

GLOBAL JOURNAL OF MEDICAL RESEARCH: G VETERINARY SCIENCE AND VETERINARY MEDICINE Volume 25 Issue 1 Version 1.0 Year 2025 Type: Double Blind Peer Reviewed International Research Journal Publisher: Global Journals Online ISSN: 2249-4618 & Print ISSN: 0975-5888

Nutritional values and varietal characteristics of two *Moringa oleifera* Lam. Morphotypes from Chad

By Barnabas Kayalto, Christophe Djekota & Abdelsalam Tidjani

University of N'Djamena

Abstract- This study aims to compare the nutritional value of leaf powder from two morphotypes of *Moringa oleifera* Lam. from Chad. In 2024, at Kolobo in the Mayo Kebbi Province in southern Chad, fresh leaf samples were collected from a large-leaf morphotype 1 of *Moringa oleifera* Lam.; the small-leaf morphotype 2 having been analyzed at CRSBAN in a previous study. The freshly harvested leaves were transported by us, sorted, washed and dried in the shade. The powders prepared from the dried leaves were analyzed using standard methods. Physico-chemical analyses revealed that leaf powder from the small-leaf morphotype 2 is rich in protein (24.28 \pm 0.22 g/100g), certain minerals (mg/100g) such as: Ca 1443.90 \pm 11.03; Mg 176.72 \pm 0.73; Fe 53.75 \pm 5.07; Zn 17.58 \pm 0.89 and vitamins (β -carotene 624.40 \pm 0.41 μ g ER, Vitamin C 65.88 \pm 0.00 mg/100g).

Keywords: Moringa, comparative study, nutritional value, morphotypes, chad.

GJMR-G Classification: NLMC: WH 155, WQ 150

NUTRITIONA LVA LUESAN DVARIETA LCHARACTERISTICSOFT WOMDRINGA OLEIFERA LAMMORPHOTY PESFROMCHAD

Strictly as per the compliance and regulations of:



© 2025. Barnabas Kayalto, Christophe Djekota & Abdelsalam Tidjani. This research/review article is distributed under the terms of the Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0). You must give appropriate credit to authors and reference this article if parts of the article are reproduced in any manner. Applicable licensing terms are at https://creativecommons.org/licenses/by-nc-nd/4.0/.

Nutritional values and varietal characteristics of two *Moringa oleifera* Lam. Morphotypes from Chad

Barnabas Kayalto ^a, Christophe Djekota ^a & Abdelsalam Tidjani ^e

Abstract- This study aims to compare the nutritional value of leaf powder from two morphotypes of Moringa oleifera Lam. from Chad. In 2024, at Kolobo in the Mayo Kebbi Province in southern Chad, fresh leaf samples were collected from a large-leaf morphotype 1 of Moringa oleifera Lam.; the smallleaf morphotype 2 having been analyzed at CRSBAN in a previous study. The freshly harvested leaves were transported by us, sorted, washed and dried in the shade. The powders prepared from the dried leaves were analyzed using standard methods. Physico-chemical analyses revealed that leaf powder from the small-leaf morphotype 2 is rich in protein $(24.28 \pm 0.22 \text{ g/100g})$, certain minerals (mg/100g) such as: Ca 1443.90 ± 11.03; Mg 176.72 ± 0.73; Fe 53.75 ± 5.07; Zn 17.58 \pm 0.89 and vitamins (β -carotene 624.40 \pm 0.41 μ g ER, Vitamin C 65.88 \pm 0.00 mg/100g). The leaf powder has a low lipids and total sugars content, respectively (7.42 \pm 1.56 g/100g) and (22.46 ± 2.02 g/100g). Dried Moringa oleifera broadleaf powder has a low protein content (6.09 ± 0.15 g/100g). Its lipid content is even lower (0.52 \pm 0.02 g/100g). However, this morphotype 1 has an ash content (9.94 \pm 0.05 g/100g) similar to the Moringa morphotype 2 with small leaves, which suggests a high mineral content.

These results show that the small-leaf morphotype 2 produced a significantly higher protein, certain minerals and β -carotene content than the large-leaf morphotype 1. This study is a contribution to the nutrition program for children and people with HIV.

Keywords: Moringa, comparative study, nutritional value, morphotypes, chad.

I. INTRODUCTION

oleifera belongs oringa Lam. to the family monogeneric shrubs and trees Moringaceae, considered to have originated in Agra and Oudh, in the north-western region of India and south of the Himalayan mountains (Mallenakuppe et al., 2019). It is now grown throughout the Middle East, almost all of the tropical belt and was introduced to East Africa from India in the early 20th century. Around 33 species have been recorded in the Moringaceae family. Of these, thirteen species namely, M. arborea, M. borziana, M. concanensis, M. drouhardi, M. hildebrandtii,

M. longituba, *M.* oleifera, *M.* ovalifolia, *M.* peregrina, *M.* pygmaea, *M.* rivae, *M.* ruspoliana, *M.* stenopetala are well known and occur worldwide (Mallenakuppe et al., 2019).

