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Results: The signs and symptoms of neuropathic pain showed a reduction ($p < 0.05$) between 4.7 and 5.5 points in DN4 in all groups irradiated with LEDs, with a clinical effect. No change in arterial blood flow of the legs was observed.

Conclusion: Photobiomodulation therapy was effective in reducing the signs and symptoms of neuropathic pain in the lower limbs of type 2 diabetic patients.

Keywords: photobiomodulation, neuropathic pain, diabetic polyneuropathy, painful polyneuropathy, type 2 diabetes mellitus, blood flow.

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I. INTRODUCTION

The causes of the onset of pain in diabetic patients are diverse and include mainly the metabolic deficit of the neuronal cell and reduced vascular supply, currently, these two factors are identified as the main reasons for early hospitalization and responsible for 50% of amputations of lower members globally ¹⁻³.

The repercussion of diabetes mellitus (DM) in face of vascular impairment is centered on the damage at the level of peripheral nerve cells, thus the micro-vascular deficit and the constant increase in free radicals generate impairment in the non-myelinated fibers of the sympathetic system and in the cholinergic-type vasodilator nerve endings, reducing the release of acetylcholine (ACh) and limiting the production of nitric oxide (ON), by means of the enzyme nitric oxide synthase (eONS) ⁴.

Nervous impairment resulting from diabetes mellitus not only affects the sensorimotor nerve fibers but, at an advanced stage, can have repercussions for the peripheral autonomic fibers ⁵, thus the diabetic distal polyneuropathy is the most frequent of the neuropathies, affecting mainly the lower extremities ⁶⁻⁸.

The sensory axons are the most affected because they have a larger mitochondrial population, leaving them in a condition more vulnerable to oxidative stress ^{9,10}. Diabetic polyneuropathy is identified as the main factor for the appearance of clinical signs and symptoms in these patients, since 20% of patients may experience neuropathic pain ¹¹.

The choice of a non-pharmacological therapeutic line, which involves the relief of neuropathic pain in the clinical environment, still deserves further investigation. Clinical and experimental evidence suggests that changes in cell function, resulting in oxidative stress, act as a major factor in the development and progression of pain diabetic neuropathies ¹².

The great effects resulting from photobiomodulation (laser or LED) are attributed to the heme cofactors existing in erythrocytes and

mitochondria. Thus, half of the photons generated by the irradiation are attenuated by the enzyme cytochrome C oxidase of the respiratory chain, although hemoglobin, myoglobin, and melanin may also present a high attenuation coefficient for photons, mainly in the red light spectrum^{13,14}.

Today, in clinical practice, two resources are used for photobiomodulation (Laser and LED), although the laser emits collimated light, different of LED light, both have the same therapeutic effect¹⁵, since this difference ends when the light reaches the biological tissues, due to its dispersion.

Scientific evidence shows that some biological tissues have a high coefficient of therapeutic light attenuation (photobiomodulation), such as the skin (higher melanin concentration)¹⁶, the erythrocyte^{17,18}, nervous tissue¹⁹⁻²¹, and muscle tissue^{22,23}. Thus, when the light is absorbed by photoreceptors, photonic energy is transformed into chemical energy, making photobiomodulation a non-pharmacological alternative that can generate an increase in energy metabolism^{24,25}, vasodilation, increased blood flow, angiogenesis, increased inflammatory response, accelerated healing process^{26,27}, increased oxygen affinity hemoglobin¹⁸, and reduction of neuropathic symptoms through cytokine stimulation and release mechanisms²⁸.

Given the above, the purpose of this study was to show that photobiomodulation with light emitting diode (LED) can improve the signs and symptoms of pain generated by the metabolic disorder of lower limbs in diabetic patients with polyneuropathy (Painful polyneuropathy), from the effects of metabolic increase and favoring biophysical photobiomodulation responses.

II. MATERIALS AND METHODS

a) Ethical aspects

This study is a randomized double-blinded placebo-controlled clinical trial, it was approved by the Ethics Committee on Experimentation with Human Beings of the Clinical Hospital - FMRP/USP (process no. 3.805.967) and registered as a clinical trial on ClinicalTrials.gov. (NCT03369834). Patients who agreed to participate in the study were informed about the objectives and procedures and signed a free and informed consent form. The study was developed from April 2018 to March 2020.

b) Sample and Randomization

The sample calculation was performed using the Ene® software (version 3.0, Barcelona, Spain). The sample size was calculated based on the study by Lorne et al. (2004)²⁹, which evaluated pain in diabetic polyneuropathy using laser as a therapeutic tool. We considered the average values of the Visual Analog Pain Scale (VAS), based on the four evaluation periods. The calculation was based on the detection of comparison

between the groups, with a mean of the reference group of 9 and the experimental group 9. Considering the statistical power of 90% and alpha 0.05, the number of 10 patients per group was estimated. Considering a sample loss of 10%, 12 patients were recruited per group.

Seventy patients with type 2 diabetes mellitus of both sexes with more than 5 years of diagnosis at the age of 45 to 70 years old were recruited at the Hospital das Clínicas de HC/USP. 58 of whom had neuropathic pain ranging from 4 to 6 points (DN4), classified as moderate to severe on the scale of diabetic distal polyneuropathy (DSDDP) in the lower limbs. The exclusion criteria involved neurological lesions that impeded the proposed exams, chronic renal failure, and patients who have had part of the saphenous vein removed for myocardial revascularization. Some of the volunteers took medication to control blood pressure, among them ACE antidiuretics and beta-blockers. All were instructed not to take analgesics or anti-inflammatory drugs during the week of the study.

