–73.
- Beersma MF, Dirven K, van Dam AP, Templeton KE, Claas EC, Goossens H. Evaluation of 12 commercial tests and the complement fixation test for *Mycoplasma pneumoniae*-specific immuneglobulin G (IgG) and IgM antibodies, with PCR used as the "gold standard". J Clin Microbiol 2005; 43: 2277–85.

© 2015 Global Journals Inc. (US)