Melom *et al.*, (2016) studied the morphological characteristics of different local varieties of *Moringa oleifera* in the Logone valley (Chad-Cameroon). These authors showed that the most decisive parameters, i.e. leaf length and width, capsule length and number of seeds per capsule, using Hierarchical Ascending Classification (CAH) enabled them to identify three morphotypes of *M. oleifera* individuals in the area. Morphotype 1, morphotypes with large leaves, the most represented with 43% of individuals; morphotype 2, morphotypes with small leaves, moderately represented with 33% of individuals and morphotype 3, the least represented, with 24% of individuals. The general characteristics of the three morphotypes are described below.

Morphotype 1:

The bipinnate and tripinnate compound leaves are imparipinnate quadripinnate, 38.95 ± 1.48 cm long and 22.64 ± 0.86 cm wide. The petioles are cylindrical, pubescent, with a thickened base and a small linear depression on the ventral part. The pale green rachis, also cylindrical, are less pubescent. The leaflets have a wedge-shaped, obtuse base and a mucronate apex; they are oval, green ventrally and greenish dorsally. The larger terminal leaflet is short-stalked, more or less glabrous, oval to rounded. The pinnate veins are barely visible (Melom *et al.*, 2016).

The panicle-shaped flowers are tubular, whitish and 3.54 ± 0.07 cm long. The five sepals, 1.46 ± 0.03 cm long, are obovate and tinged with red behind their apex. The five petals are also obovate, with inwardcurving edges and a mucronate apex. They are also tinged red at the base, with red and/or yellow spots. The androecium contains yellow anthers, while the filaments are yellowish and less pubescent. The five stamens, 1.07 ± 0.024 cm long, are fused at their more pubescent base. The pistil is green and pubescent. In the flower buds, the stamens are often separated by staminodes.

The linear, angular, 3- or 4-sided capsules are on average 31.75 \pm 1.20 cm long with 2 grooves on

Author α : Department of Life and Earth Sciences, Bongor Teachers' Training Higher School, Bongor, Chad. e-mail: bkayalto68@gmail.com Author σ : National Institute of Agronomic Sciences and Food Processing Technologies, Lai, Chad.

Author p: Food Science and Nutrition Research Laboratory, faculty of Human Health Sciences, University of N'Djamena, Chad.

each side; they are more or less beige to greyish when ripe. They are large fruits, appearing round; they are straight, but sometimes curved entirely, either at the base or at the beaks. The beaks are straight or curved, with pointed or sharp tips, while others are thin, pointed or sharp. At maturity, the capsules contain an average of 11.00 ± 2.83 seeds. The seeds are spherical, round, with a brown and/or pale-brown, semi-permeable shell. The shell has three white wings, which extend from the base to the top (Melom et al., 2016).

Morphotype 2:

The leaves are imparipinnate, of the same type as group 1, but 34.82 \pm 1.48 cm long and 20.05 \pm 0.86 cm wide. The green and sometimes reddish petioles are more pubescent than the pale green rachis; they also have a short linear depression on the ventral surface. The leaflets have a wedge-shaped base with an obtuse, wedge-shaped, mucronate apex; they are oval, green ventrally and greenish dorsally.

The flowers, 3.29 ± 0.07 cm long, are white, cruciform or tubular and pubescent. The sepals, which are moderately long $(1.31 \pm 0.11 \text{ cm})$, are also obovate, while the five petals are oblong. The stamens are shorter than those above $(1.01 \pm 0.02 \text{ cm})$.

The fruits are slender, slightly longer (33,11 ± 4.59) than those of group 1 and generally have 3 sides. The capsules are straight, but sometimes entirely curved, either at the base or at the beak. Some beaks are rounded, mucronate or simply acuminate. The capsules contain an average of 11.00 ± 2.94 brown and/or pale brown to black seeds (Melom et al., 2016).

Morphotype 3:

Individuals in this group have shorter leaves $(31.48 \pm 1.48 \text{ cm long})$, with greyish to reddish, pubescent petioles. They are cylindrical, thickened at the base with a short linear depression on the ventral surface.

The green rachises have oval leaflets with an obtuse base and mucronate apex, with a green upper surface and greenish lower surface.

The medium-length flowers $(3.45 \pm 0.07 \text{ cm})$ are white with white sepals that are longer (1.40 \pm 0.03 cm) than those of group 2. The petals, spotted red at the base, are obovate. The stamens are moderately long (1.05 \pm 0.02 cm) compared to the first two groups.

The capsules are relatively short (26.87 \pm 1.20 cm) and have 3 or 4 sides. They are straight, but sometimes entirely curved, either at the base or at the beak. The beaks are straight and/or curved, with rounded mucronate or pointed tips. Mature fruits contain an average of 10.80 \pm 4.54 seeds of the same color as those in group 2 (Melomet al., 2016). In the context of this study, leaf morphology could guide the choice of morphotypes depending on the intended use. Morphotypes 2 and 1 are best suited to green fruit production in the Logone valley. However, given the

flocculent capacity of Moringa oleifera fruit fines, the study should be completed by yield tests in order to propose the morphotype best suited to maximum powder production (Foidl, Harinder & Becker, 2001).

Moringa is known as the "tree of life" or the "tree of heaven" because of its exceptional environmental, medicinal and dietary virtues. Its leaves, flowers, fruit, bark and roots can be eaten directly. Its recognized nutritional qualities could well represent an effective solution in the fight against malnutrition (Solidarités International, 2019).