Randomization was performed using a table of random numbers in the Excel software, which was placed in sealed opaque paper envelopes, opened only in the presence of the patient. Patients were randomly divided into the following groups: Control (C, n = 11), Sham (S, n = 12), Red LED (R, n = 11), Infrared LED (IR, n = 13), Red+Infrared LED (R+IR, n = 11).

Researchers 2 and 3 and the patients themselves were blinded regarding the distribution of groups and the interventions applied. The blinding of the patient in relation to the groups, with the exception of the control, was accomplished with the use of a blindfold. As shown in the flowchart in Figure 1.

Randomization and Allocation

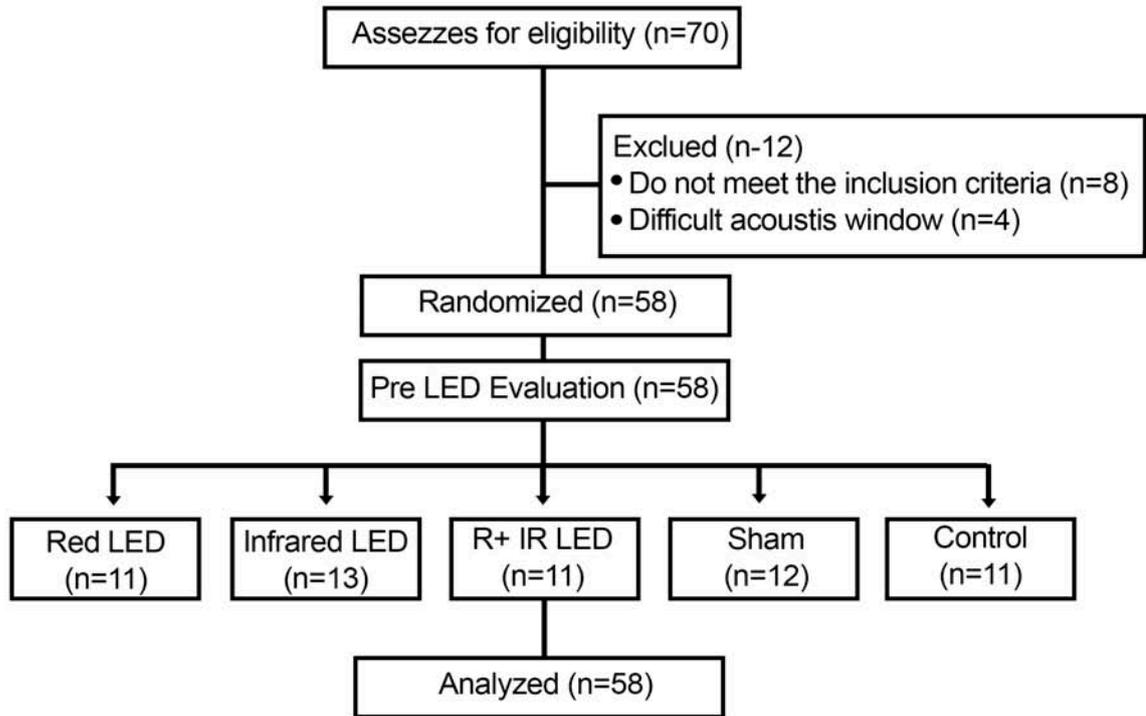


Fig. 1: Flowchart of the study design

c) Procedure

The study procedures were performed in 5 days. On the first day, the researchers collected the anthropometric and clinical data, applied the diagnostic scale for distal diabetic polyneuropathy (DSDDP) and the neuropathic pain questionnaire (DN4), and evaluated the blood flow in the posterior and dorsal tibial arteries through Doppler ultrasound. From the second to the fourth day, the patients received irradiation

(treatment group) and the simulation of application (Sham group) of photobiomodulation with light emitting diode (LED) for one minute and 37 seconds, on average, in each lower limb, totaling 3 minutes per patient, respecting the 24-hour interval between irradiations. On the fifth day, reassessments were carried out using the same assessment tools of the first day (Figure 2).

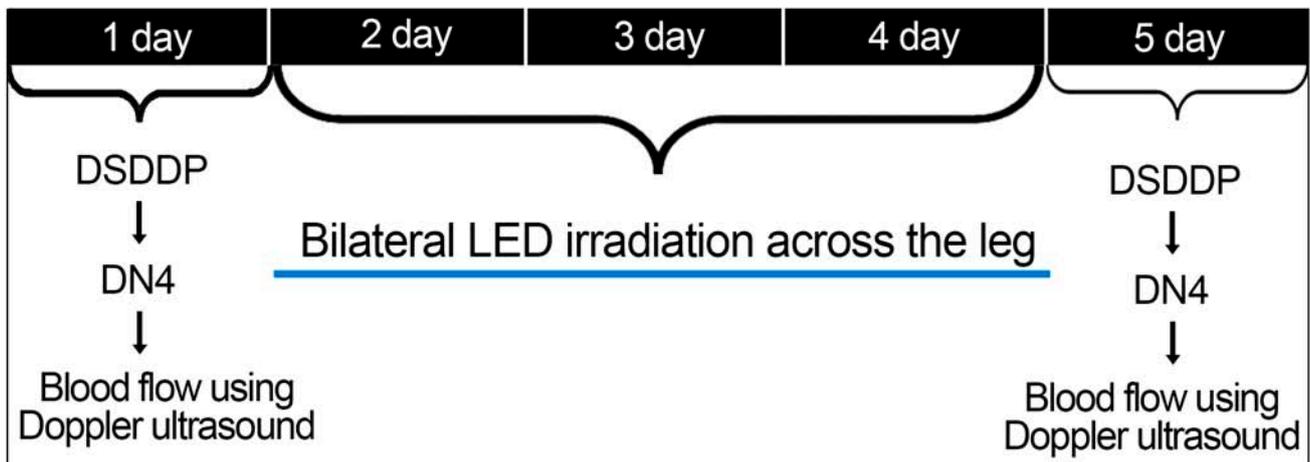


Figure 2: Description of the evaluation, intervention and reassessment during the proposed 5 days.

