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highlights

Refraction As A Function Of Atomic Gravitation

Mathieu-type series

G- Closed Sets in a Topological Space

Fixed-Dome Biodigester



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Contents of the Volume

- i. Copyright Notice
- ii. Editorial Board Members
- iii. Chief Author and Dean
- iv. Table of Contents
- v. From the Chief Editor's Desk
- vi. Research and Review Papers

- 1. Some Mathieu-type series associated with the I-function and generalized -Gauss hypergeometric function **2-9**
- 2. On g- Closed Sets in a Topological Space **10-12**
- 3. A Generalized Probability Distribution Pertaining To Product of Special Functions with Applications **13-20**
- 4. Determination of copper (II) by simple extraction procedure prior to differential pulse polarography using 4-(2-hydroxy phenyl ethaminodiol), benzene-1,3-diol **21-29**
- 5. Refraction As A Function Of Atomic Gravitation Fresh Insight Into The Natureof Space And A New Possibility In The Creation Of Metamaterials **30-32**
- 6. Fixed-Dome Biodigester Construction And Determination Of Effects Of Temperature On The Performance **33-40**

- vii. Auxiliary Memberships
- viii. Process of Submission of Research Paper
- ix. Preferred Author Guidelines
- x. Index

From the Chief Author's Desk

We see a drastic momentum everywhere in all fields now a day. Which in turns, say a lot to everyone to excel with all possible way. The need of the hour is to pick the right key at the right time with all extras. Citing the computer versions, any automobile models, infrastructures, etc. It is not the result of any preplanning but the implementations of planning.

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Some Mathieu-type series associated with the I-function and generalized ω -Gauss hyper geometric function

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Dr. V. B. L. Chourasia¹ Ravi Shanker Dubey²

Abstract- Closed integral form expression have been established pertaining to the Mathieu-type a-series and associated alternating series whose terms involve I-function and generalized ω -Gauss hyper geometric function.

Keywords- Dirichlet Series; I-Function; Integral representation of series; Laplace integral representation of Dirichlet series; Mathieu-type a-series and alternating a-series.

I. INTRODUCTION AND PRELIMINARIES

In a series of papers the authors, Srivastava and Tomovski considered the special Mathieu-type a-series and

$$I_{p_i, q_i : r}^{m, n} [z] = \frac{1}{2\pi i} \int_L \frac{\prod_{k=1}^m \Gamma(b_k - \beta_k \xi) \prod_{k=1}^n \Gamma(1 - a_k + \alpha_k \xi) z^\xi d\xi}{\sum_{i=1}^r \left\{ \prod_{k=m+1}^{q_i} \Gamma(1 - b_{ki} + \beta_{ki} \xi) \prod_{k=n+1}^{p_i} \Gamma(a_{ki} - \alpha_{ki} \xi) \right\}}. \quad (1)$$

For conditions of convergence, asymptotic behaviour and other details, see [11].

Consider the Mathieu-type a-series $\tau_{\lambda, \mu}$ and its alternating variant $\tilde{\tau}_{\lambda, \mu}$ defined by

$$\tau_{\lambda, \mu} \left(I_{p_i+1, q_i : r}^{m, n+1} ; c ; s \right) := \sum_{j=1}^{\infty} \frac{I_{p_{i+1}, q_i, r}^{m, n+1} \left(\begin{matrix} (a_k, a_k)_{1, n+1} ; (a_{ki}, a_{ki})_{n+2, p_i} \\ s \Big| c_j \end{matrix} \right)}{c_j^\lambda (c_j + s)^\mu} \quad (2)$$

$$\tilde{\tau}_{\lambda, \mu} \left(I_{p_{i+1}, q_i : r}^{m, n+1} ; c ; s \right) := \sum_{j=1}^{\infty} \frac{(-1)^{j-1} I_{p_{i+1}, q_i : r}^{m, n+1} \left(\frac{s}{c_j} \left| \begin{matrix} (a_k, a_k)_{1, n+1} ; (a_{ki}, a_{ki})_{n+2, p_i} \\ (b_k, \beta_k)_{1, m} ; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{matrix} \right. \right)}{c_j^\lambda (c_j + s)^\mu}, \quad (3)$$

where we make the convention that the real sequence $c = \{c_n\}_{n \in \mathbb{N}}$ increases and tends to ∞ , equivalently,

$$c : 0 < c_1 < c_2 < \dots c_n \uparrow \infty. \quad (4)$$

II. INTEGRAL REPRESENTATIONS OF $\tau_{\lambda, \mu} \left(I_{p_{i+1}, q_i : r}^{m, n+1} ; c ; s \right)$ and $\tilde{\tau}_{\lambda, \mu} \left(I_{p_{i+1}, q_i : r}^{m, n+1} ; c ; s \right)$