All parts of the Moringa oleifera plant (leaves, roots, flowers, pods and seeds) are edible and contain large quantities of various micronutrients, such as calcium, potassium, zinc, magnesium, iron, copper, vitamins (A, B, C, E) and phytochemicals such as tannins, sterols, terpenoids, flavonoids, saponins, anthraquinones, alkaloids and reducing sugars (Trigo et al., 2021).

Moringa oil also contains around 76% linolenic acid and oleic acid, making it a potential substitute for olive oil. Moringa leaves contain an exceptionally high amount of protein compared with other leaves consumed as food, and essential amino acids such as lysine, tryptophan, phenylalanine, valine, etc. (Trigo et al., 2021).

In developing countries, the desire to ensure food security requires intensive, input-intensive farming, which makes produce inaccessible to the majority of farmers (Ngamo, 2004). As a result, farmers continue to face unstable conditions of food insecurity. This is why new techniques that solve both the problem of food insecurity and the conservation of biodiversity have been developed in recent years, in particular agroforestry (Mapongmetsem and Zedong, 1997; De Jong, Campbell & Schröder, 2000). Agroforestry advocates the use and promotion of non-timber forest products, which can improve the living conditions of rural populations (Mapongmetsem et al., 2010). Moringa oleifera, a Middle Eastern species introduced to East Africa at the beginning of the 20th century as an ornamental and protective plant (Foidl, Harinder and Backer, 2001), is one of these resources of great interest to the rural population. Moringa is a plant which requires less water and nutrients, with rapid growth and development; it is therefore well suited to tropical Sahelian countries.

Making the most of local plant resources that are rich in protein and micronutrients and accessible at low cost is an effective strategy for fighting nutritional deficiencies (Ndong et al., 2007; Anwar et al., 2007). It makes it possible to reduce micronutrient deficiencies and improve the nutritional and health status of malnourished children. A number of studies (Ndong et al., 2007; Saint Sauveur et Broin, 2010; Compaoré et al., 2011; Zongo et al., 2013; Kayalto et al., 2013) in many countries have highlighted the exceptional nutritional qualities of the leaves of *M. oleifera* Lam. originally from India, which are used in powder form in porridges in Asia and Africa.

This study aims to compare the nutritional value of dried leaf powder from two morphotypes of *Moringa oleifera* Lam. from Chad.

II. MATERIALS AND METHODS

a) Location of the Study Area

Fresh *Moringa oleifera* broadleaf leaves, Morphotype 1, were harvested in October 2024 in the village of Kolobo, 60 km south of Bongor, Koyom subprefecture, Mayo-Boneye prefecture, Mayo-Kebbi East province.

b) Sampling

In October 2024, in Kolobo, in the Province of Mayo Kebbi East in southern Chad, samples of fresh

leaves were collected directly from a broad-leaved morphotype of Moringa oleifera Lam. These fresh leaves were transported to Bongor by us, sorted, washed and dried in the shade. The powder prepared from the dried leaves was carefully packed in clean plastic bags and sent to N'Djamena, to Foodstuffs Quality Control Center (CECOQDA) for physico-chemicals analysis using standard methods. It should be noted that Moringa broadleaf, Morphotype 1 was introduced at the Koyom missionary station, 8 km south of Kolobo by evangelist missionaries. It was originally a village pharmacy set up by the work of American doctor David Seymour in the 1950s, followed by Dr Thomas Zürcher in 1983. This pharmacy has since become the Evangelical Hospital of Koyom, renowned beyond Chad, particularly for the quality of its surgical services.



Source: Solidarités International, 2019 Photo 1: Moringa with Small Leaves, Morphotype 2 in Chad



Source: Photo KAYALTO 2024

Photo 2: Moringa with Broad Leaves, Morphotype 1 in Kolobo, Chad

c) Physico-Chemical Analyzes

Samples of *Moringa* leaf powder, Morphotype 1, were analyzed in the laboratories of the Foodstuffs Quality Control Center (CECOQDA) in N'Djamena, Chad. Moisture, ash and lipid content were determined

using the MO PC method, while crude protein was analyzed using the MO PC EA method.

To enable us to make a comparison, we assayed the samples of *Moringa* with small leaves, Morphotype 2, from our previous work, using standard methods in the laboratories of the Centre for Research

in Biological, Food and Nutritional Sciences (CRSBAN) at the University Ouaga 1 Pr Joseph KI ZERBO in Ouagadougou, Burkina Faso. The samples were analyzed in duplicate.

i. Determining Moisture

The sample (5g) was oven-dried at $105 \pm 2^{\circ}C$ for 03 hours and the difference in weight gave the moisture content (AOAC 934.06, 1990);

ii. Determining the Total Protein Content

Total protein is determined by the Kjeldahl method (AOAC 920.87, 1990) based on total mineralization of the biological material in an acid environment, followed by distillation of the nitrogen in ammonia form. The mass of total plant protein is calculated using the formula: mass of nitrogen x 6.25 (FAO/WHO, 1978).

iii. Determination of Lipids

5 g of each sample was weighed and introduced into an extraction cartridge, covered by cotton. The cartridge was placed in a 150 ml glass Soxhlet. The solvent container was weighed and 400 ml of n-hexane was added. The soxhlet was then introduced into the container placed on the heating mantle, which was then connected to the cryostat cooling thermostat. Four to six siphoning processes were conducted over 5 hours. The heating mantle was disconnected. The solvent was then evaporated in a RE 121 Rotavapor (made in Switzerland). The container with the fat was placed in an oven for 3hours at 103°C, and then in a desiccator for 30 min and then weighed. The weight difference gives the fat content of the sample.