d) *Assessment of neuropathy*

i. *Diagnostic scale for diabetic distal polyneuropathy*

To quantify the degree of polyneuropathy, the Diagnostic Scale for Diabetic Distal Polyneuropathy (DSDDP) was used, which evaluates neuropathic symptoms with six questions and neuropathic impairment with the Aquileu reflex tests and the hallux vibratory, painful and thermal sensitivity³⁰. Thus, patients should have a score equal to or greater than 5 for symptoms (ESN), associated with a score equal to or greater than 3 for signs (ECN). To characterize neuropathic pain, the patient should have a score equal to or greater than 4 on the Neuropathic Pain Diagnosis questionnaire (DN4).

ii. *Aquileu reflex test*

The Aquileu reflex test was performed using Buck's neurological hammer to assess motor changes in alpha-thick myelinated A-fibers³⁰. With the patient seated, the foot hanging and in a neutral position, percussion was performed with the reflex hammer on the Achilles tendon of both lower limbs. As a positive answer, the volunteer is expected to do reflex plantar flexion, as a consequence of percussion, and the absent or diminished reflex response means an altered result. Therefore, a second percussion was performed to confirm the altered result^{31,32}.

iii. *Vibratory sensitivity test*

For the vibration sensitivity test, a 128 Hertz frequency tuning fork was used to test changes in beta-thick myelinated A-fibers³⁰. With the patient in the supine position, the vibrating tuning fork was applied with the end of the nail, perpendicular and with constant pressure, on the dorsal portion of the distal phalanx of the hallux and medial malleolus or tibial tuberosity, as an alternative, if the volunteer could not feel it in the first region tested. Two applications were made, alternating with a false application, in which the tuning fork was not vibrating. Because of this, the result should be negative, that is, the absence of protective sensitivity, with two incorrect answers from the three applications. The test was performed on both lower limbs^{31,32}. According to Perkins et al.³³, the vibration test has a sensitivity of 53% and specificity of 99% for peripheral neuropathy.

iv. *Pain and thermal sensitivity test*

Buck's neurological hammer needle was used for the painful sensitivity test, to evaluate sensory changes in C-thin unmyelinated fibers³⁰. With the volunteer in the supine position, the tip of the reflex hammer was applied to the back of the hallux, bilaterally, with sufficient pressure to deform the skin. An altered result was the inability to feel the pressure exerted on the hallux^{31,32}. In the assessment of thermal sensitivity, the test was carried out with a test tube containing ice water to assess the sensitive alterations of delta-thin myelinated A-fibers³⁰. With the volunteer in the supine position, a test tube containing ice water was

applied to the dorsal region of the feet of both lower limbs, in which the result was considered altered when the patient did not report the cold sensation at the test site³³.

v. *DN4 Questionnaire for diagnosis of neuropathic pain*

For the qualification of neuropathic pain, the DouleurNeuropathique 4 (DN4) questionnaire was used, it has seven questions and three sensory exams, which are able to discriminate neuropathic pain from nociceptive pain. It was translated to Portuguese and validated, with 100% sensitivity and 93.2% specificity³⁴.

For scoring, DN4 assigns 1 when it is positive and 0 when it is negative (total score range varies from 0 to 10). The cut-off value for the diagnosis of neuropathic pain is a total score equal to or greater than 4

Assessment of arterial blood flow

A portable continuous-wave Doppler device with spectral analysis (Nicolet Vascular Versalab SE, San Carlos, CA, USA) was used, coupled to a notebook. To capture and process the signals, Care Fusion 7.0 software (Nicolet Vascular Versalab SE, San Carlos, CA, USA) was used, which allows the quantification of blood flow, including the peak systolic velocity and resistivity index, as well as its qualification in an interval of time.

Blood flow was assessed after 10 minutes of rest in the supine position. Collections were performed in the posterior and dorsal tibial arteries, with frequencies of 4 and 8 MHz, respectively, in both lower limbs.

Photobiomodulation

Irradiation was performed individually in 58 patients with moderate to severe diabetic distal polyneuropathy. Thus, 116 legs were irradiated during the second, third, and fourth days of therapy. The light was irradiated by a 33x42cm² LED mat, using SMD5050 diodes fed with 12-volt voltage and fixed in an ethylene-vinyl-acetate (EVA) plate with equidistant distribution (1 cm) between the LEDs³⁵. The energy applied to each leg was 180 J, on the anterior and posterior regions, bilaterally. The blanket was malleable to adapt to the contour of the leg, so the application of photobiomodulation was directly on the skin involving the entire leg of the patients (Figure 3).





Figure 3: Application of the LED mat directly on the skin involving the whole leg bilaterally.

All LEDs were previously checked at the Photobiophysics Laboratory of the Faculty of Philosophy, Sciences, and Letters of Ribeirão Preto, University of São Paulo, where the wavelengths, power,

and power density were evaluated. 285 LEDs were used in both blankets (red, infrared), and 320 LEDs for the mixed blanket, 160 LEDs in the red spectrum (620 nm), and 160 in the infrared spectrum (940 nm) (Table 1).

Table 1: Physical parameters of light-emitting diodes (LED) and photobiomodulation therapy protocol.

