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# GLOBAL JOURNAL

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# Chemical Engineering

Influence of Scan Rate

Hydrochloric Acid Solutions

Highlights

Simple Vacuum Distillation

Differential Scanning Calorimetry

VERSION 1.0

Discovering Thoughts, Inventing Future

VOLUME 15

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## GLOBAL JOURNAL OF RESEARCHES IN ENGINEERING: C Chemical Engineering

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# Simple Vacuum Distillation of Vetiver Oil from Smallholders for Quality Improvement

By I Dewa Gede Arsa Putrawan & Eric Farda

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Abstract- This research is aimed to improve the quality of vetiver oils from smallholders in Indonesia by vacuum distillation. The most important parameters of quality mentioned are total vetiverol content and color. It was shown that vetiverol contents could be increased to achieve the required minimum content of 50%. The better the initial sample, the better the distillate obtained. Distillate fractions obeying standard vetiverol content could be obtained with yield of 60%~80%. Although the initial samples were black in color, the distillates had appearance from yellow to reddish brown, as required by the standard, with Gardner scales of color ranging from 10.8 to 14.7. Distillation, however, slightly disturbed the achievement of other parameters including density, acid number and ester number. Lower distillation fractions tend to shift the values of these parameters to out of standards.

Keywords: vetiver oil, simple vacuum distillation, quality improvement.

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# Simple Vacuum Distillation of Vetiver Oil from Smallholders for Quality Improvement

I Dewa Gede Arsa Putrawan <sup>a</sup> & Eric Farda <sup>o</sup>

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#### I. INTRODUCTION

Vetiver oil is an essential oil extracted from the roots of Vetiver grass (Chrysopogon zizanioides, Linn Nash), a tropical grass growing wild or cultivated. The extraction can be done via water distillation, steam distillation, solvent extraction or expression (Dowthwaite and Rajani, 2000). As an essential oil, vetiver oil will find wider application due to the increasing tendency towards the use of natural materials. The classical roles of vetiver oil are flavor, fragrance, and aromatherapy (Lavania, 2003). It has been observed also that the vetiver oil has activities such as antimicroorganism, antioxidant, insectiside, and sedative.

Indonesia produces 60 to 75 tons of vetiver oil annually and becomes the main vetiver oil suppliers in the words, besides Haiti and La Réunion. About 90% of Indonesia vetiver oil is produced by smallholders in Garut, a small district about 200 km from Jakarta to the south-east. It is produced by water distillation at 4 to 5 bars in the conventional ways. The main equipment consists of a boiling vessel, a burner, and a condenser.Vetiver roots are packed on grids over a layer of water in the base of the vessel. Saturated steam which is produced by direct firing the vessel extracts the essential oil contained in the roots. Essential oil containing steam is then condensed to obtain vetiver oil. Due to primitive equipment and the lack of process control, the quality of most vetiver oil obtained could not satisfy the required standard. Minimum total vetiverol content of 50% is hard to obtain. In addition, the color of the vetiver oil is very dark, from dark brown to black. It is supposed due to overheating and oxidative degradation which results in tar like substances which are hard to separate by physical methods. Standard on vetiver oil requires the color from light yellow to reddish brown. Further treatment, therefore, is necessary to improve such vetiver oils in order to comply with standard qualities.

Many works have been published on vetiver oil. Aggarwal et al. (1998) studied the effects of harvest period, storage period after harvest, roots treatment and distillation time on the yield of extraction by distillation. They found that the extractable oil decreased with harvest delay and storage period. In addition, they found that cutting the roots did not affect oil yield significantly, water distillation gave oil recovery a little bit higher than steam distillation (0.28% vs 0.23%, based on dry root weight). Danh et al. (2010) have studied the extraction by an unconventional method, i.e., an extraction by supercritical CO<sub>2</sub> with ethanol as co-solvent. They found that pressure and ethanol concentration linearly affected oil yield but, in contrary, temperature in the range of 40 to 50 °C did not influence oil vield. It was found also that supercritical fluid extraction did not extract the metals in the vetiver roots giving an advantage for strict applications, such as foods and drugs. The effects of plant environments on the quantity and quality of extracted oil have attracted the attention of researchers. Adams et al. (2004) reported that cleansed vetivers gave lower oil yield with strictly different composition to those of non-cleansed vetivers. They found that the yield of oil from bacteria and fungi free vetivers was almost twenty times lower and contained large amounts of C19-C29 alkanes. The roles of microbes were also reported by Pripdeevech et al. (2006) who obtained larger oil yield and higher content of some low molecular weight volatiles from vetiver plants grown in normal soil with added microbes compared to normal soil (0.27% vs 0.18%). Adams et al. (2008), however, reported no correspondence between arbuscular mycorrhizal fungi colonization and oil yield, and postulated that the composition of vetiver oils are majorly controlled by vetiver genes. Massardo et al. (2006) reported variations in oil yield and in the contents of isovalencenol and

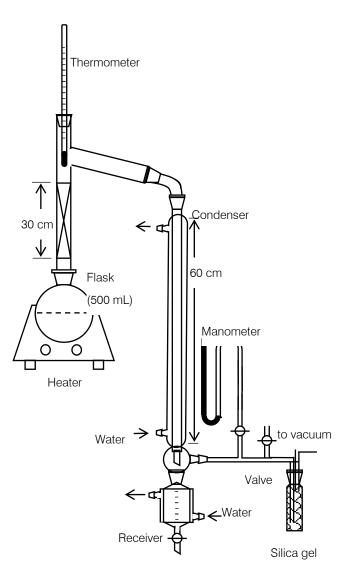
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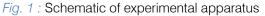
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khusimol in the obtained oils during growth of roots. The effects of climatic conditions have also been studied by Kotoky et al. (2011) who found that rain-fall played a role on the yield and quality of vetiver oils. Their data indicated that vetiver oil guality is closely related to the metabolism of its root which is influenced by climatic conditions. The complexicity of vetiver oil composition has challenged many researchers in developing gualitative and guantitative analysis, among them are Cazzausus et al. (1988), Weyerstahl et al. (200a, 200b), Paillat et al. (2012) and Filippi et al. (2013). The works of Weyerstahl et al. (2000a, 2000b) and Filippi et al. (2013) are very intensive. In recent years, biological functionalities of vetiver oil have much attention. The antibacterial and antifungal activities of vetiver oil have been reported by Prabuseenivasan (2006), Gupta et al. (2012), and Sangeetha and Stella (2012). The insecticidal activities of vetiver oil have been reported by et al. (2001a, 2001b). Zhu The antioxidant, antinociceptive, and anti-inflammatory activities, the sedative effects, and the toxicity of vetiver oil have also been studied (Kim et al., 2005, Lima et al., 2012, Thubthimthed et al., 2003, Sinha et al., 2014). Apart from the many works, no attention has been paid to upgrade the poor quality of crude vetiver oil which commonly found from smallholders.

#### II. MATERIALS AND METHODS

Fig. 1 shows the schematic of the distillation apparatus. The apparatus mainly consisted of a distillation column with insulation, a condenser and a condensate collector. The apparatus was equipped with a cooling water system with controlled temperature and connected to a vacuum system. The pressure inside the apparatus was controlled by a needle valve and was set at 10 mmHg. Boiling points were measured by using a thermometer. Crude vetiver oil of 300 ml was used for each run. Distillates were collected at every 20% recovery until 80% volume. Three vetiver oil samples from small distillers were used. The parameters observed included density, refractive index, acid number, ester number, ester number after acetylation, and vetiverol content. Density was measured by a piknometer. Refractive index was measured using a refractometer. Acid number was measured by titration. Ester number was measured by saponification. Acetylation was done using acetic anhydride. The content of free alcohol as vetiverol was calculated from ester number of original oil and that of acetilated oil. The parameters observed also included color, odor, and solubility in ethanol. Color was observed visually and also measured by a Lovibond Tintometer. Solubility in ethanol was observed as "soluble" or "not soluble" in ethanol at volume ratio of 1:1. All chemicals for analysis were of pro analysis grade.





#### III. Results and Discussion

#### a) Vetiver Oil Samples

The characteristics of three vetiver oil samples studied here are given in Table 1, columns 3 to 5. The Indonesia standard quality (SNI, 2006) are shown in the rightmost column. As shown in the table, all three samples had vetiverol contents lower than the required standard. Moreover, they smelt smoky and their color were dark black. These three quality parameters are commonly fail to achieve by most smallholders. It is due to poor distillation operation. Many factors affecting the distillation of vetiver root as discussed by Aggarwal (1998) which are not considered in the field. The smoky odor comes from soil materials which were not removed prior to distillation. The vetiverol content appeared to correlate with density. In this case, lighter vetiver oil exhibited higher vetiverol content. All samples were soluble in ethanol at 1:1 volume ratio.

Table 1 also shows the characteristics of Indonesian vetiver oil samples from Guenter [1950], columns 6 to 9. This author procured four samples of vetiver oils from Indonesia in the past, said to be genuine samples, with physicochemical properties as shown in Table 1. The data was completed with vetiverol contents which were calculated in this work from the ester numbers. In regards to vetiverol content, the samples used in this work were inferior to those from Guenter [1950]. The samples used here had higher ester numbers but lower ester numbers after acetylation resulting in lower total vetiverols. In contrast to the samples of this work, the genuine samples from Guenter [1950] complied with the required standards.

#	Deremeter	Samp	les in this	s work	Guenther (1950)†				Standardt	
#	Parameter	Α	В	С	Ι	II	III	IV	Standard‡	
1	Relative density (20/20 °C)	0.987	0.994	1.024	1.009	1.009	1.007	0.991	0.980 to 1.003	
2	Refractive index (20 °C)	1.52	1.52	1.52	1.526	1.5271	1.526	1.5258	1.52 to 1.53	
3	Acid number (mg KOH/g)	27	30	34	31.1	32.5	30.2	12.5	10 to 35	
4	Ester number (mg KOH/g)	24	22	26	2	12.6	1	4	5 to 26	
5	Ester number after acetylation (mg KOH/g)	133	124	117	141.5	150	152.1	129	100 to 150	
6	Total vetiverol (%)	46	43	39	61*	60*	67*	54*	Minimum 50%	
7	Color	Too dark	Too dark	Too dark	NA	NA	NA	NA	Pale yellow to Reddish brown	
8	Odor	Smoky	Smoky	Smoky	NA	NA	NA	NA	Vetiver characteristic	
9	Solubility in ethanol 95% (1:1)	Clear	Clear	Clear	NA	NA	NA	NA	Clear	
#	Parameter	Samp	les in this	s work	G	uenthe	r (195	0)†	Standard‡	
π	Falainelei	A	В	С	I	II	III	IV	-	
1	Relative density (20/20 °C)	0.987	0.994	1.024	1.009	1.009	1.007	0.991	0.980 to 1.003	
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7	Color	Too dark	Too dark	Too dark	NA	NA	NA	NA	Pale yellow to Reddish brown	
8	Odor	Smoky	Smoky	Smoky	NA	NA	NA	NA	Vetiver characteristic	
9	Solubility in ethanol 95% (1:1)	Clear	Clear	Clear	NA	NA	NA	NA	Clear	

Table 1 : Characteristics of vetiver oil samples

†Genuine samples analyzed by Guenther (1950). \*Calculated in this work from ester numbers. ‡National Indonesian Standard. NA: not available

#### b) Distributions of Physicochemical Properties

Fig. 2 to 7 illustrate the distribution of physicochemical properties at various distillate recovery. The densities of distillates varied in the range of 0.94 to 1.00, as shown in Fig. 2. As can be expected, lighter distillate distillate had lower relative density. All distillates from samples A and B had density lower than the required minimum density, although their initial sample obeyed the required standard density. The density of sample C was higher than the upper limit of standard density. However, its distillates from 60% to 80% recovery were in the range of standard density. As seen in the figure, the range of standard density is narrow. The deviation in densities were actually not significant, only at the second decimals. The largest was shown by 20% recovery of sample A (a deviation of 0.04 from the lower limit). Fig. 3 shows that the refractive index did not vary significantly. All distillates had refractive index in or close to the range of 1.52 to 1.53, except at 20%-recovery.