iv. Determination of Total Sugars

The determination of the total sugar content of the samples was performed in triplicate by spectrometric assay samples (Fox and Robyt, 1991). The reading of optical densities was made at 540 nm using a μ quant type plate reader (Bio-tek instrument Serial No. 157904, USA) coupled with a computer running KC integrated Junior (v1.31.5) software.

v. Détermination of ash rate

The sample (5 g) introduced in metal crucibles was mineralized in a muffle furnace (type VOLCA V50) at 550°C for five (05) hours, removed using thongs and then cooled in a desiccators for about one (01) hour before being weighed. The difference in weight gives the ash content of the sample (AOAC, 1990);

vi. Determination of Minerals

Mineralization was achieved through dry ashing. The ash obtained contains major elements (Na, Ca, Mg, K, etc.) and trace elements (Fe, Zn, etc.). These minerals were determined by *Atomic Absorption Spectrometry* (Pinta, 1973)(with a PELKIN Elmer model 3110 device (Connecticut, USA). A hollow Al-Ca-Cu-Fe-Mg-Si-Zn cathode lamp was used.

vii. Calculation of the Energy Value

The energy value corresponding to the available energy was calculated using [18] coefficients, coefficients adopted by the United Nations Food and Agriculture Organization (FAO) in 1970:

$$X = (P \times 4) + (G \times 4) + (L \times 9)$$

Where P = protein percentage, G = carbohydrates percentage, L = lipids (fats) percentage and X = energy value in Kcal/100g.

viii. Statistical Analysis

All assays were carried out in triplicate, and the averages and Standards Deviations (SD) calculation have been done with the software Exceland then transferred to Word version 2013.

III. Results

Knowledge of the physico-chemical composition of *Moringa* is an important factor in its development.

- a) Physico-Chemical Parameters of the Two Moringa Morphotypes
- i. Moringa Oleifera, Morphotype 2 Results

Table 1 presents the results of chemical analysis obtained in the CRSBAN laboratories on dried leaf powder of *Moringa oleifera*, Morphotype 2.

Table 1: Nutritional value, per 100g, of Moringa oleifera leaf powder, Morphotype 2 (mean ± SD)

Nutrients	Value	^a RDA
Energy (Kcal)	253,73	682 ^b
Moisture (%)	9,31 ± 0,18	
Ash (g)	$10,50 \pm 0,07$	
Protéins (g)	$24,28 \pm 0,22$	10,28°
Fats (g)	$7,42 \pm 1,56$	
Total Sugars (g)	$22,46 \pm 2,02$	
Vitamins		
(β-carotène)	624,40 \pm 0,41 μ g ER	400
Vitamin C (mg)	$65,88 \pm 0,00$	30
Minerals		
lron (mg)	$53,75 \pm 5,07$	11,6
Zinc (mg)	$17,58 \pm 0,89$	8.4
Calcium (mg)	1443,90 ± 11,03	400
Magnesium (mg)	$176,72 \pm 0,73$	54

RDA: Recommended Daily Allowance. ^aJoint FAO/WHO Expert Consultation, 2002. Vitamin and mineral requirements in human nutrition. Geneva: World Health Organization, 2002; ^bWHO (1998). Complementary feeding of young children in developing countries: a review of current scientific knowledge. UNICEF /University of California-Davis/WHO/ORSTOM. Geneva: WHO/NUT/98.1; ^cFAO/OMS/UNU, 1986. Energy and protein requirements. Report series.

The results obtained show that *Moringa oleifera*, Morphotype 2with small leaves, was more concentrated for most of the nutrients in our study, except for lipids (7.42 ± 1.56) and total sugars (22.46 ± 2.02): proteins (24.28 ± 0.22 g/100g), minerals, mg/100g: Ca 1443.90 ± 11.03; Mg 176.72 ± 0.73; Fe 53.75 ± 5.07; Zn 17.58 ± 0.89 and vitamins (β-carotene 624.40 ± 0.41µg ER, Vitamin C 65.88 ± 0.00 mg/100g). The high mineral content of *Moringa oleifera* powder is easily explained by its high ash content (10.50g per 100g). ii. Moringa from the Village of Kolobo, Morphotype 1 Results

Table 2 shows the results obtained in the physical and chemical laboratories of CECOQDA, Department of Physical and Chemical Quality Control of Food, Water and Beverages, sample No. 0360/CECOQDA/DCQPC/2024 dated 05/11/2024. We have only obtained the results of analyzes of water, ash, protein and lipid content. For the rest, minerals and vitamins, CECOQDA is awaiting its order for reagents.

Table 2: Nutritional Value, per 100g, of Moringa Oleifera Leaf Powder from the Village of Kolobo, Morphotype 1

Nutrients	Value	aRDA
Moisture (%)	13,73 ± 0,12	
Ash (%)	$9,94 \pm 0,05$	
Proteins (%)	$6,09 \pm 0,15$	10,28°
Fats (%)	$0,52 \pm 0,02$	

Table 3 shows the comparative chemical composition of macronutrients and ash in the two *Moringa oleifera* morphotypes.