In the course of our investigation, one of the main tool is the following result providing the Laplace transform of

$$x^{\lambda-1} I_{p_i, q_i : r}^{m, n} (wx|.), \text{ for real } w$$

$$\begin{aligned} & \int_0^\infty e^{-Ax} x^{\lambda-1} I_{p_i, q_i : r}^{m, n} \left(wx^\rho \left| \begin{matrix} (a_k, a_k)_{1, n} ; (a_{ki}, a_{ki})_{n+1, p_i} \\ (b_k, \beta_k)_{1, m} ; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{matrix} \right. \right) dx \\ &= A^{-\lambda} I_{p_{i+1}, q_i : r}^{m, n+1} \left(\frac{w}{A^\rho} \left| \begin{matrix} (1-\lambda, \rho), (a_k, a_k)_{1, n} ; (a_{ki}, a_{ki})_{n+1, p_i} \\ (b_k, \beta_k)_{1, m} ; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{matrix} \right. \right), \end{aligned} \quad (5)$$

where p_i ($i=1, 2, \dots, r$), q_i ($i=1, 2, \dots, r$), m, n are integers satisfying $0 \leq n \leq p_i$, $0 \leq m \leq q_i$ ($i=1, 2, \dots, r$); r is finite, $a_j, \beta_j, a_{ji}, \beta_{ji}$ are real and positive and a_j, b_j, a_{ji}, b_{ji} are complex numbers; $\text{Re} \left\{ \lambda + \rho \frac{b_j}{\beta_j} \right\} > 0$, $j = 1, 2, \dots, m$,

The result in (5) can be establish easily.

Theorem 1: Let $\lambda > 0$, $\mu > 0$, $s > 0$, $a_{p+1} = 1 - \lambda$, $\alpha_{p+1} = 1$ and let the sequence c satisfies (4). Then

$$\tau_{\lambda, \mu} \left(I_{p_{i+1}, q_i, r}^{m, n+1} ; c ; s \right) = \eta_c^I (\lambda + 1, \mu) + \mu \eta_c^I (\lambda, \mu + 1) \quad (6)$$

$$\tilde{\tau}_{\lambda, \mu} \left(I_{p_{i+1}, q_i, r}^{m, n+1}; c; s \right) = \tilde{\eta}_c^I (\lambda + 1, \mu) + \mu \tilde{\eta}_c^I (\lambda, \mu + 1), \quad (7)$$

where

$$\mathfrak{I}_c^I (u, v) := \int_{c_1}^{\infty} \frac{\left[c^{-1}(x) \right]}{x^u (s+x)^v} I_{p_{i+1}, q_i, r}^{m, n+1} \left(\frac{s}{x} \middle| \begin{matrix} (u, 1), (a_k, \alpha_k)_{1, n}; (a_{ki}, \alpha_{ki})_{n+1, p_i} \\ (b_k, \beta_k)_{1, m}; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{matrix} \right) dx \quad (8)$$

$$\tilde{\mathfrak{I}}_c^I (u, v) := \int_{c_1}^{\infty} \frac{\sin^2 \left(\frac{\pi}{2} [c^{-1}(x)] \right)}{x^u (r+x)^v} I_{p_{i+1}, q_i, r}^{m, n+1} \left(\frac{s}{x} \middle| \begin{matrix} (u, 1), (a_k, \alpha_k)_{1, n}; (a_{ki}, \alpha_{ki})_{n+1, p_i} \\ (b_k, \beta_k)_{1, m}; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{matrix} \right) dx, \quad (9)$$

Where $c: \mathbb{R}_+ \rightarrow \mathbb{R}_+$ is an increasing function such that $c(x)|_{x \in \mathbb{N}} = c$, $c^{-1}(x)$ denotes the inverse of $c(x)$, $[c^{-1}(x)]$ stands for the integer part of the quantity $c^{-1}(x)$.