In contrast to essential oil in general, vetiver oil exhibited high content of free fatty acid. The contents of free fatty acid as acid numbers are shown in Fig. 4. In general, acid number increased with %-recovery. Lower %-recovery tend to decrease acid number and make the acid number lower than the minimum acid number. The distillation cuts between 40% and 60% and between 60% and 80% had acid numbers significantly larger than the other lighter distillation cuts. This indicated that acid compounds in vetiver oil concentrate at the heavier fractions.

Fig. 5 and Fig. 6 shows the ester numbers before and after acetylation, respectively. Ester number before acetylation in general increased with %-recovery but ester number after acetylation decreased with %-recovery. Most of distillate obeyed the standard for ester numbers before acetylation. Ester numbers after acetylation of light distillate (20% to 40%) were higher than the maximum allowed numbers. As difference in these two ester numbers indicates vetiverol content, this indicated that the viteverol content should be higher at the lighter distillate.

Fig. 7 shows the distribution of vetiverol content. Vetiverol contents were found to decrease with distilate recovery. The better the sample, the higher the vetiverol content of the distillate obtained. Minimum vetiverol content of 50% could be obtained from samples A and B until 60% recovery and almost achieved at 80% recovery. It means that the vetiverol content of samples A and B could be improved to very close to the required standard at the expense of 20% oil lost. With sample C, however, more quantity have to be sacrificed to achieve standard content of vetiverol. At 80% recovery, sample C gave distillate with a vetiverol content of 42%. It is significantly lower than the required standard, although the vetiverol contents have improved compared to the initial samples. Minimum vetiverol content of 50% could be achieved until 60% recovery for sample C.

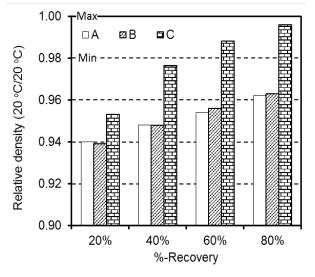
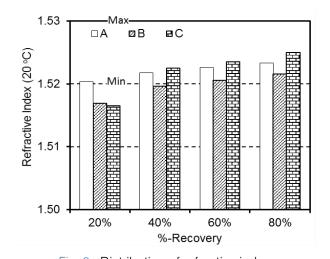
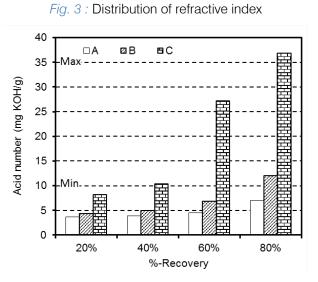


Fig. 2 : Distribution of relative density







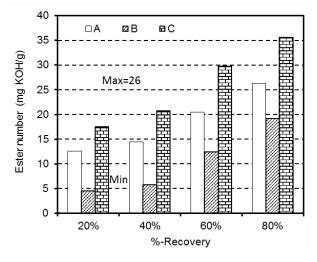


Fig. 5 : Distribution of ester number

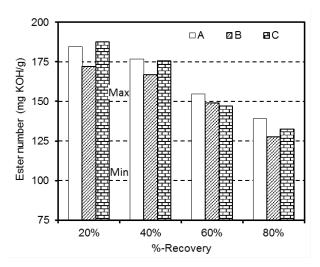
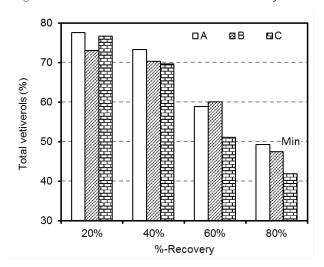


Fig. 6 : Distribution of ester number after acetylation



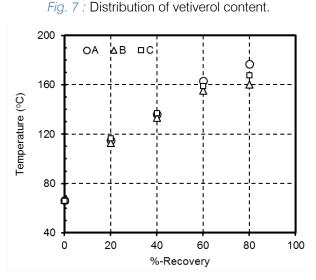


Fig. 8 : Distribution of boiling point

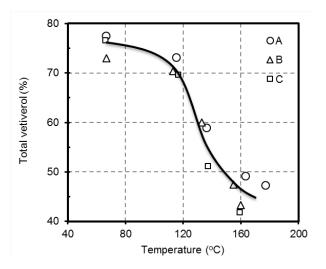


Fig. 9 : Vetiverol content vs boiling point.

Fig. 8 shows the temperature curves of distillation. The boiling temperatures of all samples were close each other, although they were getting away along with temperature. The relationship between vetiverol content and boiling point are shown in Fig. 9. Although the data is rather scattered, they could be approximated by a third order polynomial. More importantly, it clearly shows that lower distillation temperature gave higher vetiverol content. This reveals that the impurities are concentrated at the heavy fractions and distilling the samples under vacuum improved their vetiverol contents.

There are no references for the complete distribution of acid and ester numbers and vetiverol content in respect to vetiver oil distillation cuts. Gildemeister (1913) and Weyerstahl et al. (2000a) presented several distillation cuts of vetiver oils, however, for the purposes of showing the cuts in which individual components are concentrated. The results here showed that all parameters regarding compound groups in vetiver oil were continueously distributed. This indicated that vetiver oil composition is quite complex, consisting of many components with a wide boiling point range. Hundreds of components have been identified (Weyerstahl et al., 2000b and Filippi et al., 2013). However, many components are still unidentified.

#### c) Color, Odor and Ethanol Solubility

All distillates were soluble in ethanol at volume ratio of 1:1. The Indonesian standard qualitatively requires that vetiver oil has color from pale yellow to reddish brown. None of the vetiver oil samples studied here comply with this color standard. The colors of all samples were dark (almost black). In contrast, all distillates complied with the required qualitative standard. Although there is no quantitative standard for color of vetiver oils, popular color scales for oils and fats or chemicals in general were used in this study to describe the colors of distillates. Sample C was selected to study the color as sample C was the darkest sample. Year 2015

The colors of distillates from sample C were measured using a Lovibond Tintometer. Table 2 shows the results in the scales of Lovibond RYBN, AOCS RY, FAC, Gardner, and ASTM. Lovibond RYBN scale is based on different densities of red, yellow, blue and neutral. The AOCS-Tintometer Color Scale is a special red and yellow version of Lovibond RYBN. FAC Scale is divided into 5 groups: Scale 1 (1, 3, 5, 7, 9) for lighter colored fats; Scale 2 (11, 11a, 11b, 11c) predominantly for yellow fats; Scale 3 (13, 15, 17, 19) for dark fats (red cast); Scale 5 (31, 33, 35, 37, 39, 41, 43, 45) for very dark fats, predominantly red. The Gardner scale ranges from a pale yellow to a red in shade and is described in terms of the values 1-18. ASTM is a single number, one

dimensional, color scale ranging from a pale straw through to a deep red. The Lovibond RYBN and AOCS RY scales showed that the basic colors of distillates from Sample C were red and yellow. In addition, the densities of red and yellow increased with distillate recovery, causing the appearance of distillate is getting reddish brown as distillate recovery increased. Similarly, considering the FAC, Gardner and ASTM scales, it could be seen that the color of distillate became reddish as the recovery increased. Thus, brown become predominant in 80%-recovery, however, it was still reddish brown. The odor, however, could not be removed. All distillates still smell of smoky, thus, the distillates need further treatment for odor removal.

Table 2 : Color of d	stillates from Sample C
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Baseven	Lovibond RYBN			AOCS RY		FAC	Cordpor	ASTM	Visual		
Recovery	Red	Yellow	Blue	Neutral	Red	Yellow	FAC	Gardner	ASTIVI	VISUAI	
20%	2.4	39	0	0.1	2.3	32	11	10.8	2.0	Light yellow	
40%	3.0	46	0	0.1	2.9	40	11	11.2	2.1	Light yellow	
60%	3.3	57	0	0.1	3.3	50	11	11.6	2.3	Light yellow	
80%	10.9	70	0	0.1	9.9	70	33	14.7	3.9	Reddish brown	

2.

#### IV. Conclusion

A Simple vacuum distillation has been used to improve the quality of vetiver oils from smallholders. Vetiverol content, which is the difficult quality parameter to achieve could be improved. For all samples, the Indonesian standard on vetiverol content could be obeyed by distilling 60% samples. The better the sample, the less volume of initial oil that have to be sacrificed to achieve the standard vetiverol content. The standard vetiverol content could be achieved from samples A and B, which are better than sample C, until 80% recovery. Although all sample appeared to black and did not meet the standard appearance, standard in color could be achieve by all distillates. The distillate had appearance from yellow to reddish brown, with Gadner scale of color ranging from 10.8 to 14.7. Distillation, however, slightly disturbed the achievement of other parameters including density, acid number and ester number. Lower distillation fraction tend to shift the values of these parameters from their standards. The odor however could not be removed.

#### V. Acknowledgment

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## Chemical and Electrochemical Study on the Effectively of *Melilotus Officinalis* Extract as Save Corrosion Inhibitor for Aluminium in 1 M Hydrochloric Acid Solutions

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*Abstract*- Melilotus officinalis Extract (MOE), was investigated as a green corrosion inhibitor for aluminium in 1 M HCl solution using weight loss, hydrogen evolution, potentiodynamic polarization, electrochemical impedance spectroscopy (EIS) and electrochemical frequency modulation (EFM) techniques. Surface morphology was tested using scanning electron microscope (SEM). The effect of the temperature on corrosion behavior with addition of different concentrations was studied in the temperature range of 25-45 °C by weight loss method. Polarization curves reveal that the investigated extract is a mixed type inhibitor. The inhibition efficiency was found to increase with increase in the investigated extract concentration and increase with increase in solution temperature. The adsorption of the inhibitor on aluminium surface was found to obey the Temkin's adsorption isotherm. The activation and adsorption parameters were calculated and discussed. The results obtained from chemical and electrochemical techniques are in good agreement.

Keywords: acidic corrosion, aluminium, melilotus officinalis extract, EIS, EFM, SEM.

GJRE-C Classification: FOR Code: 290699

# CHEMICALANDELECTROCHEMICALSTUDYONTHEEFFECTIVELYOFMELILOTUSOFFICINALISEXTRACTASSAVECORROSIONINHIBITORFORALUMINIUMINIMHYDROCHLORICACIDSOLUTIONS

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# Chemical and Electrochemical Study on the Effectively of *Melilotus Officinalis* Extract as Save Corrosion Inhibitor for Aluminium in 1 M Hydrochloric Acid Solutions

Abd El-Aziz S. Fouda <sup> $\alpha$ </sup>, Rabab Abo-Shohba <sup> $\sigma$ </sup>, Walaa M. Husien <sup> $\rho$ </sup> & Esraa S. Ahmed <sup> $\omega$ </sup>

Abstract- Melilotus officinalis Extract (MOE), was investigated as a green corrosion inhibitor for aluminium in 1 M HCl solution using weight loss, hydrogen evolution, potentiodynamic polarization, electrochemical impedance spectroscopy (EIS) and electrochemical frequency modulation (EFM) techniques. Surface morphology was tested using scanning electron microscope (SEM). The effect of the temperature on corrosion behavior with addition of different concentrations was studied in the temperature range of 25-45 °C by weight loss method. Polarization curves reveal that the investigated extract is a mixed type inhibitor. The inhibition efficiency was found to increase with increase in the investigated extract concentration and increase with increase in solution temperature. The adsorption of the inhibitor on aluminium surface was found to obey the Temkin's adsorption isotherm. The activation and adsorption parameters were calculated and discussed. The results obtained from chemical and electrochemical techniques are in good agreement.

Keywords: acidic corrosion, aluminium, melilotus officinalis extract, EIS, EFM, SEM.