Table 3: Comparative Macronutrient Composition of the Two Morphotypes of Moringa Oleifera (Mean±SD)

Morphotypes	Moisture (%)	Proteins (%)	Fats (%)	Ash (%)
Morphotype 2	9,31 ± 0,18	$24,28 \pm 0,22$	7,42 ± 1,56	$10,50 \pm 0,07$
Morphotype 1	$13,73 \pm 0,12$	$6,09 \pm 0,15$	$0,52 \pm 0,02$	$9,94\pm0,05$

IV. DISCUSSION

Small-leaf *Moringa* is a nutrient concentrate, as shown by the results summarized in Table 1, except for lipids (7.42 ± 1.56) and total sugars (22.46 ± 2.02) . In one of our previous studies (Barnabas *et al.*, 2024), we used the powder of these leaves in the nutritional recovery of 416 moderately acutely malnourished children (MAM) at the Chagoua Hospital Notre Dame of

Apostles (HNDA). The results of this study showed that moderately malnourished children given porridges with dried *Moringa* leaf powder took a maximum of three weeks to recover from malnutrition. The millet (Pennisetum typhoïdes) porridge with *Moringa* gave the best results, with 96% of children recovering after an average stay of 17.5 days. Children on red sorghum porridge from Bongor with *Moringa* showed the highest rate of haemoglobin gain at 13.5 g/dl. Children on maize porridge without *Moringa* (control) took the longest to recover in the study, averaging 32.7 days.

Comparing our results with those of Compaoré et al. (2011) in Burkina Faso, *Moringa* from Burkina has a high lipid content (43.56 ± 0.03 g/100g) and a low total sugar content (9.17 ± 0.25) which is the opposite for Chad, respectively 7.42 \pm 1.56 g/100g for lipids and 22.46 \pm 2.02 for total sugars.

Moringa broadleaf has a low protein content (6.09 g/100g) compared with *Moringa* obtained at Gounou Gaya. Its lipid content is very low (0.52 g/100g). However, this species has an ash content (9.94 g/100g) similar to the *Moringa* obtained at Gounou Gaya (10.50 \pm 0.07 g/100g), which suggests a high minerals content that was not analyzed due to a break in the reagents at the time of our study.

Ndong *et al.* (2007) in Senegal, in their study of dried *Moringa oleifera* leaf powder, obtained higher results than ours, in terms of protein, carbohydrate, calcium, potassium, magnesium, ash and energy content. These values were respectively: 39.69 ± 0.01 ; 35.33; 1526.74 ± 50.03 ; 888.50 ± 38.30 ; 428.87 ± 85.96 ; 11.39 ± 0.66 and 358.73. As for iron and zinc content, we obtained 53.75 ± 5.07 and 17.58 ± 0.89 respectively, higher results than those obtained in their study, which were 18.86 ± 1.20 and 2.13 ± 0.07 respectively. However, in their study, Ndong *et al.* found low iron bioavailability (%) in dried *Moringa oleifera* leaf powder, i.e. 2.24 ± 0.65 . Our results regarding the lipid content of dried *Moringa* leaves from Chad (7.42 ± 1.56 g/100g) are similar to those from Senegal, i.e. 7.85 ± 0.28 .

As regards the minerals composition of broadleaf *Moringa* leaves from Chad, the reagents for these analyses were not available at the time of our study in the laboratories of the Foodstuffs Quality Control Center (CECOQDA) in Chad. For information purposes, we give here the average values taken from page 209 of the FAO table of food composition for West Africa.

Table 4: Minerals (mg/100 g) in fresh raw Moringa leaves and fresh boiled leaves

Minerals	Fresh raw leaves	Fresh boiled leaves
Calcium	595	633
Iron	10,3	10,9
Magnesium	68	73
Phosphorus	91	103
Potassium	405	428
Sodium	9	10
Zinc	1,20	1,28
Copper	0,21	0,22

Source: Vincent et al, (2020), FAO/INFOODS food composition table for West Africa (2019), p.209

According to Manzo et al. (2016), in Niger, the use of local foods has been identified in the national protocol for the management of malnutrition as an alternative to the ready-to-use therapeutic foods currently in use. Moringa oleifera dried leaf powder was identified as one of these local foods. They analyzed four Moringa samples from the three main Moringa production regions in Niger, namely Tillabéri and Niamev in the river basin and the south of Maradi in the Sahelo-Sudanian zone. According to the above authors, the dried Moringa leaf powder produced in Niger was found to be rich in protein, with an average of 24.8%. Depending on the region, the composition varied between 51.9 and 55.12 mg/100g for iron; 0.45 and 1.58 mg/100g for zinc; 1192.5 and 1957.5 mg/100g for calcium; 414.37 and 714.37 mg/100g for magnesium; 1587 and 2037 mg/100g for potassium; 207.75 and 326.25 mg/100g for sodium; 32 and 61 mg/100g for phosphorus.

Our results for dried *Moringa* leaf powder from Chad are similar to those from Niger in terms of protein, iron and calcium. *Moringa* from Niger has an average magnesium content (551 mg/100g) three times higher than that from Chad (176.72 \pm 0.73 mg/100g). On the other hand, dried *Moringa* leaf powder from Chad has a zinc content (17.58 \pm 0.89 mg/100g) 19 times higher than that from Niger (0.92 mg/100g).