Variables	Red	Infrared	Red+Infrared
Wave-length	620±10nm	940±10nm	
Number of diodes	285	285	320
Diode diameter	0.125 cm ²	0.178 cm ²	
Power Density	52.86 mW/cm ²	33.7 mW/cm ²	
Diode power	0.0066 W	0.006 W	
Total blanket power	1.88 W	1.71 W	2.01 W
Application time	96 s	106 s	90 s
Total energy per leg	180 J	180J	180 J

mW: milliwatt; s: Second; j: joules; W: watt

e) Statistical analysis

Statistical analysis was performed using SPSS for Windows (version 20; SPSS Inc., Chicago, IL). The mean differences (initial minus final value) of the groups and the 95% confidence intervals (CI) were calculated using mixed linear models, with the baseline as a covariate. The Bonferroni correction test was used to compare the groups, with a p value of 0.05.

To determine the size of the clinical effect of the proposed therapies, Cohen's d was used, with interpretation of the values based on the classification

established by Cohen (1988): less than 0.2, small effect; around 0.5, moderate effect; and above 0.8, great effect.

III. RESULTS

Of the 70 recruited patients, 58 were included and randomized. To characterize the homogeneity of the sample, anthropometric assessments and some clinical routes are available in table 2.

Table 2: Anthropometric and clinical data of patients with type 2 diabetes mellitus, distributed in the experimental groups.

	C n=11	S n=12	R n=11	IR n=13	R+IR n= 11	
Age	63.66 (4.18)	61 (5.37)	61.38 (4.44)	60.08 (7.31)	61.3 (6.99)	
Sex	Male	5	9	5	4	
	Female	3	7	4	8	7
Weight	80.15 (13.78)	80.5 (18.46)	89.36 (21.78)	86.38 (13.42)	97.54 (19.65)	
BMI	30 (4.50)	30.25 (7.72)	32.49 (5.72)	32.39 (4.78)	36.73 (4.97)	
Time of diagnosis (years)	17.23 (6.67)	14.91 (9.17)	13.92 (6.34)	14.08 (6.76)	16.27 (8.79)	
Arterial hypertension	9	10	8	10	9	
DSDDP DM2	Symptoms	6.46±1.89	6.76±2.16	7.8±1.72	6.25±2.22	7.00±1.63
	Signals	4.61±1.12	4.07±0.75	4.54±1.21	4.00±0.73	4.23±0.83
DN4 (pain initial)	6.3 (1.94)	5.6 (1.42)	6 (2.30)	6.9 (3.66)	6.7(1.42)	

BMI: body mass index; DM2: type 2 diabetes mellitus; DSDDP: scale for the diagnosis of diabetic distal polyneuropathy; DN4: Douleur Neuropathique 4 Questionnaire * $p < 0.05$. R: red group, IR: infrared group, R+IR: red+infrared group, S: sham group, C: control group.

As for the assessment of neuropathic pain by the DN4 questionnaire, after irradiation with LED, there was a significant difference in the comparison between the groups that received the treatment (LED) with the control and sham groups. Specifically, when comparing LED red groups with control and sham, the difference ($p < 0.001$) between the means (lower limit and upper limit) was -4.18 (-5.95 -2.40) and -4.08 (-5.81 -2.35), respectively; when comparing infrared LED with control and sham there were also differences ($p < 0.001$) with values of -4.21 (-5.91 -2.50) and -4.11 (-5.77 -2.45)

respectively; as well as LED red+infrared compared to control and sham with differences ($p < 0.001$) of -4.58 (-6.37 -2.80) and -4.49 (-6.23 - 2.74), respectively. In the comparisons between the different groups irradiated with LED, there was no significant difference, as well as between control and sham. The size of the clinical effect of photobiomodulation therapy in the different groups, using Cohen's d, had a great effect on relieving neuropathic pain for all groups irradiated with LED (Table 3).

Table 3: Values of differences between means, 95% confidence interval, upper and lower limits, and assessment of effect size of different groups, considering signs and symptoms of neuropathic pain from the neuropathic Douleur 4 questionnaires (DN4).

Comparison between groups	Difference of means	95% confidence intervals		Effect size Cohen's d
		Inferior limit	Upper limit	
R x IR	0.03	-1.66	1.73	-0.02
R x R+IR	0.40	-1.36	2.18	-0.08
R x C	-4.18*	-5.95	-2.40	-2.08
R x S	-4.08*	-5.81	-2.35	-1.92
IR x R+IR	0.37	-1.33	2.08	0.05
IR x C	-4.21*	-5.91	-2.50	-2.15
IR x S	-4.11*	-5.77	-2.45	-1.61
R+IR x C	-4.58*	-6.37	-2.80	-2.18
R+IR x S	-4.49*	-6.23	-2.74	-1.07
C x S	-0.09	-1.82	1.63	-0.01

* $p < 0.05$. R: red group, IR: infrared group, R+IR: red+infrared group, S: sham group, C: control group.

As for the qualitative assessment of the signs and symptoms of DN4, there was a reduction from 75% to 100% for responses to pain qualification after the application of LED and from 50% to 85% in responses to physical examination of pain. Considering the groups, the table shows that the reduction in pain, with regard to their qualification, occurred in the three groups irradiated in an equitable way, unlike the groups sham and control that did not present any changes. For the

answers to the exams, it is observed that only in red+infrared there was pain reduction (Table 4).

Table 4: Number of patients who present the characteristics/symptoms of pain before and after application of LED, in the Douleur neuropathic 4 questionnaire (DN4).