#### I. INTRODUCTION

orrosion is a fundamental process playing an important role in economics and safety, particularly for metals. The use of inhibitors is one of the most practical methods for protection against corrosion, especially in acidic media [1]. Most wellknown acid inhibitor are organic compounds containing nitrogen, sulfur, and oxygen atoms. Among them, organic inhibitors have many advantages such as high inhibition efficiency and easy production [2-5]. Organic heterocyclic compounds have been used for the corrosion inhibition of iron [6-9], copper [10], aluminum [11-13], and other metals [14-15] in different corroding media. Although many of these compounds have high inhibition efficiencies, several have undesirable side effects, even in very small concentrations, due to their toxicity to humans, deleterious environmental effects, and high-cost [16].

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Plant extract is low-cost and environmental safe, so the main advantages of using plant extracts as corrosion inhibitor are economic and safe environment. Up till now, many plant extracts have been used as effective corrosion inhibitors for aluminium in acidic media, such as: Garlic [17], Black Mulberry [18], Piper Guineense seed [19] Red onion skin [20]. The inhibition performance of plant extract is normally ascribed to the presence of complex organic species, including tannins. alkaloids and nitrogen bases, carbohydrates and proteins as well as hydrolysis products in their composition. These organic compounds usually contain polar functions with nitrogen, sulfur, or oxygen atoms and have triple or conjugated double bonds with aromatic rings in their molecular structures, which are the major adsorption centers.

Melilotus officinalis extract (MOE), belongs to the family Leguminosae (Fabaceae). It exhibits several medicinal properties, this plant is mainly used for agricultural purposes. It is grown as hay despite its toxic properties when moldy. It is considered an excellent green manure. Sweet clover is a major source of nectar for domestic honey bees. Flowers and seeds can be used as flavoring Melilotus officinalis has been used as a phytoremediation—phytodegradation plant for treatment of soils contaminated with dioxins [21].

The present work was designed to study the inhibitory action of Melilotus officinalis for the corrosion of aluminium in 1 M HCl using Chemical and electrochemical techniques, and to study the effect of temperature on the rate of corrosion

#### II. Experimental Methods

#### a) Materials and Solutions

Aluminium used has the chemical composition (% weight) 0.30 Si; 0.60 Fe; 0.10 Cu; 1.40 Mn; 0.05 Mg; 0.05 Cr; 0.05 Ti and the rest aluminium. The auxiliary electrode was a platinum wire (1 cm<sup>2</sup>), while a saturated calomel electrode (SCE) connected to a conventional electrolytic cell of capacity 100 ml via a bridge with a Luggin capillary, the tip of which was very close to the surface of the working electrode to minimize the IR drop. The aggressive solution used was prepared by dilution

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of analytical reagent grade 37% HCl with bidistilled water. The stock solution (1000 ppm) of Melilotus officinalis was used to prepare the desired concentrations by dilution with bidistilled water. The concentration range of MOE used was 50-300 ppm.

#### b) Preparation of plant extracts

Fresh aerial parts of MOE sample were crushed to make fine powder. The powdered materials (250 g) were soaked in 500 ml of dichloromethane for 5 days and then subjected to repeated extraction with  $5 \times 50$  ml until exhaustion of plant materials. The extracts obtained were then concentrated under reduced pressure using rotary evaporator at temperature below 50°C. The dichloromethane evaporated to give solid extract that was prepared for application as corrosion inhibitor. Chemical studies have demonstrated that the main chemical constituents of Melilotus officinalis are the glycosides of coumaric acid, especially melitoside, which by hydrolysis of lactonises gives coumarin. Free coumarin, 3,4-dihydroxycoumarin (melilotin), scopoletin and umbelliferone are also present [22].

#### c) Weight loss measurements

Seven parallel aluminium sheets of  $2 \times 2 \times 0.2$  cm were abraded with emery paper (grade 320–500–1200) and then washed with bidistilled water and acetone. After accurate weighing, the specimens were immersed in a 250 ml beaker, which contained 100 ml of HCl with and without addition of different concentrations of Melilotus officinalis. All the aggressive acid solutions were open to air. After 180 minutes, the specimens were taken out, washed, dried, and weighed accurately. The average weight loss of seven parallel aluminium sheets could be obtained. The inhibition efficiency (IE%) and the degree of surface coverage,  $\theta$  of MOE for the corrosion of Al were calculated as follows [23]:

$$IE\% = \theta \times 100 = \left[1 - \frac{W}{W^{\circ}}\right] \times 100 \tag{1}$$

where W<sup>o</sup> and W are the values of the average weight losses without and with addition of the inhibitor, respectively.

#### d) Gasometric measurements

The gasometric method assembly used for the measurement of hydrogen gas evolution from the corrosion reaction was designed following the method described by Onuchukwu [24]. The gasometric assembly measures the volume of hydrogen gas evolution from the reaction system. Sevenaluminium coupons of dimension 2 x 2 x 0.2cm were used in the experiments for test solutions containing 1 M HCl with the six different concentrations of MOE and the blank at 25°C. A 50ml of each test solution was introduced into the reaction vessel connected to a burette through a delivery tube. The initial volume of air in the burette was recorded. Thereafter, one aluminium coupon was dropped into the corrodent and the reaction vessel

quickly closed. Variation in the volume of hydrogen gas evolved with time was recorded every 1min. for 80 min. Each experiment was conducted on a fresh specimen of metal coupon. The hydrogen gas evolved displaced the paraffin water in the gasometric set-up and the displacement representing the volume of hydrogen evolved was read directly. The experiment was repeated in the presence of the six different concentrations of MOE, 50 to 300 ppm asused in the weight loss experiments.

#### e) Electrochemical measurements

Electrochemical measurements were performed using a typical three-compartments glass cell consisting of the aluminium specimen as working electrode (1 cm<sup>2</sup>), saturated calomel electrode (SCE) as a reference electrode, and a platinum wire as a counter electrode. The reference electrode was connected to a Luggin capillary and the tip of the Luggin capillary is made very close to the surface of the working electrode to minimize IR drop. All the measurements were done in solutions open to atmosphere under unstirred conditions. All potential values were reported versus SCE. Prior to each experiment, the electrode was abraded with successive different grades of emery paper, degreased with acetone, also washed with bidistilled water, and finally dried. Tafel polarization curves were obtained by changing the electrode potential automatically from (-0.8 to 1 V vs. SCE) at open circuit potential with a scan rate of 1 mVs<sup>-1</sup>. Stern-Geary method [25], used for the determination of corrosion current is performed by extrapolation of anodic and cathodic Tafel lines to a point which gives (log  $i_{\mbox{\scriptsize corr}})$  and the corresponding corrosion potential  $(E_{corr})$  for inhibitor free acid and for each concentration of inhibitor. Then (icorr) was used for calculation of inhibition efficiency (IE %) and surface coverage ( $\theta$ ) as in equation 2:

$$IE\% = \Theta \times 100 = \left[1 - \frac{i_{corr\ (in\ h)}}{i_{corr\ (free\ )}}\right] \times 100$$
(2)

Where  $i_{\text{corr(free)}}$  and  $i_{\text{corr(inh)}}$  are the corrosion current densities in the absence and presence of inhibitor, respectively.

Impedance measurements were carried out in frequency range (2x10<sup>4</sup> Hz to 8x10<sup>-2</sup> Hz) with amplitude of 5 mV peak-to-peak using AC signals at open circuit potential. The experimental impedance was analyzed and interpreted based on the equivalent circuit. The main parameters deduced from the analysis of Nyquist diagram are the charge transfer resistance R<sub>ct</sub> (diameter of high-frequency loop) and the double layer capacity C<sub>dl</sub>. The inhibition efficiencies and the surface coverage ( $\theta$ ) obtained from the impedance measurements are calculated from equation 3:

$$IE\% = \Theta \times 100 = \left[1 - \left(\frac{R_{ct}}{R_{ct}}\right)\right] \times 100$$
(3)

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Where  $R^{o}_{ct}$  and  $R_{ct}$  are the charge transfer resistance in the absence and presence of inhibitor, respectively.

Electrochemical frequency modulation, EFM, was carried out using two frequencies 2 and 5 Hz. The large peaks were used to calculate the corrosion current density ( $i_{corr}$ ), the Tafel slopes ( $\beta_a$  and  $\beta_c$ ) and the causality factors CF-2&CF-3 [26]. The electrode potential was allowed to stabilize 30 min before starting the measurements. All the experiments were conducted at 25°C.

All electrochemical measurements were performed using Gamry Instrument (PCI4/750) Potentiostat/ Galvanostat/ZRA. This includes a Gamry framework system based on the ESA 400. Gamry applications include DC105 software for potentiodynamic polarization, EIS 300 software for electrochemical impedance spectroscopy, and EFM 140 software for electrochemical frequency modulation measurements via computer for collecting data. Echem Analyst 6.03 software was used for plotting, graphing, and fitting data. To test the reliability and reproducibility of the measurements, duplicate experiments, which performed in each case at the same conditions.

#### f) Surface morphology

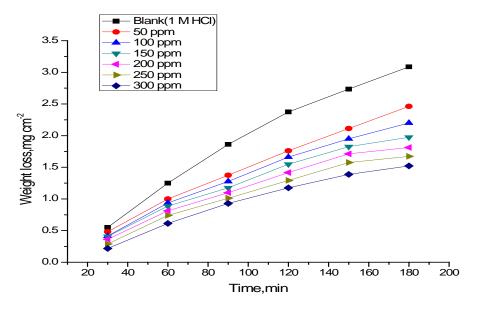
For morphological study, surface features (2 x 2 x 0.2cm) of aluminium were examined before and after

exposure to 1 M HCl solutions for 24 hour with and without extract. JEOL JSM-5500 scanning electron microscope was used for this investigation.

#### III. Results and Discussion

#### a) Weight loss measurements

Weight loss measurements were carried out for aluminium in 1 M HCl in the absence and presence of different concentrations of Melilotus officinalis and are shown in Figure (1). The inhibition efficiency (IE%) values calculated are listed in Table (1). From this table, it is noted that the IE% increases steadily with increasing the concentration of Melilotus officinalis and with increasing temperature from 25-45°C. The inhibition efficiency (IE%) and surface coverage  $(\theta)$ were calculated by equation (1). The observed inhibition action of the MOE could be attributed to the adsorption of its components on aluminium surface. The formed layer, of the adsorbed molecules, isolates the metal surface from the aggressive medium which limits the dissolution of the latter by blocking of their corrosion sites and hence decreasing the corrosion rate, with increasing efficiency as their concentrations increase [27].



*Figure 1 :* Weight loss-time curves for the corrosion of aluminium in 1 M HCl in the absence and presence of different concentrations of MOE at 25°C

#### b) Gasometric measurements

The volume of hydrogen evolved during the corrosion reaction of aluminium in 1 M HCl solutions devoid of and containing different concentrations of extract is measured as a function of the reaction time, and the data are represented graphically in Figure 2.

Inspection of the figure reveals that, the hydrogen evolution starts after a certain time from the immersion of aluminum coupon in the test solution. It may be expected thatthis time corresponds to the period needed by the acid to destruct the pre-immersion oxide film before the start of the metal attack, and it is known as the incubation period. Further inspection of Figure 2 reveals linear relationship between the time of reaction and the volume of hydrogen evolved, in all of the tested solutions. However, the presence of the extract decreases, markedly, the slope of the straight line. Since the slope of the line represents the corrosion reaction rate, it could be concluded that the *Melilotus offecinalis* 

extract has an excellent ability to inhibit the corrosion of aluminium in the acid solution. The values of inhibition efficiencies of different concentrations of the extract are given in Table 2. Inspection of Table 2 reveals that the IE increases as the concentration of the extract is increased.

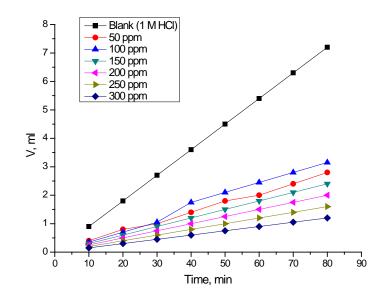
Table 1 : Variation of corrosion rate (k <sub>corr</sub> ), surface coverage(Θ)and inhibition efficiency(IE%) with different	
concentrations of MOE after 120 minutes of immersion in 1 M HCl at different temperatures.	