Proof: On taking $z = c_n + s$ in the following known formula

$$\Gamma(\mu) z^{-\mu} = \int_0^{\infty} e^{-zt} t^{\mu-1} dt \quad (Re\{z\} > 0, Re\{\mu\} > 0), \quad (10)$$

And on specifying $A = c_j$, $\rho = s$ in (5) finally inserting $a_{p+1} = 1 - \lambda$, $\alpha_{p+1} = 1$, we conclude

$$\begin{aligned} \tau_{\lambda, \mu} \left(I_{p_{i+1}, q_i, r}^{m, n+1}; c; s \right) &= \sum_{j=1}^{\infty} \frac{I_{p_{i+1}, q_i, r}^{m, n+1} \left(\frac{s}{c_j} \middle| \begin{matrix} (1-\lambda, 1), (a_k, \alpha_k)_{1, n}; (a_{ki}, \alpha_{ki})_{n+1, p_i} \\ (b_k, \beta_k)_{1, m}; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{matrix} \right)}{c_j^{\lambda} (c_j + s)^{\mu}} \\ &= \sum_{j=1}^{\infty} \int_0^{\infty} e^{-c_j \theta} \theta^{\lambda-1} I_{p_{i+1}, q_i, r}^{m, n+1} \left(\theta s \middle| \begin{matrix} (a_k, \alpha_k)_{1, n}; (a_{ki}, \alpha_{ki})_{n+1, p_i} \\ (b_k, \beta_k)_{1, m}; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{matrix} \right) ds \int_0^{\infty} \frac{t^{\mu-1}}{\Gamma \mu} e^{-(c_j + s)t} dt \\ &= \frac{1}{\Gamma \mu} \int_0^{\infty} \int_0^{\infty} \left(\sum_{j=1}^{\infty} e^{-c_j(\theta+t)} \right) e^{-st} \theta^{\lambda-1} t^{\mu-1} I_{p_i, q_i, r}^{m, n} \left(\theta s \middle| \begin{matrix} (a_k, \alpha_k)_{1, n}; (a_{ki}, \alpha_{ki})_{n+1, p_i} \\ (b_k, \beta_k)_{1, m}; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{matrix} \right) d\theta dt, \quad (11) \end{aligned}$$

where $\mu > 0$.

The inside Dirichlet Series

$$D_c(\theta + t) = \sum_{j=1}^{\infty} e^{-(\theta+t)c_j} \quad (12)$$

Possesses the Laplace integral form representation [1, 2], such that we can express it as follows:

$$\begin{aligned} D_c(\theta, t) &= (\theta + t) \int_0^{\infty} e^{-(\theta+t)x} \left(\sum_{j: c_j \leq x} 1 \right) dx \\ &= (\theta + t) \int_{c_1}^{\infty} e^{-(\theta+t)x} [c^{-1}(x)] dx \end{aligned} \quad (13)$$

with $[c^{-1}(x)] = 0$ for $(x \in [0, c_1])$.

Therefore, we conclude that

$$\begin{aligned} \tau_{\lambda, \mu} \left(I_{p_{i+1}, q_i, r}^{m, n+1}; c; s \right) &= \frac{1}{\Gamma(\mu)} \int_0^{\infty} \int_0^{\infty} \int_{c_1}^{\infty} e^{-t(s+x)-\theta x} \theta^{\lambda} t^{\mu-1} I_{p_i, q_i, r}^{m, n} \left(\theta s \left| \begin{smallmatrix} (a_k, a_k)_{1, n}; (a_{ki}, a_{ki})_{n+1, p_i} \\ (b_k, \beta_k)_{1, m}; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{smallmatrix} \right. \right) [c^{-1}(x)] d\theta dt dx \quad (H_s) \\ &+ \frac{1}{\Gamma(\mu)} \int_0^{\infty} \int_0^{\infty} \int_{c_1}^{\infty} e^{-t(s+x)-\theta x} \theta^{\lambda-1} t^{\mu} I_{p_i, q_i, r}^{m, n} \left(\theta s \left| \begin{smallmatrix} (a_k, a_k)_{1, n}; (a_{ki}, a_{ki})_{n+1, p_i} \\ (b_k, \beta_k)_{1, m}; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{smallmatrix} \right. \right) [c^{-1}(x)] d\theta dt dx. \quad (H_t) \end{aligned}$$