DN4	R		IR		R+IR		S		C	
	Pre	Post	Pre	Post	Pre	Post	Pre	Post	Pre	Post
Features / Symptoms										
Burning	8	0	8	1	8	2	6	7	7	7
Painful cold sensation	7	0	5	0	6	1	5	5	4	4
Electric shock	8	1	8	1	7	0	5	5	7	7
Tingling	7	1	11	1	8	1	8	8	10	10
Pinned and needled	8	0	11	1	7	1	6	6	6	6
Fall asleep	7	1	10	1	6	0	7	7	8	7
Itching	5	1	8	2	4	0	3	4	3	3
Exams										
Hypoesthesia to the touch	2	3	1	1	7	1	5	5	6	6
Needle prick hypoesthesia	2	3	1	1	8	3	5	5	6	6
Brushing	6	3	6	6	6	3	5	5	6	6

R: red group, IR: infrared group, R+IR: red+infrared group, S: sham group, C: control group.

Considering that neuropathic pain could be associated with circulatory restriction, we opted for the Doppler flow assessment of the lower limb arteries, distally. The results did not show significant changes ($p > 0.050$) for the PVS, IR, IP, and HR parameters for any of the groups, despite presenting a clinical effect size between mild to moderate for the groups irradiated with LED (Table 5).



Table 5: Values of the difference in means and the confidence interval of blood flow variables and do Effect Size of the Doppler ultrasound variables for the tibial and dorsal arteries of the lower limbs.

Varíaveis	R x IR	R x R+IR	R x S	R x C	IR x R+IR	IR x S	IR x C	R+IR x S	R+IR x C	S x C
Dorsal R	PVS 4,21 (-10,07 18,50)	2,56 (-12,40 17,53)	3,36 (-10,94 17,66)	-1,38 (-16,67 13,91)	-1,65 (-15,65 12,34)	-0,85 (-14,12 12,41)	-5,59 (-20,02 8,82)	0,80 (-13,15 14,76)	-3,94 (-18,82 10,94)	-4,74 (-19,00 9,51)
ES	0,34	0,00	0,08	-0,48	-0,32	-0,80	-0,28	0,07	-0,46	0,57
IR	0,09 (-0,03 0,22)	0,03 (-0,10 0,17)	-0,02 (-0,15 0,11)	-0,00 (-0,14 0,13)	-0,05 (-0,18 0,07)	-0,11 (-0,24 0,00)	-0,09 (-0,23 0,03)	-0,06 (-0,19 0,07)	-0,04 (-0,18 0,09)	0,01 (-0,11 0,15)
ES	0,74	0,42	-0,22	0,00	-0,40	-0,98	-0,73	-0,67	-0,40	-0,22
Tibial R	PVS 1,71 (-16,15 19,59)	-1,82 (-20,81 17,17)	-2,66 (20,55 15,22)	-12,89 (-31,34 5,54)	-3,54 (-22,32 15,24)	-4,38 (-22,06 13,30)	-14,61 (-32,82 3,59)	-0,84 (-19,64 17,95)	-11,07 (-30,39 8,24)	-10,23 (-28,46 7,99)
ES	0,11	-0,07	-0,16	-0,78	-0,22	-0,27	-0,89	-0,09	-0,79	0,73
IR	0,05 (-0,06 0,18)	0,00 (-0,13 0,13)	-0,03 (-0,16 0,08)	-0,02 (-0,15 0,10)	-0,05 (-0,19 0,07)	-0,09 (-0,22 0,02)	-0,08 (-0,21 0,04)	-0,04 (-0,17 0,09)	-0,02 (-0,16 0,10)	0,01 (-0,11 0,14)
ES	0,59	0,00	-0,46	-0,29	-0,61	-1,05	-0,82	-0,50	-0,31	-0,11
Dorsal L	PVS -0,40 (-17,88 17,08)	-8,11 (-26,71 10,49)	-15,77 (-33,26 1,71)	-9,59 (-27,87 8,68)	-7,70 (-25,94 10,52)	-15,37 (-32,48 1,73)	-9,18 (-27,10 8,72)	-7,66 (-25,91 10,59)	-1,47 (-20,47 17,51)	6,18 (-11,73 24,10)
ES	-0,03	-0,54	-1,02	-0,54	-0,63	-1,17	-0,63	-1,17	-0,09	-0,39
IR	-0,01 (-0,15 0,12)	-0,02 (-0,16 0,12)	-0,08 (-0,21 0,05)	-0,04 (-0,19 0,09)	-0,00 (-0,14 0,13)	-0,06 (-0,19 0,06)	-0,03 (-0,17 0,10)	-0,05 (-0,19 0,08)	-0,02 (-0,17 0,12)	0,03 (-0,10 0,17)
ES	-0,08	-0,15	-0,62	-0,34	-0,09	-0,70	-0,34	-0,53	-0,20	-0,43
Tibial L	PVS 4,39 (-15,34 24,12)	4,79 (-17,58 27,18)	-1,68 (-22,12 18,76)	-10,81 (-31,83 10,19)	0,40 (-21,25 22,06)	-6,07 (-25,71 13,56)	-15,20 (-35,45 5,03)	-6,47 (-28,76 15,80)	-15,61 (-38,44 7,21)	-9,13 (-30,08 11,81)
ES	0,29	0,38	-0,18	-0,63	0,04	-0,83	-0,42	-0,48	-0,89	0,42
IR	0,04 (-0,11 0,20)	0,04 (-0,13 0,22)	-0,02 (-0,19 0,13)	-0,04 (-0,20 0,12)	0,00 (-0,17 0,17)	-0,08 (-0,23 0,07)	-0,08 (-0,25 0,07)	-0,08 (-0,26 0,09)	-0,08 (-0,27 0,09)	0,00 (-0,16 0,17)
ES	0,30	0,34	-0,35	-0,45	0,00	-0,69	-0,82	-0,65	-0,76	0,09

Dorsal (R, L): right and left dorsal artery, Tibial (R, L): right and left posterior tibial artery, PVS: peak systolic velocity, IR: resistivity index, R: red group, IR: infrared group, R+IR: red+infrared group, S: sham group, C: control group, ES: Effect Size

IV. DISCUSSION

The hypothesis that photobiomodulation with LED could be effective in reducing the signs and symptoms of neuropathic pain of the lower limbs was confirmed, being an effective therapy in short-term intervention, with a great clinical effect for all groups irradiated with LED.