Temp. °C	[Inh] ppm	Weight loss, mg cm <sup>-2</sup>	k <sub>corr,</sub> mg cm <sup>-2</sup> min <sup>-1</sup>	θ	
	Blank	3.24	0.027		
	50	1.48	0.012	0.544	54.4
	100	1.46	0.012	0.546	54.6
25	150	1.41	0.011	0.560	56.0
	200	1.31	0.011	0.599	59.9
	250	1.30	0.011	0.595	59.5
	300	1.20	0.010	0.623	62.3
	Blank	7.44	0.062		
	50	2.55	0.021	0.657	65.7
	100	2.41	0.020	0.674	67.4
30	150	2.22	0.020	0.701	70.1
50	200	1.81	0.018	0.757	75.7
	250	1.73	0.014	0.767	76.7
	300	1.66	0.013	0.776	77.6
-	Blank	23.67	0.198		
	50	6.70	0.055	0.717	71.7
	100	6.49	0.054	0.726	72.6
35	150	6.10	0.050	0.744	74.4
	200	5.24	0.043	0.778	77.8
-	250	4.87	0.040	0.794	79.4
-	300	4.39	0.036	0.814	81.4
	Blank	34.92	0.291		
-	50	5.82	0.049	0.834	83.4
-	100	5.39	0.044	0.845	84.5
40	150	4.55	0.037	0.869	86.9
-	200	4.10	0.034	0.883	88.3
-	250	3.65	0.030	0.895	89.5
-	300	3.25	0.027	0.907	90.7
	Blank	51.96	0.433		
	50	8.10	0.068	0.845	84.5
	100	7.35	0.061	0.858	85.8
45	150	6.50	0.054	0.874	87.4
	200	5.57	0.046	0.893	89.3
	250	4.75	0.039	0.908	90.8
	300	3.80	0.032	0.926	92.6

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Table 2 : Inhibition efficiency obtained from hydrogen evolution method for Al after 80 minutes from immersion in 1 MHCl for various concentrations of MOEat 25°C

[Inh] Ppm	Volume of hydrogen gas evolvedml	IE %
Blank	6.3	-
50	2.7	56.8
100	2.4	61.9
150	2.1	66.6
200	1.8	72.2
250	1.4	77.7
300	11	83.3

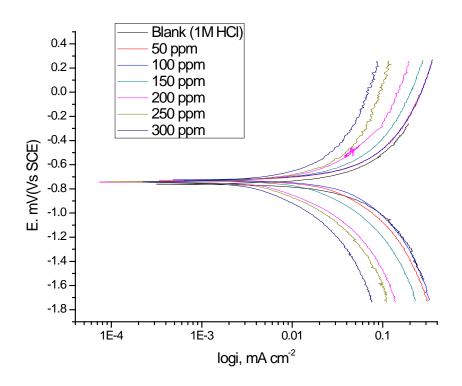


*Figure 2 :* volume of hydrogen gas evolved during the corrosion of aluminium in 1M HCl in the absence and presence of different concentrations of MOE

#### c) Polarization curves

Figure 3 shows potentiodynamic polarization curves recorded for aluminium in 1 M HCl solutions in the absence and presence of various concentrations of MOE at 25°C.

Lee and Nobe [28] reported the occurrence of a current peak between the apparent-Tafel and limitingcurrent regions during potential sweep experiments. The presence of MOE shifts both anodic and cathodic branches to the lower values of corrosion current densities and thus causes a remarkable decrease in the corrosion rate. The parameters derived from the polarization curves in Figure 3 are given in Table 3. In 1 M HCl solution, the presence of MOE causes a remarkable decrease in the corrosion rate i.e., shifts both anodic and cathodic curves to lower current densities. In other words, both cathodic and anodic reactions of aluminium electrode are retarded by MOE in 1 M HCl solution. The Tafel slopes of  $\beta_a$  and  $\beta_c$  at 25°Cdo not change remarkably upon addition of MOE, which indicates that the presence of MOE does not change the mechanism of hydrogen evolution and the metal dissolution process. Generally, an inhibitor can be classified as cathodic type if the shift of corrosion potential in the presence of the inhibitor is more than 85 mV with respect to that in the absence of the inhibitor [29, 30]. In the presence of MOE,  $E_{corr}$  shifts to less negative but this shift is very small (about 20-30 mV), which indicates that MOE can be arranged as mixed inhibitor.



*Figure 3 :* Potentiodynamic polarization curves for the corrosion of aluminium in 1 M HCl solution without and with various concentrations of MOE at 25°C

Table 3 : Corrosion potential ( $E_{corr}$ ), corrosion current density ( $i_{corr}$ ), Tafel slopes ( $\beta_c$ , $\beta_a$ ), degree of surface cove	rage
$(\theta)$ , and inhibition efficiency (IE%) of AI in 1M HCI at 25°C	

[Inh] ppm	-E <sub>corr</sub> mV vs SCE	i <sub>corr</sub> m A cm <sup>-2</sup>	<sup>β</sup> a mV dec <sup>-1</sup>	<sup>β</sup> c mV dec <sup>-1</sup>	C.Rx10 <sup>-3</sup> mpy	θ	IE%
0	725	275	250	420	164		
50	760	136	150	170	58	0.505	50.5
100	730	127	80	100	57	0.538	53.8
150	732	82	60	100	37	0.700	70.0
200	743	62	30	90	10	0.773	77.3
250	742	56	20	90	5.3	0.796	79.6
300	744	30	30	90	5.0	0.889	88.9

# d) Electrochemical impedance spectroscopy (EIS) measurements

Figure 4 shows impedance plots for aluminium M HClsolution without and with in 1 different impedance concentrations of MOE. The spectra consists of a Nyquist semicircle type without appearance of diffusive contribution to the total impedance (Z) indicating that the corrosion proceeds mainly under charge-transfer control and the presence of extract do not alter the mechanism of corrosion reaction.It is found that the obtained Nyquist plots are not perfect semicircle due to frequency dispersion and this behavior can be attributed to roughness and in homogeneities of the electrode surface [31, 32]. When there is non-ideal frequency response, it is common practice to use distributed circuit elements in an equivalent circuit. The most widely employed is the constant phase element (CPE). In general a CPE is used in a model in place of a capacitor to compensate for inhomogeneity in the system [33]. It was found that the diameters of the semicircle increases with increasing the concentration of the investigated extract. This indicates that the polarization resistance of the oxide layer increases with increasing the concentration of MOE and the depressed capacitive semicircle are often referred to the surface roughness and inhomogeneity, since this capacitive semicircle is correlated with dielectric properties and thickness of the barrier oxide film [34]. The data revealed that, each impedance diagram consists of a large capacitive loop with low frequencies dispersion (inductive arc). This inductive arc is generally attributed to anodic adsorbed intermediates controlling the anodic process [35-36]. By following this, inductive arc was disregarded. The electrical equivalent circuit model shown in Figure 5 was used to analyze the obtained impedance data. The model consists of the solution resistance ( $R_s$ ), the charge-transfer resistance of the interfacial corrosion reaction ( $R_{ct}$ ) and the constant phase angle element (CPE). The value of frequency power (n) of CPE can be assumed to correspond to capacitive behavior. However, excellent fit with this model was obtained with our experimental data. The admittance of CPE is described as:

$$Y_{CPE} = Y_o(j\omega)^n \tag{4}$$

where j is the imaginary root,  $\omega$  the angular frequency, Y<sub>o</sub> the magnitude and *n* the exponential term [37].

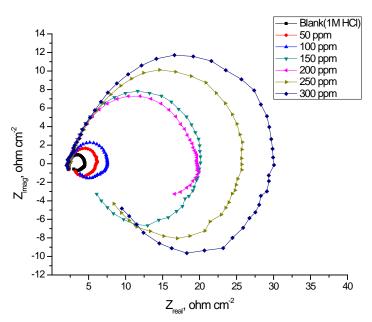
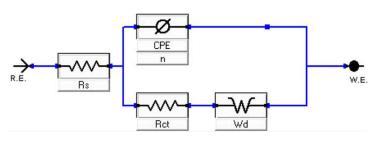


Figure 4 : Nyquist plots for aluminium in 1 M HCl solutions in the absence and presence of various concentrations of MOE at 25°C

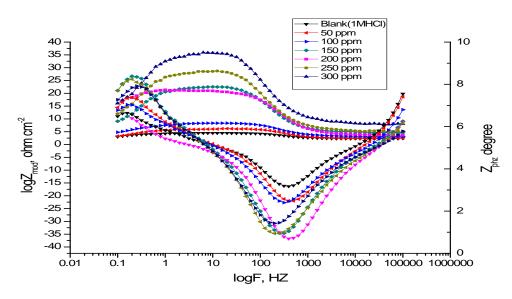


*Figure 5* : Equivalent circuit used to model impedance data for aluminium in 1 M HCl solutions

A long Warburg diffusion tail was observed at low frequency values. The tails are inclined at an angle of 45° to the real-axis at the very low frequencies; A diffusion controlled process is therefore exists. Studies reported in the literature [38] showed that the diffusion process is controlled by diffusion of dissolved oxygen from the bulk solution to the electrode surface and the Warburg impedance, which is observed in the low frequency regions, is ascribed to diffusion of oxygen to the alloy surface. This diffusion tail still appears, even in presence of high concentrations of the investigated extract. This means that the corrosion behavior of alloy in the absence as well as in the presence of MOE is influenced by mass transport.

Also, Bode plots for the aluminium in 1 M HCl solution are shown in Figure 6. In which the high frequency limit corresponding to the electrolyte

resistance (ohmic resistance)  $R_{\Omega}$ , while the low frequency represents the sum of  $(R_{\Omega} + R_{ct})$ , where  $R_{ct}$  is in the first approximation determined by both electrolytic conductance of the oxide film and the polarization resistance of the dissolution and repassivation process. At both low and high frequency limits, the phase angle between the current and potential ( $\theta$ ), assumes a value of about 0°, corresponding to the resistive behavior of  $R_{\Omega}$  and ( $R_{\Omega} + R_{ct}$ ).



*Figure 6 :* Bode plots for aluminium in 1 M HCl solutions in the absence and presence of various MOE concentrations at 25°C

The main parameters deduced from the analysis of Nyquist diagram are:

- The resistance of charge transfer R<sub>ct</sub>(diameter of high frequency loop)
- The capacity of double layer C<sub>dl</sub> which is defined as:

$$C_{\rm dl} = \frac{1}{2\pi R_{ct} f_{max}} \tag{5}$$

Where  $f_{max}$  is the maximum frequency at which the  $Z_{imag}$  of the impedance is a maximum. Since the electrochemical theory assumed that (1/R<sub>ct</sub>) is directly proportional to the capacity of double layer C<sub>dl</sub>, the inhibition efficiency (IE%) of the inhibitor for aluminium in 1 M HCl solution was calculated from R<sub>ct</sub> values obtained from impedance data at different inhibitor concentration the following equation:

$$IE\% = \left(1 - \frac{R_{ct}^{\circ}}{R_{ct}}\right) \times 100 \tag{6}$$

Where  $R^{\circ}_{ct}$  and  $R_{ct}$  are the charge transfer resistance in the absence and Presence of investigated extract, respectively. From the impedance data given in Table 4, we can conclude that the value of  $R_{ct}$  increases with the increase in the concentration of the investigated extract and this indicates the formation of a protective film on the Al surface by the adsorption and an increase in the corrosion inhibition efficiency in acidic solution. While the value of  $C_{dl}$  decreases with increasing the concentrations of extract in comparison with that of blank solution (uninhibited), as a result from the replacement of water molecules by inhibitor molecules which lead to increase in local dielectric constant and/or an increase in the thickness of the electric double layer formed on the metal surface [39,40].