Since

$$\begin{aligned} H_s &= \frac{1}{\Gamma(\mu)} \int_{c_1}^{\infty} \left(\int_0^{\infty} e^{-\theta x} \theta^{\lambda} I_{p_{i+1}, q_i, r}^{m, n} \left(\theta s \left| \begin{smallmatrix} (a_k, a_k)_{1, n}; (a_{ki}, a_{ki})_{n+1, p_i} \\ (b_k, \beta_k)_{1, m}; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{smallmatrix} \right. \right) d\theta \right) \left(\int_0^{\infty} e^{-(x+s)t} t^{\mu-1} dt \right) [c^{-1}(x)] dx \\ &= \int_{c_1}^{\infty} \frac{[c^{-1}(x)]}{x^{\lambda+1} (s+x)^{\mu}} I_{p_{i+1}, q_i, r}^{m, n+1} \left(\frac{s}{x} \left| \begin{smallmatrix} (-\lambda, 1), (a_k, a_k)_{1, n}; (a_{ki}, a_{ki})_{n+1, p_i} \\ (b_k, \beta_k)_{1, m}; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{smallmatrix} \right. \right) dx, \end{aligned}$$

introducing the auxiliary integral

$$\mathfrak{I}_c^I(u, v) = \int_{c_1}^{\infty} \frac{[c^{-1}(x)]}{x^u (s+x)^v} I_{p_i, q_i, r}^{m, n} \left(\frac{s}{x} \left| \begin{smallmatrix} (1-u, 1)(a_k, a_k)_{1, n}; (a_{ki}, a_{ki})_{n+1, p_i} \\ (b_k, \beta_k)_{1, m}; (b_{ki}, \beta_{ki})_{m+1, q_i} \end{smallmatrix} \right. \right) dx$$

And

$$\begin{aligned}
H_t &= \frac{1}{\Gamma(\mu)} \int_{c_1}^{\infty} \left(\int_0^{\infty} e^{-\theta x} \theta^{\lambda-1} I_{p_{i+1}, q_i; r}^{m, n} \left(\theta s \left| \begin{smallmatrix} (a_k, a_k)_{1, n}; (a_{ki}, a_{ki})_{n+1, p_i} \\ (b_k, b_k)_{1, m}; (b_{ki}, b_{ki})_{m+1, q_i} \end{smallmatrix} \right. \right) d\theta \right) \left(\int_0^{\infty} e^{-(x+s)t} t^{\mu} dt \right) \left[c^{-1}(x) \right] dx \\
&= \mu \int_{c_1}^{\infty} \frac{\left[c^{-1}(x) \right]}{x^{\lambda} (s+x)^{\mu+1}} I_{p_{i+1}, q_i; r}^{m, n+1} \left(\frac{s}{x} \left| \begin{smallmatrix} (-\lambda, 1), (a_k, a_k)_{1, n}; (a_{ki}, a_{ki})_{n+1, p_i} \\ (b_k, b_k)_{1, m}; (b_{ki}, b_{ki})_{m+1, q_i} \end{smallmatrix} \right. \right) dx.
\end{aligned}$$

Now we can say that

$$H_s = \mathfrak{I}_c^I(\lambda+1, \mu) \quad \text{and} \quad H_t = \mu \mathfrak{I}_c^I(\lambda, \mu+1).$$

These proves the first assertion of the Theorem 1.

The derivation (7) is similar to the previous procedure; the only novelty is the new inside alternating Dirichlet series [3, section 4, p. 77], define as follows

$$\begin{aligned}
\tilde{D}_c(\theta+t) &= \sum_{j=1}^{\infty} (-1)^{j-1} e^{-c_j(\theta+t)} \\
&= (\theta+t) \int_0^{\infty} e^{-(\theta+t)x} \left(\sum_{j: c_j \leq x} (-1)^{j-1} \right) dx \\
&= \frac{(\theta+t)}{2} \int_0^{\infty} e^{-(\theta+t)x} \left(1 - (-1)^{\left[c^{-1}(x) \right]} \right) dx \\
&= (\theta+t) \int_0^{\infty} e^{-(\theta+t)x} \sin^2 \left(\frac{\pi}{2} \left[c^{-1}(x) \right] \right) dx. \tag{14}
\end{aligned}$$

This formula helps in proving the theorem.