According to Kallenborn-Gerhardt et al. (2013)³⁶, the main causes of pain in peripheral neuropathy still need further studies. In diabetic patients, the emergence of neuropathy is believed to be due to multiple factors, including the increased production of reactive oxygen species at the mitochondrial level, reduced antioxidant capacities such as superoxide dismutase catalase and glutathione in enzymatic and non-enzymatic cells³⁷, and vascular impairment generated by chronic hyperglycemia^{38,39}. These factors are identified as the main causes of reverse neuroinflammation, which promotes an imbalance in the production of ATP and the induction of apoptosis in nerve cells. Thus, the results of this study open a window so that photobiomodulation can be used as a non-pharmacological alternative, contributing to the reduction of pain generated by painful neuropathy.

In the light of the evidence, the results of Cg et al. (2015)⁴⁰ corroborated the results found in this study, based on the hypothesis that the biophysical responses generated by photobiomodulation may control neurological pain in diabetic patients with polyneuropathy. Thus, it is highlighted that the three LED irradiations in the leg were sufficient to reduce pain in the lower limbs. Gobbi et al. (2020)⁴¹ report that PBMT applied for a short period does not bring important gains for the muscular performance and functionality of diabetic individuals.

As it is a subjective experience, pain has some limitations regarding its measurement. The use of a standardized and validated questionnaire that involves other aspects than just its intensity is important since it contemplates the subjective experience, involving the affective responses associated with pain. Thus, the application of DN4 sought to expand this analysis.

The quantification of pain through DN4 showed a reduction between 4.7 and 5.5 points, on a 10 point scale, in the LED groups, so it is believed that this response is due to the effects of metabolic increment and favoring biophysical responses generated by photobiomodulation, as proposed by Janzadeh et al. (2016)⁴² in an animal model, where they emphasized that photobiomodulation can generate an increase in the levels of antioxidants, such as superoxide dismutase catalase and glutathione, improve mitochondrial function, safeguard the survival of neural cells, and improve symptoms of pain.

It speculates that the affective responses associated with pain reduction in the red+infrared LED group are caused by the proportional increase in the metabolic effect generated by the combined irradiation of the two light spectra. Since the two lengths tend to improve electrochemical activity and increase ATP re-synthesis¹⁴. In addition, irradiation with LED in the infrared spectrum promotes an increase in the concentration of oxyhemoglobin and total oxymyoglobin, which increases the availability of oxygen¹⁸ that may favor aerobic metabolism.

Although the main factor for the emergence of neuropathic pain is assigned to metabolic stresses (ROS)⁴³, it is also emphasized that the vascular impairment, generated by reduced flow and hypoperfusion due to hyperglycemia, can lead to the failure of several tissues and contribute to the onset of neuropathic pain early⁴⁴.

Although there were no significant changes in blood flow, the clinical effect was also in favor of the irradiated groups.

Considering that oxidative stress plays a central role in the development of microvascular complications of diabetes⁴⁵, as well as a secondary variable, the blood flow of the posterior and dorsal tibial arteries was measured in order to support the context of blood circulation in the appearance of neuropathic pain. Although there was no significant improvement in the quantitative assessments of Doppler ultrasound, it is believed that photobiomodulation tends to improve blood flow and reduce arterial resistance, since there was an improvement from mild to moderate in the effect size in the irradiated groups, for the peak systolic velocity variables and the resistivity index in the three therapeutic intervention groups. The mechanisms proposed to explain the vascular changes induced by glucose and lipids in diabetes include the accelerated formation of advanced glycation end products (AGEs), activation of protein C kinase, inflammatory signaling, and oxidative stress⁴⁶. Accordingly, the deficits in the bioavailability of nitric oxide and the large concentrations of ROS are the main responsible for vasculopathy in diabetic patients,^{47,48} responses that can be reversed with PBM.

According to our searches, in order to confront the findings of the influence of LED light on blood flow, there was an absence of studies that analyzed vascular resistance (IR) and improved blood flow (PVS) in patients with pain secondary neuropathic diabetes mellitus, what leads to a limitation in the discussion of the results presented.

In view of the results obtained, we believe that the irradiation time of three days is one of the factors that can be extended, allowing a longer response time of the tissues in face of photobiomodulation, allowing an effect that can generate changes that can be reflected

at the level of macrocirculation. In addition, using different means of evaluation, such as microcirculation, tissue perfusion of the lower limbs, biomarkers of neuronal dysfunction, and systemic inflammation, could complement the analysis of neuropathic pain and vascular condition, secondary to diabetes mellitus.

Thus, despite the positive limitations faced due to lack of equipment to better assess the painful polyneuropathy caused by type 2 diabetes mellitus, this study opens a new window for photobiomodulation by light-emitting diode (LED) in the red, infrared, or associated spectrum is now filled as a non-pharmacological therapeutic line for reducing the signs and symptoms of neuropathic pain in patients with type 2 diabetes with polyneuropathy of the lower limbs.

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Conflict of Interest

The authors declare no conflicts of interest.