Table 4 : Electrochemical kinetic parameters obtained from EIS technique for AI in 1M HCI in the absence and presence of different concentrations of investigated plant extracts at 25°C

[Inh] ppm	R <sub>p</sub> Ω Cm <sup>2</sup>	C <sub>d</sub> x10⁻⁵ µF Cm⁻²	θ	IE%
0	1.7	21		
50	2.9	5.9	0.468	46.8
100	3.2	4.4	0.595	59.5
150	4.2	3.2	0.730	73.0
200	6.3	3.0	0.760	76.0
250	7.1	2.5	0.840	84.0
300	10.9	2.3	0.844	84.4

# e) Electrochemical frequency modulation (EFM) measurements

EFM is a nondestructive corrosion measurement technique that can directly determine the corrosion current value without prior knowledge of Tafel slopes, and with only a small polarizing signal. These advantages of EFM technique make it an ideal candidate for online corrosion monitoring [41]. The great strength of the EFM is the causality factors which serve as an internal check on the validity of EFM measurement. The causality factors CF-2 and CF-3 are calculated from the frequency spectrum of the current

responses. Figure 7 show the frequency spectrum of the current response of pure Aluminium in 1 M HCI solution, contains not only the input frequencies, but also contains frequency components which are the sum, difference, and multiples of the two input frequencies. The EFM intermodulation spectrums of Aluminium in 1 M HCl solution containing (50ppm- 300ppm) of the MOE extract at 25°C is shown in Figure 7. The harmonic and intermodulation peaks are clearly visible and are much larger than the background noise. The two large peaks, with amplitude of about 200  $\mu$ A, are the response to the 40 and 100 mHz (2 and 5 Hz) excitation frequencies. It is important to note that between the peaks there is nearly no current response (<100 mA). The experimental EFM data were treated using two different models: complete diffusion control of the cathodic reaction and the "activation" model. For the latter, a set of three nonlinear equations had been solved, assuming that the corrosion potential does not change due to the polarization of the working electrode [42]. The larger peaks were used to calculate the corrosion current density ( $i_{corr}$ ), the Tafel slopes ( $\beta_c$  and  $\beta_a$ ) and the causality factors (CF-2 and CF-3). These electrochemical parameters were simultaneously determined by Gamry EFM 140 software, and listed in Table 5 indicating that this extract inhibit the corrosion of aluminium in 1 M HCI through adsorption. The causality factors obtained under different experimental conditions are approximately equal to the theoretical values (2 and 3) indicating that the measured data are verified and of good quality [43]. The inhibition efficiencies IE<sub>FEM</sub> % increase by increasing the studied extract concentrations and was calculated as follows:

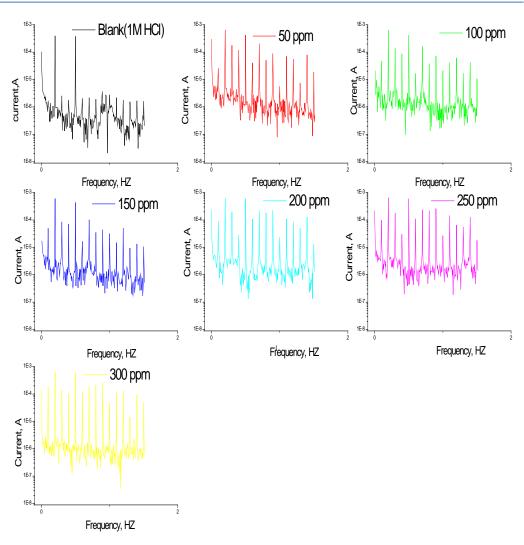
$$IE \%_{EFM = \left(1 - \frac{i_{corr}}{i_{corr}^{*}}\right) \times 100}$$
(7)

Where  $i^{o}_{\text{corr}}$  and  $i_{\text{corr}}$  are corrosion current densities in the absence and presence of MOE extract, respectively.

Table 5 : Electrochemical kinetic parameters obtained from EFM technique for aluminium in 1M HCl in the absence
and presence of different concentrations of MOE

[Inh] ppm	i <sub>corr</sub> m A cm <sup>-2</sup>	<sup>B</sup> a mV dec <sup>-1</sup>	<sup>β</sup> c mV dec <sup>-1</sup>	CF-2	CF-3	CRx10 <sup>-3</sup> mpy	θ	IE%
0	1100	182	195	1.1	2.3	667		
50	400.9	32	102	2.0	2.7	238	0.642	64.2
100	348.8	31	66	1.1	2.9	207	0.688	68.8
150	345.7	34	55	1.7	2.2	204	0.691	69.1
200	342.8	24	36	1.8	2.5	146	0.694	69.4
250	235.5	19	33	1.2	2.3	140	0.789	78.9
300	214.3	18	25	2.0	2.3	127	0.809	80.9

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*Figure 6 :* Intermodulation spectrums for the corrosion of aluminium in 1 M HCl without and with various concentrations of MOE at 25°C.

#### f) Adsorption isotherms

The mode and interaction degree between an inhibitor and a metallic surface have been widely studied with the application of adsorption isotherms. The adsorption of an organic molecule occurs because the interaction energy between an inhibitor and a metallic surface is higher than that between water molecules and metallic surface [44, 45]. To obtain the adsorption isotherms, the degree of surface coverage ( $\theta$ ) obtained from weight loss method was determined as a function of inhibitor concentration. The values of  $\theta$  were then plotted to fit the most suitable model of adsorption [46]. Attempts were made tofit experimental data to various isotherms including Frumkin, Langmuir, Temkin, Freundlich, isotherms. By far the results were besfitted by Temkin adsorption isotherm as seen in Figure 7 [47].

$$a/2.303\Theta = \log K_{ads} + \log C$$
 (9)

The equilibrium constant of adsorption  $K_{\mbox{\scriptsize ads}}$  obtained from the intercepts of Temkin adsorption

The equilibrium constant of adsorption  $K_{ads}$  obtained from the intercepts of Temkin adsorption isotherm is related to the free energy of adsorption  $\Delta G^\circ$  ads as follows:

$$K_{ads} = 1/55.5 \exp[(-\Delta G^{\circ}_{ads})/RT]$$
(8)

where 55.5 is the molar concentration of water in the solution in  $M^{-1}$ . The values obtained are given in Table 6.

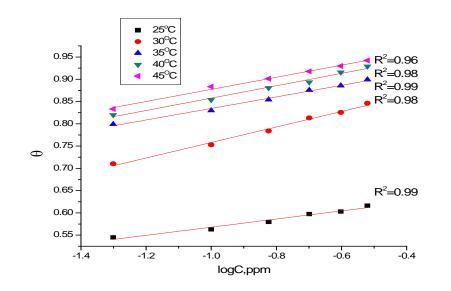


Figure 7 : Temkin adsorption plots for aluminium in 1 M HCl containing various concentrations of MOE at 25°C

Plot of  $(\Delta G^{\circ}_{ads})$  versus T (Figure 8) gave the heat of adsorption  $(\Delta H^{\circ}_{ads})$  and the entropy  $(\Delta S^{\circ}_{ads})$  according to the thermodynamic basic equation 5:

 $\Delta G^{\circ}_{ads} = \Delta H^{\circ}_{ads} - T \Delta S^{\circ}_{ads}(5)$ 

Table 6 clearly shows a good dependence of  $\Delta G^{o}_{ads}$  on T, indicating the good correlation among thermodynamic parameters. The negative value of  $\Delta G^{o}_{ads}$  reflect that the adsorption of studied inhibitors on aluminium surface from 1 M HCl solution is spontaneous process and stability of the adsorbed layer on the aluminium surface. Generally, values of  $\Delta G^{o}_{ads}$  around -20 kJ mol<sup>-1</sup> or lower are consistent with the electrostatic interaction between the charged molecules and the charged metal (physical adsorption); those around -40 kJ mol<sup>-1</sup> or higher involves charge sharing or transfer from organic molecules to the metal surface to form a coordinate type of bond (chemisorption) [48].

From the obtained values of  $\Delta {\rm G^{\circ}}_{\rm ads}$  it was found the existence of chemical adsorption). The values of thermodynamic parameter for the adsorption of inhibitors Table 6 can provide valuable information about the mechanism of corrosion inhibition. Endothermic adsorption process ( $\Delta H^{o}_{ads} >$ is 0) attributed unequivocally to chemisorption [49], an exothermic adsorption process ( $\Delta H^{o}_{ads} < 0$ ) may involve either physisorption or chemisorption or mixture of both processes. In the presented case, the calculated values of  $\Delta H^{o}_{\mbox{ ads}}$  for the adsorption of extract in 1 M HCl indicating that this extract may be chemically adsorbed. The values of  $\Delta S^{\circ}_{ads}$  in the presence of extract is large and positive that is accompanied with endothermic adsorption process. This indicates that decrease in disorder takes places on going from reactants to the metal-adsorbed reaction complex [50].

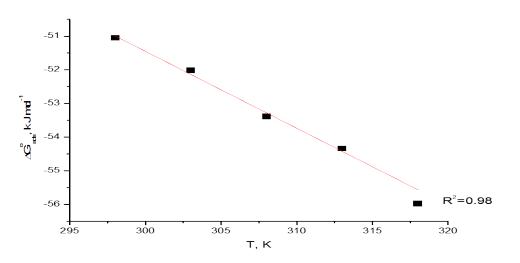


Figure 8 : Variation of  $\Delta G^{\circ}_{ads}$  versus T for the adsorption of MOE on aluminium surface in 1 M HCl at different temperatures

Table 6: Thermodynamic parameters for the adsorption of MOE on aluminium in 1MHCl at different	emperatures.
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Temp. ℃	K <sub>ads</sub> x10⁻ <sup>6</sup> M⁻¹	-∆G° <sub>ads</sub> k J mol⁻¹	-ΔH° <sub>ads</sub> k J mol <sup>-1</sup>	∆Sº <sub>ads</sub> J mol⁻¹ k⁻¹
25	12.2	50.4		226.7
30	15.6	51.8		227.8
35	20.6	53.4	17.2	229.2
40	26.9	54.9		230.5
45	33.9	56.5		231.6

#### g) Kinetic-thermodynamic corrosion parameters

Weight loss method was carried out at different temperature ( $25^{\circ}C-45^{\circ}C$ ) in the presence of different concentration of MOE. It has been found that he corrosion rate decreases with the increase in temperature for MOE (Table 1). The corrosion rate of aluminium in the absence of MOE increased steeply from 25 to  $45^{\circ}C$  whereas; in the presence of MOE the corrosion rate decreased slowly. The inhibition efficiency was found to increase with temperature. The corrosion parameter in the absence and presence of extract in the temperature range  $25-45^{\circ}C$  has been summarized in (Table 1).The apparent activation energy ( $E_{a}^{*}$ ) for dissolution of aluminium in 1 M HCl was calculated from the slope of plots by using Arrhenius equation:

$$\log k = \frac{-E_a^*}{2.303 \, R \, T} + \log A \tag{9}$$

where k is rate of corrosion,  $E_a^*$  is the apparent activation energy, R is the universal gas constant, T is absolute temperature and A is the Arrhenius pre-exponential factor.

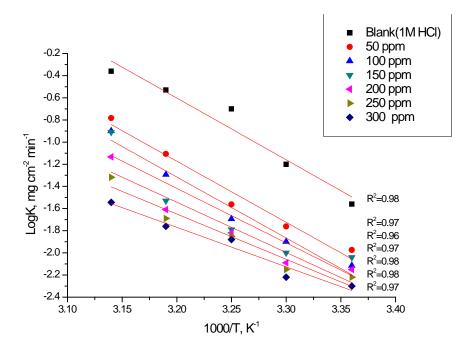
By plotting log k against 1/T the values of activation energy  $(E_a^*)$  has been calculated  $(E_a^*)$ (slope) 2.303 x R) (Figure 9). Activation energy for the reaction of aluminium in 1M HCl decreases in the presence of extract (Table 7). This decreas indicates the formation of chemical bonds were strengthen by increasing the temperature. However, the extent of the rate increment in the inhibited solution is lower than that in the free acid solution. Therefore, the inhibition efficiency of the MOE increases markedly with increasing temperature. This result supports the idea that the adsorption of extract components on the aluminium surface may be chemical in nature. Thus, as the temperature increases the number of adsorbed molecules increases leading to an increase in the inhibition efficiency. This could be done by adsorption on the aluminium surface making a barrier for mass and charge transfer. However, such types of inhibitors perform a good inhibition at high temperature with considerable increase in inhibition efficiency at elevated temperatures [51]. Moreover, the relatively high value of activation energy in presence of MOE suggests a chemical adsorption process.