III. MATHIEU-TYPE SERIES WITH THE GENERALIZED ω -GAUSS HYPERGEOMETRIC FUNCTION

Here we give the series representation of ${}_{e+f} \mathbf{R}_{g+h}^{\omega} \left(\begin{smallmatrix} a_1, \dots, a_e; b_1, \dots, b_f \\ c_1, \dots, c_g; d_1, \dots, d_h \end{smallmatrix} \middle| z \right)$ which is the generalization of the ω -Gauss hyper geometric function [12, 13, 14].

$${}_{e+f} \mathbf{R}_{g+h}^{\omega} \left(\begin{smallmatrix} a_1, \dots, a_e; b_1, \dots, b_f \\ c_1, \dots, c_g; d_1, \dots, d_h \end{smallmatrix} \middle| z \right) = \frac{\Gamma(d_1) \dots \Gamma(d_h)}{\Gamma(b_1) \dots \Gamma(b_f)} \sum_{k=0}^{\infty} \frac{(a_1)_k (a_2)_k \dots (a_e)_k}{(c_1)_k (c_2)_k \dots (c_g)_k} \frac{\Gamma(b_1 + \omega k) \dots \Gamma(b_g + \omega k)}{\Gamma(d_1 + \omega k) \dots \Gamma(d_h + \omega k)} \frac{z^k}{k!}, \tag{15}$$

where a_e, b_f, c_g, d_h are complex numbers, $b_g + \omega k$ and $d_h + \omega k \neq 0, -1, -2, \dots$ the series converges uniformly in the region $|z| < 1$, $\sum d_h - b_g > 0$.

Consider the Mathieu-type \mathbf{a} -series $\mathbf{V}_{\lambda, u}$ and its alternating variant $\tilde{\mathbf{V}}_{\lambda, u}$ defined by

$$\mathbf{V}_{\lambda, \mu} \left(e_{+1+f} \overset{\omega}{\mathbf{R}}_{g+h} ; \mathbf{c} ; \mathbf{r} \right) := \sum_{j=1}^{\infty} \frac{e_{+1+f} \overset{\omega}{\mathbf{R}}_{g+h} \left(-\frac{\mathbf{r}}{\mathbf{c}_j} \middle| \mathbf{a}_{e+1} ; \mathbf{b}_f \right)}{\mathbf{c}_j^{\lambda} (\mathbf{c}_j + \mathbf{r})^{\mu}}, \quad (16)$$

and

$$\tilde{\mathbf{V}}_{\lambda, \mu} \left(e_{+1+f} \overset{\omega}{\mathbf{R}}_{g+h} ; \mathbf{c} ; \mathbf{r} \right) := \sum_{j=1}^{\infty} \frac{(-1)^{j-1} e_{+1+f} \overset{\omega}{\mathbf{R}}_{g+h} \left(-\frac{\mathbf{r}}{\mathbf{c}_j} \middle| \mathbf{a}_{e+1} ; \mathbf{b}_f \right)}{\mathbf{c}_j^{\lambda} (\mathbf{c}_j + \mathbf{r})^{\mu}}, \quad (17)$$

where sequence \mathbf{c} is already defined

IV. Integral Representations of $\mathbf{V}_{\lambda, u} \left(e_{+1+f} \overset{\omega}{\mathbf{R}}_{g+h} ; \mathbf{c} ; \mathbf{r} \right)$ and $\tilde{\mathbf{V}}_{\lambda, u} \left(e_{+1+f} \overset{\omega}{\mathbf{R}}_{g+h} ; \mathbf{c} ; \mathbf{r} \right)$