Moringa oleifera Lam. is a leafy vegetable that is very rich in nutrients and is increasingly recommended for populations suffering from malnutrition. Moringa leaves, commonly known in local Arabic as "Haloum", are a staple food in some Chadian households. They are used in the preparation of sauces. "I use Moringa leaves to make sauce, and I boil the leaves to make herbal tea to treat malaria. I also use the roots of this plant to treat tooth decay. My children and me eat the seeds for stomach problems such as typhoid. I also use soaps made from *Moringa* to make my skin smooth", these are the answers given by some of the women interviewed by Solidarités International. According to traditional healers, Moringa leaves have been around for centuries. People all over the world have used Moringa leaves as a food, but also for its medicinal properties (Solidarités International, 2019).

HOUNDJI et al. (2013) assessed the effect of a daily intake of 10 g of Moringa oleifera leaf powder on the nutritional status of moderately acutely malnourished children aged 6 to 30 months after 6 months of supplementation. Two groups were formed from 84 infants selected from a nutritional recovery center in the village of Lissèzoun (Centre-Benin): one received PFMo and the other, chosen as a control, did not receive this powder. The nutritional status of the children was assessed using anthropometric indicators. At the end of the 6 months, the results showed that daily supplementation with PFMo significantly improved the nutritional status of the children, both for wasting (Zscore Weight/Height of -1.0 \pm 0, 9 at the start of the intervention to 0.7 ± 1.0 at the end of 6 months), stunting (Z-score Height/Age from -2.6 \pm 0.7 to 0.4 \pm 0.7) and underweight (Z-score Weight/Age from -2.2 \pm 0.6 to 0.7 \pm 0.7). The improvement in Z-scores was greater in the intervention group than in the control group and resulted in zero prevalence for the three types of malnutrition at the end of the experimental period.

According to Nikiema *et al.* (2009), in Burkina Faso, natural substances are also recommended by traditional health practitioners for immunological and nutritional recovery, early treatment of HIV infection and reducing the side effects of ARV (antiretroviral) treatment. The most important of these are the leaves of *Moringa oleifera* Lam. (Moringaceae), the pulp of the fruit of *Detarium microcarpum* Guill. & Perr. (Fabaceae), spirulina and pollen from the beehive.

TETE-BENISSAN et al. (2012) evaluated the influence of leaf powder consumption on serum protein changes during nutritional recovery in malnourished subjects. They reached the following conclusions: Infants (29 HIV-negative and 26 HIV-positive) and children (27 HIV-negative and 32 HIV-positive) of both sexes aged between 12 months and 8 years were compared. After 16 weeks of *M. oleifera* use, anthropometric parameters were measured, and assays of total protein, creatinine, ASAT, ALAT and GT were performed, along with a proteinogram. Nutritional recovery with M. oleifera showed that BMI increased significantly in both HIV-negative and HIV-positive subjects (p<0.001). The significant increase in albumin concentrations correlated with a significant decrease in total protein, $\alpha 1$, $\alpha 2$, β , γ , globulin fractions, creatinine, AST, ALT and GT after use of the dietary supplement. The results of the study also showed that the improvement in nutritional status, inflammatory status and immune status was greater in HIV-negative subjects than in HIV-positive subjects. This study confirms the nutritional qualities and pharmacological properties of M. oleifera leaves in the fight against malnutrition and micronutrient deficiencies. Thus, despite the profound metabolic disruption caused by viral infection, M. oleifera effectively helps to reduce the inflammatory state in HIV-positive subjects.

Moringa oil also contains around 76% linolenic acid and oleic acid, making it a potential substitute for olive oil. *Moringa* leaves contain an exceptionally high amount of protein compared with other leaves consumed as food, and essential amino acids such as lysine, tryptophan, phenylalanine, valine, etc. (Trigo *et al.*, 2021).

The *Moringa* tree has been reported to have high economic and cultural values, which has also led to the emergence of *Moringa* plantation and processing businesses with its implication in job creation and therefore poverty reduction. Many families, particularly women, are involved in the distribution and sale of fresh *Moringa* leaves. They make a living from supplying urban centers on a daily basis (Omotesho *et al.*, 2013).

In Nigeria, the average yield of wet leaves per *Moringa* plant is 4.5 kg per year, equivalent to 1 kg of dry leaf powder, giving a ratio of 4.3 kg to 1 kg of dry organic leaf powder. On average, one hectare of *Moringa* plant will yield 50,616 kg of dry leaf powder and a gross income of \$75,924 per year. It is important to note that the *Moringa* plant produces for seven years or more (Omotesho *et al.*, 2013).

Moringa leaves contain antinutritional factors such as oxalate and phytate and tannins that can reduce the bioavailability of certain nutrients. Treatments such as roasting (Alidou *et al.*, 2016) and boiling (Sallau *et al.*, 2012) have been reported to effectively reduce the content of these anti-nutrients, thereby increasing nutrient bioavailability. Boiling leaves before consumption is a traditional practice of local communities in Zaria, Nigeria.