The values of change of entropy ( $\Delta S^*$ ) and change of enthalpy ( $\Delta H^*$ ) can be calculated by using the formula:

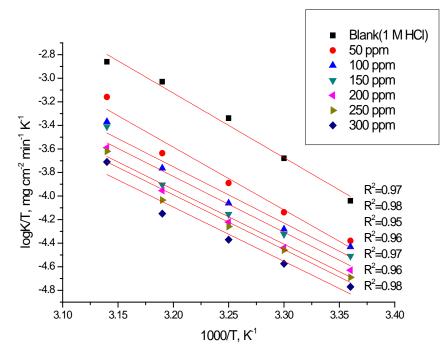
$$k = \left(\frac{RT}{Nh}\right) exp\left(\frac{\Delta S^*}{R}\right) exp\left(\frac{\Delta H^*}{RT}\right)$$
(10)

where k is rate of corrosion, h is Planck's constant, N is Avogadro number,  $\Delta S^*$  is the entropy of activation, and  $\Delta H^*$  is the enthalpy of activation. A plot of log (k/T) vs. 1/T (Figure 10) should give a straight line, with a slope of ( $\Delta H^*/2.303R$ ) and an intercept of [log (R/Nh)+ $\Delta S^*/$ 2.303R], from which the values of  $\Delta S^*$  and  $\Delta H^*$  can be calculated (Table 7). The positive value of  $\Delta S^*$  for the extract indicates that activated complex in the rate determining step represents a dissociation rather than an association step, meaning that an increase in disorder takes place during the course of transition from reactant to the activated complex [52] The positive sign of  $\Delta H^*$  indicates that the adsorption of extract molecules is an endothermic process. Generally, an endothermic process signifieschemisorption process.

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*Figure* 9 : log k (corrosion rate) – 1/T curves for aluminium in 1 M HCl in the absence and presence of different concentrations of MOE



*Figure 10 :* log k (corrosion rate) /T – 1/T curves for aluminium in 1 M HCl in the absence and presence of different concentrations of MOE

Table 7: Activation parameters for dissolution of aluminium in the absence and presence of different concentrations of MOE in 1 M HCI

Conc. ppm	Ea <sup>*</sup> , kJ mol⁻1	ΔH <sup>*</sup> , kJ mol <sup>-1</sup>	∆S <sup>*</sup> , J mol <sup>-1</sup> K <sup>-1</sup>
1 M HCI	106.6	45.5	78.1
50	105.5	44.3	60.1
100	105.3	39.7	23.2
150	94.2	39.4	19.4
200	86.7	38.6	11.7
250	78.1	38.5	10.7
300	68.1	38.2	6.1

*h*) Surface analysis by scanning electron microscopy

Figure11shows an SEM photograph recorded for aluminium samp-les polished (a) and exposed for 24 h in 1 M HCl solution without (b) and with (c) 300 ppm of MOE at 25C<sup>0</sup>. A photograph of the polished aluminium surface before immersion in 1 M HCl solution is shown in Figure 11a. The photograph shows the surface was smooth and without pits. The SEM micrographs of the corroded aluminium in the presence of 1 M HCl solution are shown in Figure 11b. The faceting seen in this figures was a result of pits formed due to the exposure of aluminium to the acid. The influence of the inhibitor addition 300 ppm on the aluminium in 1 M HCl solution

is shown in Figure 11c. The morphology in Figure 11c shows a rough surface, characteristic of uniform corrosion of aluminium in acid, as previously reported [52], that corrosion does not occur in presence of inhibitor and hence corrosion was inhibited strongly when the inhibitor was present in the hydrochloric acid, and the surface layer is very rough. In contrast, in the presence of 300 ppm of MOE, there is much less damage on the aluminium surface, which further confirm the inhibition action. Also, there is an adsorbed film adsorbed on aluminium surface (Figure 11c). In accordance, it might be concluded that the adsorption film can efficiently inhibits the corrosion of aluminium.

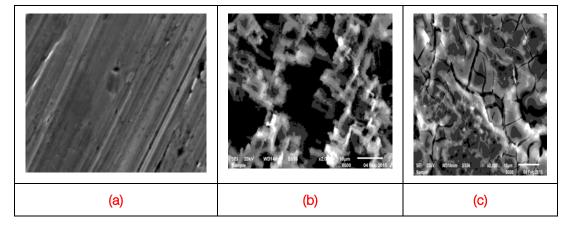


Figure 11: SEM micrographs of aluminium surface (a) before of immersion in 1 M HCl, (b) after 24 h of immersion in 1 M HCl and (c) after 24 h of immersion in 1 M HCl + 300 ppm of MOE at 25°C

#### Mechanism of the corrosion inhibition i)

The adsorption of organic compounds can be described by two main types of interactions: physical and chemisorptions adsorption. In general, physical adsorption requires the presence of both the electrically charged surface of the metal and charged species in solution. The surface charge of the metal is due to the electric field existing at the metal/solution interface. A chemisorption process, on the other hand, involves charge sharing or charge transfer from the inhibitor molecules to the metal surface to form a coordinate type

of a bond. This is possible in case of a positive as well as a negative charge of the surface. The presence of a transition metal, having vacant, low-energy electron orbitals and an inhibitor with molecules having relatively loosely bound electrons or heteroatoms with a lone pair of electrons is necessary for the inhibiting action [53]. Generally, two types of mechanisms of inhibition were proposed. One was the formation of polymeric complexes with aluminium ions (Al<sup>3+</sup>) depending on the applied conditions[54, 55]. The other was the chemical adsorption of MOE components on aluminium surface

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[56, 57]. The inhibition action of MOE does not occur by the simple blocking at the surface of aluminium, especially at high temperature. This might be attributed to the different adsorption capacities of the MOE extract on the aluminium surface at different temperatures. It has been studied that with the increase in temperature, the desorption effect of MOE on aluminium surface increased. Some of the hydrophilic groups with positively charged atoms (O<sup>+</sup>) desorbed from the surface of aluminium and did more work to prevent the H<sup>+</sup> from getting nearer to the metal surface. Therefore, MOE preferentially inhibited both cathodic and anodic corrosion processes at high temperature.

#### IV. CONCLUSIONS

From the overall experimental results the following conclusions can be deduced:

- 1. The MOE shows good performance as corrosion inhibitor in 1 M HCl.
- 2. The results obtained from weight loss showed that the inhibiting action increases with the MOE concentration and also increase with the increasing in temperature.
- 3. Double layer capacitances decrease with respect to blank solution when the plant extract is added. This fact confirms the adsorption of plant extract molecules on the aluminium surface.
- 4. The MOE inhibits the corrosion by getting adsorbed on the metal surface following Temkin adsorption isotherm.
- 5. The inhibition efficiencies determined by weight loss, potentiodynamic polarization and EIS techniques are in reasonably good agreement.

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## Influence of Scan Rate on Simulation of Differential Scanning Calorimetry Profiles of Protein Denaturationt

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Keywords: carbonic anhydrase, melting temperature, molecular dynamics simulation, scan rate, simulated annealing.

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# Influence of Scan Rate on Simulation of Differential Scanning Calorimetry Profiles of Protein Denaturationt

Maryam Ghadamgahi <sup>a</sup> & Davood Ajloo <sup>o</sup>

Abstract- The heat capacity has played a major role in proteins. Its calculation by atomistic simulation methods remains a significant challenge due to the complex and dynamic nature of protein structures and this work compares the denaturation effect of bovine carbonic anhydrase (BCA) by heat, pH and scan rate dependence of protein denaturation by molecular dynamics (MD) simulation. To better understand this factor on calculating a protein heat capacity and  $T_m$ , we have provided a comparative analysis of simulation models that differ in their scan rate and pH description. Our model protein system is the carbonic anhydrase, and a series of 20 ns simulated DSC with different scan rate (v= 0.10, 0.0125, 0.015 and 0.02 K/ps) and pH have been reported by simulated annealing performed at temperatures ranging from 250 to 575 K, starting from the carbonic anhydrase native structure. It was observed that, our systems were guite sensitive to the description and the calculated melting temperature ( $T_m$ ) varied in the range 353-438 K and was higher for higher scan rates systems and lower for acidic condition. It was also demonstrated that increasing scan rate causes a slight shift to right and acidic pH cause a shift to left in  $T_m$  value.

Keywords: carbonic anhydrase, melting temperature, molecular dynamics simulation, scan rate, simulated annealing.

#### I. INTRODUCTION

arbonic anhydrase (CA) is a clinically relevant and biochemically well-characterized protein. It catalyzes hydration of carbon dioxide to carbonic acid and is involved in vital physiological processes such as pH and  $CO_2$  homeostasis, transport of bicarbonate and  $CO_2$ , biosynthetic reactions, bone resorption, calcification, tumorigenicity, and other physiological or pathological processes. Therefore, this enzyme is an important target for inhibitors with clinical applications, primarily for use as antiglaucoma agents but also for the therapy of various pathologies such as epilepsy and Parkinson's disease. Many groups have used carbonic anhydrase—both bovine and human—as a model protein for studies of folding and unfolding.<sup>1-3</sup>

Differential scanning calorimetry (DSC) is a technique able to study thermally induced transitions and particularly, the conformational transitions of biological macromolecules (for example between the folded and the unfolded structure of a protein).

It measures the excess heat capacity of a solution  $(C_{\rm p})$ of the molecule of interest as a function of temperature and has been extensively used to study protein thermal denaturation.<sup>4</sup> A variety of techniques have evolved which can be used to gain structural information on protein stability. DSC has become one of the key physicochemical methods to study the stability of protein biopharmaceuticals.<sup>5,6</sup> In experimental study of carbonic anhydrase unfolding by DSC the enthalpy of unfolding in the temperature range of 39 to 72 °C by carrying out DSC experiments at various pH was determined.<sup>7</sup>  $T_m$  (effectively the transition peak) is defined as the temperature at which 50% of the protein molecules are unfolded or as a midpoint in a thermal ramp and represents a temperature where the free energy of the natives and nonnative forms are equivalent. Protein melting temperatures can be determined by numerous methods, including differential scanning calorimetry and optical methods (circular dichroism, fluorescence or absorbance spectroscopy). These techniques have low throughput, are time consuming, and require significant amounts of protein and, thus, are not generally utilized when testing the large numbers of compounds generated during drug development.

Scan rate dependence was determined using the methods described by Sanchez-Ruiz et al.<sup>8</sup> The scan-rate-dependent shift in  $T_m$  for denaturation was fitted to the equation:

$$\frac{scan\ rate}{T_m^2} = \frac{AR}{E_a} e^{-E_a/RT_m}$$
(1)

Such that a plot of  $\ln(\operatorname{scan rate}/T_m^2)$  against  $1/T_m$  yields a slope-  $E_a/R$ , where  $E_a$  is the activation energy for denaturation, R the gas constant and A the pre-exponential factor in the Arrhenius equation. The effects of scan rate on the DSC profiles are given. These results are expressed in the form of DSC profiles (excess  $C_{pr}$  vs. T).

The calorimetric transitions for carbonic anhydrase denaturation are highly scanning-rate dependent. which indicates that the thermal denaturation is under kinetic control.9-12 All above results are confirmed by a MD simulation study and a good agreement was found for the DSC data with experimental values.