In the course of our investigation, one of the main tool in the following result providing the Laplace transform of $\mathbf{x}^{\lambda-1} e_{+f} \overset{\omega}{\mathbf{R}}_{g+h} (w\mathbf{x} | \cdot)$, that is

$$\int_0^{\infty} e^{-Ax} \mathbf{x}^{\lambda-1} e_{+f} \overset{\omega}{\mathbf{R}}_{g+h} \left(\begin{matrix} a_e; b_f \\ c_g; d_h \end{matrix} \middle| -w\mathbf{x} \right) d\mathbf{x} = A^{-\lambda} \Gamma(\lambda) e_{+1+f} \overset{\omega}{\mathbf{R}}_{g+h} \left(\begin{matrix} \lambda, a_e; b_f \\ c_g; d_h \end{matrix} \middle| -\frac{w}{A} \right), \quad (18)$$

where $\operatorname{Re}\{A\} > 0$, $\operatorname{Re}\{\lambda\} > 0$, a_e, b_f, c_g, d_h are complex numbers, $w \in \mathbb{R}^+$, $b_g + \omega k$ and $d_h + \omega k \neq 0, -1, -2, \dots$ the series converges uniformly in the region $|z| < 1$ and $\sum d_h - b_g > 0$.

Theorem 2: Let $\lambda > 0$, $\mu > 0$, $r > 0$, $a_{e+1} = \lambda$ and let the sequence \mathbf{c} satisfies (4). Then

$$\mathbf{V}_{\lambda, \mu} \left(e_{+1+f} \overset{\omega}{\mathbf{R}}_{g+h} ; \mathbf{c} ; \mathbf{r} \right) = \left(\mathfrak{I}_c^R (\lambda+1, \mu) + \mu \tilde{\mathfrak{I}}_c^R (\lambda, \mu+1) \right) \quad (19)$$

and

$$\tilde{\mathbf{V}}_{\lambda, \mu} \left(e_{+1+f} \overset{\omega}{\mathbf{R}}_{g+h} ; \mathbf{c} ; \mathbf{r} \right) = \left(\tilde{\mathfrak{I}}_c^R (\lambda+1, \mu) + \mu \tilde{\mathfrak{I}}_c^R (\lambda, \mu+1) \right), \quad (20)$$

where

$$\mathfrak{I}_c^{\omega}(u, v) := \int_{c_1}^{\infty} \frac{\left[c^{-1}(x) \right]}{x^u (x+r)^v} {}_{e+1+f}^{\omega} R_{g+h} \left(\begin{matrix} a_e, u; b_f \\ c_g; d_h \end{matrix} \middle| -\frac{r}{x} \right) dx \quad (21)$$

and

$$\tilde{\mathfrak{I}}_c^{\omega}(u, v) := \int_{c_1}^{\infty} \frac{\sin^2 \left(\frac{\pi}{2} \left[c^{-1}(x) \right] \right)}{x^u (x+r)^v} {}_{e+1+f}^{\omega} R_{g+h} \left(\begin{matrix} a_e, u; b_f \\ c_g; d_h \end{matrix} \middle| -\frac{r}{x} \right) dx, \quad (22)$$

where $c : R_+ \rightarrow R_+$ is an increasing function.

Proof: On taking $A = c_j$, $w = r$ in (18) and inserting $a_{e+1} = \lambda$, we conclude that

$$\begin{aligned} \mathfrak{V}_{\lambda, \mu} \left({}_{e+1+f}^{\omega} R_{g+h}; c; r \right) &= \sum_{j=1}^{\infty} \frac{{}_{e+1+f}^{\omega} R_{g+h} \left(\begin{matrix} \lambda, a_e; b_f \\ c_j; d_h \end{matrix} \middle| -\frac{r}{c_j} \right)}{c_j^{\lambda} (c_j + r)^{\mu}} \\ &= \sum_{j=1}^{\infty} \frac{1}{\Gamma(\lambda)} \int_0^{\infty} e^{-c_j \theta} \theta^{\lambda-1} {}_{e+f}^{\omega} R_{g+h} \left(-r\theta \Big| {}_{c_g; d_h}^{a_e; b_f} \right) d\theta \int_0^{\infty} \frac{t^{\mu-1}}{\Gamma(\mu)} e^{-(c_j + r)t} dt \\ &= \frac{1}{\Gamma(\lambda) \Gamma(\mu)} \int_0^{\infty} \int_0^{\infty} \left(\sum_{j=1}^{\infty} e^{-c_j(\theta+t)} \right) e^{-rt} \theta^{\lambda-1} t^{\mu-1} {}_{e+f}^{\omega} R_{g+h} \left(-r\theta \Big| {}_{c_g; d_h}^{a_e; b_f} \right) d\theta dt, \end{aligned}$$