Moringa leaves are rich in polyphenols, the main components of which are flavonoids and phenolic acids. The flavonoids found are mainly guercetin and kaempferol, while the phenolic acids are mainly gallic, chlorogenic, ellagic and ferulic acids. The leaves also contain saponins, tannins, catecholic tannins, anthraquinones and alkaloids (Omede, 2016). Moringa has remarkable health benefits, including antioxidant, antimicrobial, anticancer, anti-inflammatory and antidiabetic activity (Trigo et al., 2021; Ghimire et al., 2021; Kashyap et al., 2022). There is also growing interest in using Moringa as a value-added ingredient in the development of functional foods (Kashyap et al., 2022).

V. CONCLUSION

At the end of this study, a number of physicochemical parameters were determined. The study revealed that *Moringa oleifera* leaf powder with small leaves, Morphotype 2 is rich in protein, calcium, magnesium, iron, zinc and β -carotene. Dried large-leaf *Moringa oleifera*, Morphotype 1 leaf powder is low in protein and very low in lipids. Its ash content is almost the same as that of the small leaves. From a nutritional point of view, large-leaf Moringa oleifera, Morphotype 1 is not very suitable for treating cases of protein and lipid deficiency.

The processing of Moringa leaves into powder remains a good practice for better preservation of the product but also for a good concentration of nutrients. Analysis of the chemical composition revealed the particularities of dried Moringa oleifera leaf powder with small leaves, which would be of great interest from a nutritional point of view.

References

- 1. Alidou C, Salifou A, Djossou J, Mazou M, Tchobo FP, Soumanou MM (2016). Roasting effect on antinutritional factors of the Moringa oleifera leaves. Int. J. of Adv. Res. 4, 78-85.
- Anwar F, Latif S, Ashraf M and Gilani AH. (2007). 2. Moringa oleifera: a food plant with multiple medicinal uses. Phytotherapy Research 21(1): 17-25.
- AOAC, "Official Methods of Analysis," 15th Edition, З. Association of Official Analytical Chemists, Washington DC, 1990, pp. 808,831-835, 1113.
- Barnabas K., Cheikna Z., Raketa W. C., Aly S., Brahim B. O., Alfred S. T., 2013. Study of the nutritional value and hygienic quality of local infant flours from Chad, with the aim of their use for improved infant flours preparation. Food and Nutrition Sciences, 2013, 4, 59-68. http://dx.doi. org/10.4236/fns.2013.49A2009 Published Online September 2013 (http://www.scirp.org/journal/fns).
- Barnabas K., Djimadoum K., Brahim B.O., Emilienne S., Alfred S.T., 2024. Assessment of Biological Effectiveness of Infant Flours on the Iron Status and Speed of Recovery of MAM Children Admitted to HNDA Hospital at N'Djamena (Chad). Central African Journal of Public Health, Vol. 10, N°6, pp. 285-296.
- Bidossessi VSH, Sam F. B, Serge BML, Romaric O, 6. Alfred Α. Kou'Santa SE. AMOUZOU et HOUNMENOU, 2013. Amélioration de l'état nutritionnel des enfants âgés de 6 à 30 mois à Lissèzoun (Centre-Bénin) par la poudre de feuilles de Moringa oleifera(Lam.). Int. J. Biol. Chem. Sci. 7(1): 225-235, February 2013
- Compaoré WR, Nikièma PA, Savadogo A, Bassole 7. HIN, Mouecoucou J, Pignatelli S, Simporé J and Traoré SA. (2011). Nutritional and Hematological Recovery of Severely Malnourished Children with New Flour Formulas Developed at Ouagadougou (Burkina Faso). British Journal of Dairy Sciences 2 (1): 11-17, 2011.
- 8. Hounhouigan D. J. and Traoré S. A., 2011. Chemical Composition and Antioxidative Properties of Seeds of Moringa oleifera and Pulps of Parkiabiglobosa and Adansoniadigitata Commonly used in Food

Fortification in Burkina Faso. Current Research Journal of Biological Sciences 3(1): 64-72, 2011