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Finally, according to the two-state irreversible model, the scanning rate effect on the DSC transitions is given by:<sup>13</sup>

$$ln\left(\frac{v}{T_m^2}\right) = const - \frac{E}{RT_m}$$
(2)

Scan rate and pH dependence of carbonic anhydrase stability were determined by measuring the  $T_m$  values at various scan rates and pH to generate a DSC curve. Models relating the scan rate-dependent increase in protein thermal stability to association constants require an accurate knowledge of the thermodynamics of protein stability. Thus, carbonic anhydrase stability was studied by MD simulation of differential scanning calorimetry (DSC), giving a complete thermodynamic description of the Gibbs free energy, calorimetric enthalpy, and heat capacity of unfolding .<sup>14-20</sup>

We chose to conduct the study on carbonic anhydrase—a protein commonly used as a model for biophysical and physical-organic studies by us and others.<sup>21-24</sup>

Thermal denaturations of carbonic anhydrase have been examined using simulation of differential scanning calorimetry by molecular dynamic simulation. Thermal denaturation has never been directly examined previously theoretically. Carbonic anhydrase have been examined as a function of scan rate. In this work we have found denaturation of protein. Finally, it is noteworthy that CA could become important biotechnological materials. Therefore investigation of thermal stability of the CA has not only academic, but also applied, interest. This model correctly predicts scan rate and pH-dependent changes in  $T_m$  of BCA. These  $T_m$ values are then compared to those obtained by experimental methods that are in good agreement.

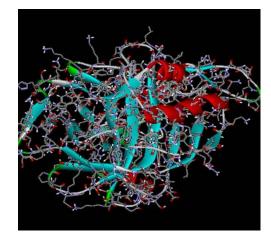
#### II. EXPERIMENTAL

#### a) Molecular dynamics simulations

All MD simulations were carried out using the GROMACS 4.5.0 package together with the GROMOS96 force field in parallel by the BirgHPC. The starting structure of bovine carbonic anhydrase was constructed based on the X-ray crystal structure of BCA (PDB ID: 1CA2, Fig. 1). The simple point charge (SPC) model was used to describe water. A different time step was used to integrate the equations of motion with the Verlet algorithm. A non bond pair list cutoff of 0.9 nm was used. Temperatures and pressures were controlled by a Nose-Hoover thermostat and Parrinello-Rahman barostate with coupling constants of 0.1 and 0.5, respectively. For all simulations, the atomic coordinates were saved every 50 ps for analysis. A cubic simulation box of the volume 321 nm<sup>3</sup> was made and then water molecules were randomly added into the simulation box and initial configurations were minimized using steepest descent algorithm with 5000 integration step. BirgHPC (Bioinformatics Research Group High Performance

Computing) which is a free Linux Live CD distribution based on Pelican HPC and Debian Live including 56 processors was also used for our simulations. BirgHPC has been developed to create high-performance clusters for bioinformatics and molecular dynamics studies using any Local Area Network (LAN)-networked computers. The latest versions of GROMACS 4.5.0 was run in parallel by the birgHPC.

Variations of temperature were adjusted in mdp file. In order to study thermal denaturation, temperatures varied in the range of 273 to 405 K to calculate the stability of the protein.



*Fig. 1 :* Protein structure (1CA2) taken from protein data bank www.RCSB.org

Simulated annealing is a special case of MD or MC simulation, in which the temperature is gradually reduced during the simulation. Often, the system is first heated and then cooled. Thus, the system is given the opportunity to surmount energetic barriers in a search for conformations with energies lower than the local-minimum energy found by energy minimization. One of the applications of MD is involved in utilization of MD, often with simulated annealing protocols, to determine or refine structures with data obtained from experiments.<sup>25</sup>

Creation of different scan rates by variation of temperature in each step was adjusted in mdp file by simulated annealing. DSC calculations were performed, keeping a constant pressure of 1 atm over the simulation. Different scanning rates within the range 0.010-0.02 K/ps were employed. In order to simulate scan rate of 0.01 K/ps, temperature increased from 250 to 450 K during 20 ns. It means that in each 500 step temperature increased 5 K. For the scan rate of 0.0125, initial and final temperature was 225 to 475 and in each 2000 ps step temperature increased 25 K, for scan rate of 0.015, initial and final temperature was 270 to 570 and in each 1000 ps step temperature increased 15 K and for scan rate of 0.02, initial and final temperature was 150 to 545 K and in each 250Ps step, temperature increased 5 K.

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In order to simulate lower pH (acidic form), all carboxyl groups (COO-) were protonated and converted to COOH by definite tools implemented in GROMACS. In addition, at neutral pH, we used the crystal structure (pdb code; 1CA2) downloaded from the protein data bank. All MD simulations for comparison of pH effect were carried out for three scan rates of 0.01, 0.0125 and 0.015 K/ps during 20 ns. Variations of RMSD, CD<sub>222,nm</sub>, hydrogen bond (HB), solvent accessible surface (sas) area, radius of gyration and Hamiltonian energy were also calculated.

#### b) Analyses

The conformational changes of the protein during MD simulations were monitored by the rootmean-square derivations (RMSD) with its X-ray structure as a reference. The RMSD value, a measure of molecular mobility, is calculated by translating and rotating the coordinates of the instantaneous structure to superimpose the reference structure with a maximum overlap. The RMSD is defined as

$$RMSD = \sqrt{\frac{\sum_{i=1}^{N} m_i (r_i - r_i^0)}{\sum_{i=1}^{N} m_i}}$$
(3)

Where  $m_i$  is the mass of atom *i*.  $r_i$  and  $r_i^{\circ}$  are the coordinates of atom *i* at a certain instance during MD simulations and at its reference state, respectively. RMSDs were calculated, for the trajectories, from the starting structures of carbonic anhydrase as a function of time. In the all systems, RMSDs reach a stable value within the first nanosecond of all the analyses.

The simulation trajectories were analyzed using several auxiliary programs provided with the GROMACS package. The programs include g\_energy that calculate all energies such as Hamiltonian, total pressure, box volume etc. and displays averages. Calculation of the heat capacity at constant pressure ( $C_P$ ) can be used to directly compare with experimental DSC results. In general, a straightforward but difficult method to accomplish this is to use the trajectory energy fluctuations to determine the  $C_P$ , directly. From a trajectory, one can determine the trajectory average energy and the enthalpy of each step *i* H<sub>I</sub>, to determine the heat capacity:

$$H = E + PV \tag{4}$$

$$C_{\rm P} = \frac{\langle {\rm H}^2 \rangle - \langle {\rm H} \rangle^2}{{\rm RT}^2} \tag{5}$$

$$\langle H \rangle = \frac{\sum_{V} \sum_{j} H_{i} e^{-E_{Vj} / kT} e^{-pV / kT}}{\sum_{V} \sum_{j} e^{-E_{Vj} / kT} e^{-pV / kT}}$$
(6)

 $\langle H \rangle$  is the average value of enthalpy,  $H_i$  is enthalpy of ith state and k is bultzman constant. It is

important to note that in all calculations of DSC profiles, heat capacity has been subtracted from solvent heat capacity and it has been done for all scan rates calculation.<sup>26-30</sup>

#### III. Results and Discussion

#### a) Thermal denaturation (Tm of carbonic anhydrase)

We used the MD simulation of differential scanning calorimetry (DSC) to monitor the thermal unfolding of BCA. The melting temperature of BCA in temperature range from 273 to 405 K was about 345 K which is near the experimental value.7, 31-32 Fig. 2 shows DSC profile or representative thermo gram in several temperatures for unfolding of CA protein. Thermodynamic of CA unfolding was studied by DSC using various temperature conditions previously demonstrated to give  $T_m$  using MD simulation. This observation was consistent with experiments, where the  $T_{\rm m}$  for BCA was estimated to be 343 K.

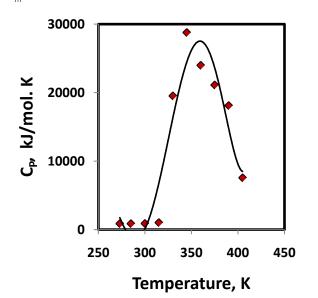


Fig 2 : DSC profile for the unfolding of BCA protein

Here, we describe simulations of CA at various temperatures, focusing on the unfolding process. Increasing temperature accelerates protein unfolding without changing the pathway of unfolding. Temperature is believed to alter the structure of hydrogen bonds network of protein in water and increase the SAS and protein size and decrease the intermolecular hydrogen bond, electrostatic and hyrophobic interactions of proteins. Structure parameters were obtained from MD simulation for each temperature and results were averaged. The structure information such as solvent accessible surface, inter molecular hydrogen bonding (HB) between CA and solvent molecules, gyrate radii (Rg), CD<sub>222 nm</sub> and RMSD were obtained and averaged at each temperature. Fig. 3 show parameters of total SAS, hydrophobic SAS, radius of gyration and RMSD

that increase by temperature due to unfolding of protein while intermolecular hydrogen bond, hydrophilic SAS and CD<sub>222,nm</sub> decrease with temperature.

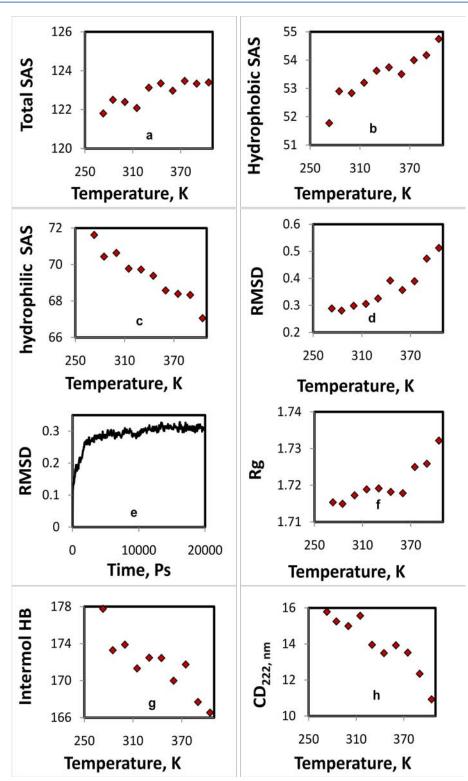
The variation of surface area during 20 ns time evolution was significant and obtained. Fig. 3a shows the averaged value of total solvent accessible surface area of CA in 20 ns time interval in the temperature ranged from 273 to 405. These figure shows increase of surface by increasing temperature. This proves that the CA structure has been unfolded and it is obvious that surface area of CA in system with 273 K is less than higher temperatures. Fig. 3b and 3c show average solvent accessible surface area of hydrophobic and hydrophilic part for CA in 20 ns time interval vs. temperature respectively. Solvent accessible surface area of hydrophobic part and total surface area of CA is more in the presence of higher temperatures. This proves that the CA structure has been unfolded more in the presence of higher temperature so the surface areas of protein increase due to unfolding process. Totally temperature cause more interaction and structural change in CA and this result is in good agreement with experiment data.

Root mean square deviation (RMSD) of the CA for all temperatures was obtained. Fig. 3d shows the average of CA RMSD in the 20 ns time interval in all temperatures. The figure shows that CA has more structural changes (RMSD) in the higher temperature. Fig. 3e shows the RMSD of CA in the 20 ns time interval for 273 K which has been selected randomly. It shows that the system reaches a stable state after about 5 ns.

Fig. 3f shows the average values of radius gyration of CA in 20 ns time interval vs. temperature. This figure shows increase of radius gyration of CA in the presence of higher temperature. This result is in good accordance with increase of hydrophobic and total surface area of CA. This proves that the CA structure has been unfolded and it is obvious that surface area and therefore radius gyration of CA in system with 405 K temperature is increased more due to more increase of CA surface area. This proves that the CA structure has been unfolded more in the higher temperatures.