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Ethics approval

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REFERENCES RÉFÉRENCES REFERENCIAS

1. Ulbrecht JS, Cavanagh PR, Caputo GM. Foot problems in diabetes: an overview. *Clinical infectious diseases: an official publication of the Infectious Diseases Society of America*. Aug 1 2004; 39 Suppl2: S73-82.
2. Boulton AJ, Vileikyte L, Ragnarson-Tennvall G, Apelqvist J. The global burden of diabetic foot disease. *The Lancet*. 2005; 366(9498): 1719-1724.
3. Schreml S, Berneburg M. The global burden of diabetic wounds. *The British journal of dermatology*. Apr 2017; 176(4): 845-846.
4. Bello A, Biliaminu S, Wahab K, Sanya E. Distal symmetrical polyneuropathy and cardiovascular autonomic neuropathy among diabetic patients in Ilorin: Prevalence and predictors. *The Nigerian postgraduate medical journal*. Apr-Jun 2019; 26(2): 123-128.
5. Chan AC, Wilder-Smith EP. Small fiber neuropathy: Getting bigger! *Muscle & nerve*. May 2016; 53(5): 671-682.
6. Gwathmey KG, Pearson KT. Diagnosis and management of sensory polyneuropathy. *Bmj*. May 8 2019; 365: l1108.

7. Callaghan BC, Cheng HT, Stables CL, Smith AL, Feldman EL. Diabetic neuropathy: clinical manifestations and current treatments. *The lancet NEUROLOGY*. 2012; 11(6): 521-534.
8. Leahy JL. Pathogenesis of type 2 diabetes mellitus. *Archives of medical research*. May-Jun 2005; 36(3): 197-209.
9. Lotfy M, Adeghate J, Kalasz H, Singh J, Adeghate E. Chronic Complications of Diabetes Mellitus: A Mini Review. *Current diabetes reviews*. 2017; 13(1): 3-10.
10. Marfella R, Nappo F, De Angelis L, Paolisso G, Tagliamonte MR, Giugliano D. Hemodynamic effects of acute hyperglycemia in type 2 diabetic patients. *Diabetes care*. May 2000; 23(5): 658-663.
11. Avetisov SE, Chernenkova NA, Surnina ZV. [Clinical features and diagnosis of diabetic polyneuropathy]. *Vestnik oftalmologii*. 2017; 133(5): 98-102.
12. Babizhayev MA, Stokov IA, Nosikov VV, et al. The Role of Oxidative Stress in Diabetic Neuropathy: Generation of Free Radical Species in the Glycation Reaction and Gene Polymorphisms Encoding Antioxidant Enzymes to Genetic Susceptibility to Diabetic Neuropathy in Population of Type I Diabetic Patients. *Cell biochemistry and biophysics*. Apr 2015; 71(3): 1425-1443.
13. Pastore D, Greco M, Passarella S. Specific helium-neon laser sensitivity of the purified cytochrome c oxidase. *International journal of radiation biology*. Jun 2000; 76(6): 863-870.
14. Hamblin MR. Mechanisms and Mitochondrial Redox Signaling in Photobiomodulation. *Photochemistry and photobiology*. Mar 2018; 94(2): 199-212.
15. Heiskanen V, Hamblin MR. Photobiomodulation: lasers vs. light emitting diodes? *Photochemical & photobiological sciences: Official journal of the European Photochemistry Association and the European Society for Photobiology*. Aug 8 2018; 17(8): 1003-1017.
16. Brondon P, Stadler I, Lanzafame RJ. Melanin density affects photobiomodulation outcomes in cell culture. *Photomedicine and laser surgery*. Jun 2007; 25(3): 144-149.
17. Wajih N, Basu S, Ucer KB, et al. Erythrocytic bioactivation of nitrite and its potentiation by far-red light. *Redox biology*. Jan 2019; 20: 442-450.
18. Linares SN, Beltrame T, Ferraresi C, Galdino GAM, Catai AM. Photobiomodulation effect on local hemoglobin concentration assessed by near-infrared spectroscopy in humans. *Lasers in medical science*. Apr 2020; 35(3): 641-649.
19. Rosso MPO, Buchaim DV, Kawano N, Furlanette G, Pomini KT, Buchaim RL. Photobiomodulation Therapy (PBMT) in Peripheral Nerve Regeneration: A Systematic Review. *Bioengineering*. Jun 9 2018; 5(2).