- De Jong W., Campbell B. M. & Schroder, J-M., 2000. 9. "Sustanining incomes from non timber forest products: introduction and and synthesis". International Tree Crops Journal, 10 (4): 427-275.
- 10. Foidl, N., Harinder, P. S. & Becker K, 2001. Potentiel du Moringa oleifera pour les besoins agricoles et industrielsin L'arbre de la vie, Les multiples usages du Moringa. CTA etCTA et CWS, Dakar, 45-78 p.
- 11. Fox J. D. and Robyt J. F., 1991. "Miniaturization of Three Car- bohydrates Analyzes Using a Microplate Reader," Analytical Biochemistry, Vol. 195, No. 1, 1991, pp. 93-96. doi:10.1016/0003-2697(91)90300-l
- 12. Ghimire S, Subedi L, Acharya N, Gaire BP. (2021). Moringa oleifera: A Tree of Life as a Promising Medicinal Plant for Neurodegenerative Diseases. J. Agric. Food Chem. 69, 14358-14371 doi: 10.1021/ acs.jafc.1c04581
- 13. Nikiéma J. B., K. Dijerro, J. Simpore, D. Sia, S. Sourabié, C. Gnoula et I.P. Guissou, 2009. Stratégie d'utilisation des substances naturelles dans la prise en charge despersonnes vivant avec le VIH: expériencedu Burkina Faso. Ethnopharmacologia, n°43, juillet 2009. Dossier spécial: Médecine traditionnelle en Afrique.
- 14. Kashyap P, Kumar S, Riar CS, Jindal N, Baniwal P, Guine RF, Correia PMR, Mehra R, Kumar H. (2022). Recent Advances in Drumstick (Moringa oleifera) Leaves Bioactive Compounds: Composition, Health Benefits, Bioaccessibility, and Dietary Applications. Antioxidants (Basel), 11, 402, doi: 10.3390/antiox11 020402
- 15. Manzo ML, Halidou DM, Hallarou M, Illo A, Rabani A, Donnen P and M Dramaix, 2016. Composition de la poudre des feuilles sèches de Moringa oleifera dans trois régions du Niger. Afric. J. of Food Agric. Nut. and Dev., 2016; 16(4). DOI: 10.18697/ajfand. 76.15115
- 16. Mapongmetsem P. M. & Zedong C. H., 1997. Situation du bois de chauffe dans les savanes humides de l'Adamaoua. Le flambovant. 42: 29-33.
- 17. Mapongmetsem P. M., Nkongmeneck B. A., Alexandre D., Salba I. & Haoua M., 2010. Jardins de case et sécurité alimentaire dans les savanes soudano-guinéennes (Adamaoua-Cameroun). Nanterre, Franve, Symposium Mega-Tchad 2002. 12 p.
- 18. Melom S, Mbayngone E et Mapongmetsem PM. (2016). Caractérisation variétale de Moringa oleifera Lam. (Moringaceae) dans la vallée du Logone (Tchad-Cameroun). Annales de l'Université de N'Djamena Série C N°8.
- 19. Ndong M, Wade S, Dossou N, Guiro AT and Gning RD. (2007). Valeur nutritionnelle du Moringa oleifera, étude de la biodisponibilité du fer, effet de l'enrichissement de divers plats traditionnels sénégalais avec la poudre des feuilles. African

Journal of Food, Agriculture, Nutrition and Development7 (3): 17 pages.

- 20. Ngamo L.S.T., 2004. Rapport technique (septembre 2004). 2003-Février Store protect, proiet interuniversitaire ciblé (PIC), N'Gaoundéré, Cameroun. 34 p.
- 21. Omede A. (2016). Total polyphenolic content and antioxidant properties of Moringa oleifera leaf extracts. Animal Research International 13, 2454 -2462.
- 22. Omotesho, K. F., Sola-Ojo, F. E., Fayeye, T. R., Babatunde, R. O., Otunola, G. A., & Aliyu, T. H. (2013). The potential of Moringa tree for poverty alleviation and rural development: Review ofevidences on usage and efficacy. International Journal of Development and Sustainability, 2(2), 799-813.
- 23. Pinta M., "Méthodes de Référence pour la Détermination des Éléments Minéraux dans les Végétaux. Détermination des Éléments Ca. Mg. Fe. Mn, Zn et Cu par Absorption Atomique," Oléagineux, Vol. 28, No. 2, 1973, pp. 87-92.
- 24. Saint Sauveur A, Broin M. (2010). Produire et transformer les feuilles de Moringa. Editions CTA, CDE, Horizon Gémenoéd., France, 69 pages.
- 25. Sallau B, Mada SB, Ibrahim S, Ibrahim U. (2012). Effect of boiling, simmering and blanching on the antinutritional content of Moringa oleifera leaves. Int. J. Food Nutr. Saf. 2, 1–6.
- 26. Solidarités international, 2019. Moringa: contre la malnutrition au Tchad, l'arbre de vie, 22 mai 2019.
- 27. Tete-Benissana, Quashiem-La, Lawsonevik, Kokouk, Gbeassorm, 2012. Récupération nutritionnelle chez les sujets malnutris VIH positifs et VIH négatifs après utilisation de feuilles de Moringa oleiferaLam. Journal of Animal & Plant Sciences, 2012. Vol.15, Issue 2: 2184-2199
- 28. Trigo C, Castello ML, Ortol MD, Garcia-Mare FJ, Desamparados Soriano M. (2021). Moringa oleifera: An Unknown Crop in Developed Countries with Great Potential for Industry and Adaptedto Climate Change, Foods, 10, 31. doi: 10.3390/foods100 10031
- 29. Urbain Zongo, Steve Léonce Zoungrana, Aly Savadogo, Alfred S. Traoré, (2013). Nutritional and Clinical Rehabilitation of Severely Malnourished Children with Moringa oleifera Lam. Leaf Powder in Ouagadougou (Burkina Faso). Food and Nutrition Sciences 4: 991-99
- 30. Vincent A, Grande F, Compaore E, AmponsahAnnor G, Addy PA, Aburime LC, Ahmed D, BihLoh AM, Dahdouh Cabia S, Deflache N, Dembele FM, Dieudonne B, Edwige OB, Ene-Obong HN, Fanou Fogny N, Ferreira M, OmaghomiJemide J, Kouebou PC, Muller C, Najera Espinosa S, Ouattara F, Rittenschober D, Schonfeldt H, Stadlmayr B, van Deventer M, Razikou Yiagnigni A & Charrondiere UR.

Table de composition des aliments (2020). FAO/INFOODS pour l'Afrique de l'Ouest (2019) Guide d'utilisation & table de composition des aliments condensée. Rome, FAO.