Fig. 3g shows averaged value of intermolecular hydrogen bond of CA in 20 ns time interval vs. temperature. Reduction of this parameter proves that the CA structure has been unfolded in high temperatures and this result is in good agreement with above results. The number of intermolecular hydrogen bonds decreased in the process of increasing temperatures and protein structure is unfolded and denatured. The number of hydrogen bonds between protein-solvent is also increased due to unfolding of protein in high temperatures and is not shown here. In some of the temperatures these trends are reverse due to formation of helix and beta sheets. Decrease of SAS may be due to formation of beta sheet and helix structures. Fig. 3h shows averaged  $CD_{222,nm}$  of CA by molecular dynamics vs. temperature this figure shows the helicity as  $CD_{222,nm}$  which decreases in the high temperatures due to unfolding and denaturation of protein.

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*Fig. 3*: a) averaged total solvent accessible surface area of CA by molecular dynamics b) and c) averaged solvent accessible surface area of hydrophobic and hydrophilic part of CA by molecular dynamics, respectively d) averaged RMSD by molecular dynamics for CA e) Calculated RMSD by molecular dynamics for CA in the 273 K f) averaged radius gyration of CA by molecular dynamics g) averaged inter molecular hydrogen bond of CA by molecular dynamics h) averaged CD222,nm of CA by molecular dynamics vs.temperature

#### b) Scan rate dependence

In the case of the thermal denaturation of CA, however, the DSC transitions are strongly scanning-rate dependent. This indicates that the state of the transitions at any given temperature (within the denaturation range) depends on the time required to reach that temperature; therefore, the thermal denaturation of CA is under kinetic control and the DSC transitions are distorted by changing scan rates. The thermal denaturation of several soluble proteins such ac CA has been found to conform to this model. Fig. 4 shows temperaturedependence of excess heat capacity for CA at several scan rates.

 $T_{m}(350)$ 

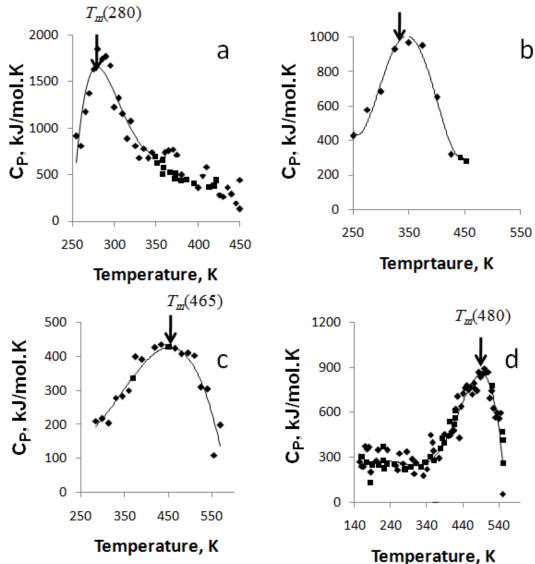
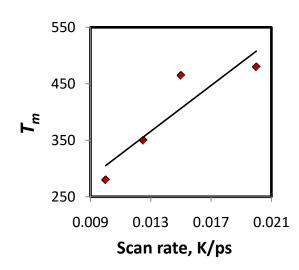


Fig. 4: Characteristic heat-capacity curves for CA, with four scan rates a) 0.01, b) 0.0125, c) 0.015 and d) 0.02 K/ps

As it may be seen in this figure the traces were scan rate-dependent. It may be concluded, therefore, that the thermal denaturation of CA is kinetically controlled under the conditions employed. The denaturation shapes of the transitions agree in general with the results of Brouillette et al.<sup>19</sup> The results displayed in this figure show, however, that the DSC transitions are highly scanning rate-dependent. The curves of  $C_{\rm P}$ , vs *T* are shifted to higher temperature with increasing scan rate (Fig. 3). This illustrates the

importance of varying the scan rate or the rate of unfolding on the profiles. The temperature of maximum  $C_{\rm P}$ , ( $T_{\rm max}$ ) obeys the equation (2). As scan rate increases further, the profile shifts to higher temperature which is obvious in our results. Fig. 5 shows scan rate dependence of CA denaturation.



*Fig.* 5 : Scan rate dependence of thermal denaturations of CA

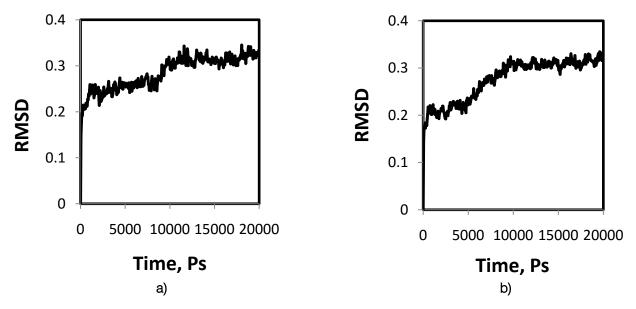


Fig. 6 : Calculated RMSD by molecular dynamics for CA for scan rates of a) 0.01 and b) 0.0125K/ps

The area under a DSC curve normally yields the calorimetric enthalpy of denaturation of the protein,  $\Delta H$ . The position of the peak yields the  $T_m$  for denaturation. CA thermally denature as a single peak in DSC. The

fitted values for the calorimetrically determined apparent thermodynamic parameters for the denaturation of the CA in each scan rate are shown in Table 1.

Calculated RMSD by molecular dynamics for

CA for studied scan rates is depicted in Fig. 6. It shows

that all of the systems reach a stable state.

*Table 1 : T*<sub>m</sub> and thermodynamic parameters of CA denaturation in different scan rates.

Scan rate	$\Delta H$ (neutral) (kJ/mol)	$\Delta S$ (kJ/mol)	7 <sub>m</sub> (neutral)	T <sub>m</sub> (acidic)
0.01	62207	222.17	280	278
0.0125	76986	219.96	350	341
0.015 0.02	78914 79638	169.71 165.91	465 480	415 Not done

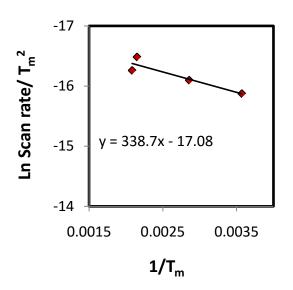
Where  $\Delta S_{\text{Tm}}$  is the entropy of unfolding at the melting temperature obtained by:

$$\Delta S_{T_m} = \frac{\Delta H}{T_m} \tag{7}$$

Since the Gibbs free energy of unfolding at  $T_m$  is equal to zero:  $\Delta G_{T_m} = 0$ 

The enthalpy of melting  $\Delta H$  was determined by integrating the area under the peak. It is common practice to determine the enthalpy of protein unfolding by differential scanning calorimetry (DSC). This approach limits the available data to higher temperatures where most proteins denature, ranging from about 273 to 405 K.

Proteins have to be artificially destabilized to reduce their melting temperature. One of the most common means to destabilize a protein is to reduce the pH. It was shown using CA as a model protein that the enthalpy of unfolding determined by simulated DSC at various pHs is equal to the enthalpy of unfolding determined by isothermal calorimetric titration of the protein with acid.<sup>9</sup> The free energy of denaturation at a reference temperature *T* was calculated.



*Fig.* 7 : Plot of  $Ln(V/T_m^2)$  Versus  $1/T_m$ , where each data point refers to one of the four scan rates used

Fig. 7 shows Arrhenius plots of the scan-rate dependent changes in  $T_{\rm m}$ . The slopes of these lines provide the apparent activation energies of denaturation.<sup>31</sup>

Since Sanchez-Ruiz and co-workers and other authors <sup>33, 34</sup> reported more or less equivalent activation energies with each of the different Arrhenius-based analysis methods, we have demonstrated only one of these methods here (in Figure 7).  $T_m$  values for CA were found to be scan-rate-dependent. Kinetic activation energies for irreversible denaturation were derived from the scan rate dependence of the DSC transitions using Arrhenius plots as described by Sanchez-Ruiz et al., <sup>18</sup> and these plots of ln(scan rate/ $T_m^2$ ) against  $1/T_m$  are also given in Fig. 7. The slopes of these plots provide the activation energies for irreversible denaturation, which are 338.7/R kJ/mol for CA which R is gas constant.

#### c) pH Effect

Further exploration of the denaturation of CA was also performed as a function of scan rate in acidic pH. Protein stability could be altered dramatically by changing pH or by changing pH in a scan rate mode. For example, lowering the pH to acidic value lowered the  $T_{\rm m}$  by 8, 16, or 25 °C experimentally.<sup>9</sup> Here we used the MD simulation and showed thermal denaturation of the sample was pH and scan-rate dependent. We have made DSC studies into the thermal stability of CA within different pH at neutral and acidic pH values and different scan rates and compared these calculation results to those of the native protein in experimental condition. Effects of the pH on  $T_m$  of both scan rate and thermal denaturation can be taken as an interesting result. The scan rates used were 0.01, 0.0125 and 0.015 K/ps in a lower acidic media which all carboxyl groups (COO<sup>-</sup>) were protonated. Fig. 8 shows a typical DSC profiles for a protein in three different scan rates and acidic media. Melting temperature as a function of pH for different scan rates was obtained.  $T_{\rm m}$  values in each scan rate for acidic condition are presented in Table 1.

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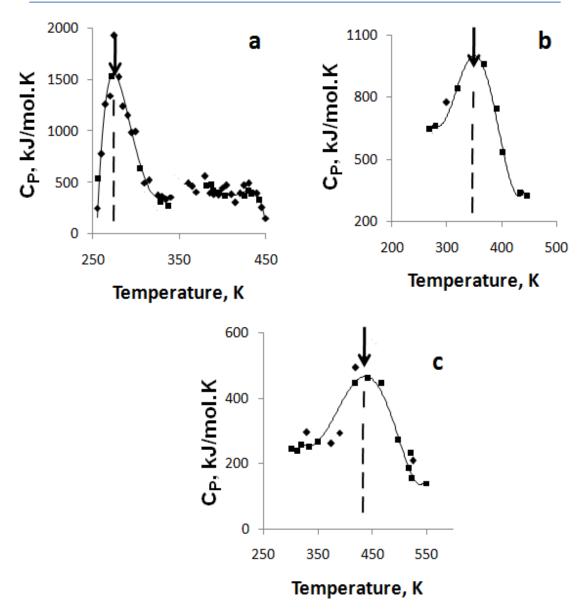


Fig. 8 : DSC profiles for CA in three different scan rates in created acidic media

Thermal stability was essentially pH dependent. In lower pH, the  $T_m$  gradually decreased as the pH became more acidic, as expected for a protein that binds protons more tightly in the non-native state.<sup>15</sup>

The calorimetric enthalpy of unfolding was calculated as the area of the unfolding peak, normalized to the molar protein concentration. The unfolding enthalpy was linearly proportional to  $T_{\rm m}$ , and the slope of H vs  $T_{\rm m}$  yielded  $C_{\rm p}$ .

#### IV. CONCLUSION

BCA is a protein that is commonly used as a model for biophysical studies, and this work provides additional information on the effect of different denaturants of scan rate, pH and temperature on the stability of BCA. It demonstrates that increasing the scan rate in acidic media reduces protein stability. Increasing the scan rate in neutral media improves the stability of BCA. Calculations were performed using molecular dynamics simulation with four scan rates. The calorimetric traces were found to be scan-rate-dependent under the conditions employed.

### V. Acknowledgments

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#### Approach:

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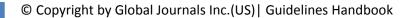
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- Present a background, such as by describing the question that was addressed by creation an exacting study.
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- Do not present the similar data more than once.
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#### Approach

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- Give details all of your remarks as much as possible, focus on mechanisms.
- Make a decision if the tentative design sufficiently addressed the theory, and whether or not it was correctly restricted.
- Try to present substitute explanations if sensible alternatives be present.
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- Recommendations for detailed papers will offer supplementary suggestions.

#### Approach:

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Result	Well organized, Clear and specific, Correct units with precision, correct data, well structuring of paragraph, no grammar and spelling mistake	Complete and embarrassed text, difficult to comprehend	Irregular format with wrong facts and figures	
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References	Complete and correct format, well organized	Beside the point, Incomplete	Wrong format and structuring	

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