20. Shen CC, Yang YC, Huang TB, Chan SC, Liu BS. Low-Level Laser-Accelerated Peripheral Nerve Regeneration within a Reinforced Nerve Conduit across a Large Gap of the Transected Sciatic Nerve in Rats. *Evidence-based complementary and alternative medicine: eCAM*. 2013; 2013: 175629.
21. Hennessy M, Hamblin MR. Photobiomodulation and the brain: a new paradigm. *Journal of optics*. Jan 2017; 19(1): 013003.
22. Dos Santos SA, Serra AJ, Stancker TG, et al. Effects of Photobiomodulation Therapy on Oxidative Stress in Muscle Injury Animal Models: A Systematic Review. *Oxidative medicine and cellular longevity*. 2017; 2017: 5273403.
23. Ferraresi C, Bertucci D, Schiavinato J, et al. Effects of Light-Emitting Diode Therapy on Muscle Hypertrophy, Gene Expression, Performance, Damage, and Delayed-Onset Muscle Soreness: Case-control Study with a Pair of Identical Twins. *American journal of physical medicine & rehabilitation*. Oct 2016; 95(10): 746-757.
24. Miranda EF, de Oliveira LV, Antonialli FC, Vanin AA, de Carvalho Pde T, Leal-Junior EC. Phototherapy with combination of super-pulsed laser and light-emitting diodes is beneficial in improvement of muscular performance (strength and muscular endurance), dyspnea, and fatigue sensation in patients with chronic obstructive pulmonary disease. *Lasers in medical science*. Jan 2015; 30(1): 437-443.
25. Nampo FK, Cavalheri V, Dos Santos Soares F, de Paula Ramos S, Camargo EA. Low-level phototherapy to improve exercise capacity and muscle performance: a systematic review and meta-analysis. *Lasers in medical science*. Dec 2016; 31(9): 1957-1970.
26. Desmet KD, Paz DA, Corry JJ, et al. Clinical and experimental applications of NIR-LED photobiomodulation. *Photomedicine and laser surgery*. Apr 2006; 24(2):121-128.
27. Eells JT, Wong-Riley MT, VerHoeve J, et al. Mitochondrial signal transduction in accelerated wound and retinal healing by near-infrared light therapy. *Mitochondrion*. Sep 2004; 4(5-6): 559-567.
28. Wong-Riley MT, Liang HL, Eells JT, et al. Photobiomodulation directly benefits primary neurons functionally inactivated by toxins: role of cytochrome c oxidase. *The Journal of biological chemistry*. Feb 11 2005; 280(6): 4761-4771.
29. Zinman LH, Ngo M, Ng ET, Nwe KT, Gogov S, Bril V. Low-intensity laser therapy for painful symptoms of diabetic sensorimotor polyneuropathy: a controlled trial. *Diabetes care*. Apr 2004; 27(4): 921-924.
30. Moreira RO, Castro AP, Papelbaum M, et al. Tradução para o português e avaliação da confiabilidade de uma escala para diagnóstico da polineuropatia distal diabética. *Arquivos Brasileiros de Endocrinologia & Metabologia*. 2005; 49: 944-950.
31. Milech A, Angelucci AP, Golbert A, Matheus A, Carrilho AJF, Ramalho A. Diretrizes da sociedade brasileira de diabetes (2015-2016). São Paulo: AC Farmacêutica. 2016:13.
32. Boulton AJ, Armstrong DG, Albert SF, et al. Comprehensive foot examination and risk assessment. *Endocrine practice: official journal of the American College of Endocrinology and the American Association of Clinical Endocrinologists*. Jul-Aug 2008; 14(5): 576-583.
33. Perkins BA, Olaleye D, Zinman B, Bril V. Simple screening tests for peripheral neuropathy in the diabetes clinic. *Diabetes care*. Feb 2001; 24(2): 250-256.
34. Van Seventer R, Vos C, Meerding W, et al. Linguistic validation of the DN4 for use in international studies. *European journal of pain*. Jan 2010; 14(1): 58-63.
35. de Jesus Guirro RR, de Carvalho G, Gobbi A, de Oliveira Assuncao FF, de Souza Borges NC, Bachmann L. Measurement of Physical Parameters and Development of a Light Emitting Diodes Device for Therapeutic Use. *Journal of medical systems*. Mar 12 2020; 44(4): 88.
36. Kallenborn-Gerhardt W, Schroder K, Geisslinger G, Schmidtko A. NOXious signaling in pain processing. *Pharmacology & therapeutics*. Mar 2013; 137(3): 309-317.
37. Dongre UJ, Meshram VG, Pitale S. Oxidative Stress and 3243 A/G Mitochondrial Dna Mutation In Maternally Inherited Type 2 Diabetes Mellitus.
38. Sandireddy R, Yerra VG, Areti A, Komirishetty P, Kumar A. Neuroinflammation and oxidative stress in diabetic neuropathy: futuristic strategies based on these targets. *International journal of endocrinology*. 2014; 2014: 674987.
39. Said G. Diabetic neuropathy--a review. *Nature clinical practice. Neurology*. Jun 2007; 3(6): 331-340.
40. Cg SK, Maiya AG, Hande HM, Vidyasagar S, Rao K, Rajagopal KV. Efficacy of low level laser therapy on painful diabetic peripheral neuropathy. *Laser therapy*. Oct 2 2015; 24(3): 195-200.
41. Gobbi A, de Carvalho G, Sapalo AT, de Jesus Guirro RR. Acute application of photobiomodulation does not bring important gains for the muscular performance and functionality of diabetic individuals. *Lasers in medical science*. Jul 2021; 36(5): 995-1002.
42. Janzadeh A, Nasirinezhad F, Masoumipoor M, Jameie SB, Hayat P. Photobiomodulation therapy reduces apoptotic factors and increases glutathione levels in a neuropathic pain model. *Lasers in medical science*. Dec 2016; 31(9): 1863-1869.
43. Volpe CMO, Villar-Delfino PH, Dos Anjos PMF, Nogueira-Machado JA. Cellular death, reactive

oxygen species (ROS) and diabetic complications.
Cell death & disease. Jan 25 2018; 9(2): 119.

44. Kim SE, Park KM, Park J, et al. Vascular factors and neuropathy in lower limb of diabetic patients. *Journal of clinical neuroscience: official journal of the Neurosurgical Society of Australasia*. Jan 2019; 59: 130-135.
45. Domingueti CP, Dusse LM, Carvalho M, de Sousa LP, Gomes KB, Fernandes AP. Diabetes mellitus: The linkage between oxidative stress, inflammation, hypercoagulability and vascular complications. *Journal of diabetes and its complications*. May-Jun 2016; 30(4): 738-745.
46. Potenza MA, Gagliardi S, Nacci C, Carratu MR, Montagnani M. Endothelial dysfunction in diabetes: from mechanisms to therapeutic targets. *Current medicinal chemistry*. 2009; 16(1): 94-112.
47. Giacco F, Brownlee M. Oxidative stress and diabetic complications. *Circulation research*. Oct 29 2010; 107(9): 1058-1070.
48. Hink U, Li H, Mollnau H, et al. Mechanisms underlying endothelial dysfunction in diabetes mellitus. *Circulation research*. Feb 2 2001; 88(2): E14-22.