and on using the result of (13) and (14), we obtain

$$\begin{aligned} \mathfrak{V}_{\lambda, \mu} \left({}_{e+1+f}^{\omega} R_{g+h}; c; r \right) &= \frac{1}{\Gamma(\lambda) \Gamma(\mu)} \int_0^{\infty} \int_0^{\infty} \int_{c_1}^{\infty} e^{-(r+x)t - \theta x} \theta^{\lambda} t^{\mu-1} {}_{e+f}^{\omega} R_{g+h} \left(-r\theta \Big| {}_{c_g; d_h}^{a_e; b_f} \right) \left[c^{-1}(x) \right] d\theta dt dx \\ &\quad + \frac{1}{\Gamma(\lambda) \Gamma(\mu)} \int_0^{\infty} \int_0^{\infty} \int_{c_1}^{\infty} e^{-(r+x)t - \theta x} \theta^{\lambda-1} t^{\mu} {}_{e+f}^{\omega} R_{g+h} \left(-r\theta \Big| {}_{c_g; d_h}^{a_e; b_f} \right) \left[c^{-1}(x) \right] d\theta dt dx \\ &= \int_{c_1}^{\infty} \frac{\left[c^{-1}(x) \right]}{x^{\lambda+1} (x+r)^{\mu}} {}_{e+f}^{\omega} R_{g+h} \left(\begin{matrix} a_e, \lambda+1; b_f \\ c_g; d_h \end{matrix} \middle| -\frac{r}{x} \right) dx \\ &\quad + \mu \int_{c_1}^{\infty} \frac{\left[c^{-1}(x) \right]}{x^{\lambda} (x+r)^{\mu+1}} {}_{e+f}^{\omega} R_{g+h} \left(\begin{matrix} a_e, \lambda; b_f \\ c_g; d_h \end{matrix} \middle| -\frac{r}{x} \right) dx, \end{aligned}$$

and finally, we get

$$\mathbf{U}_{\lambda,\mu} \left({}_{e+1+f}^{\omega} \mathbf{R}_{g+h}^{\omega} ; \mathbf{c} ; \mathbf{r} \right) = \mathfrak{I}_c^R (\lambda+1, \mu) + \mu \mathfrak{I}_c^R (\lambda, \mu+1).$$

In a similar manner, we can easily obtain

$$\tilde{\mathbf{U}}_{\lambda,\mu} \left({}_{e+1+f}^{\omega} \mathbf{R}_{g+h}^{\omega} ; \mathbf{c} ; \mathbf{r} \right) = \tilde{\mathfrak{I}}_c^R (\lambda+1, \mu) + \mu \tilde{\mathfrak{I}}_c^R (\lambda, \mu+1)$$

V. CONSEQUENCES AND IMPORTANT SPECIAL CASES

Many well known special functions, such as $\text{erf}(x)$, and the Bessel, Fox-Wright $\tilde{\psi}$, Whittaker, Meijer G-, and generalized hyper geometric functions ${}_pF_q$, and Jacobi polynomials and other elliptic functions are included in the class of functions which can be expressed in terms of I-function. We give below certain known results pertaining to the Theorem 1 and Theorem 2.

- For $r = 1$, the result given in Theorem 1 reduced to a known result recently obtained in [7].
- Taking $r = 1$ and $\alpha_j = \beta_j = \alpha_{ji} = \beta_{ji} = 1$, the Theorem 1 reduced to another known result established in [5].
- Taking $\omega = 0$, the Theorem 2 reduces to the known result given in [7].

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On g- Closed Sets in a Topological Space

Dedicated to Professor S. Chowdhary on his 65th birthday

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The purpose of this research article is to explain the meaning of g-closed sets, which is more understandable to the readers.

I. INTERIOR OF A SET

Let A be a subset of a topological space X . A point $p \in A$ is called an interior point of A , if p belongs to an open set G contained in A i.e. $p \in G \subset A$.

The set of interior points of A is denoted by $\text{int}(A)$ or A° , which is called the interior of A .

II. CLOSURE OF A SET

Let A be a subset of a topological space X . The closure of A is defined as the intersection of all closed super sets of A .

Closure of A is denoted by \bar{A} or A^-

III. g-CLOSED SET

Let (X, τ) be a topological space and A be its subset, then A is called an g-closed set if $\tau - \text{Cl}(A) \subseteq u$, whenever $A \subseteq u$ and $u \in \tau$ (i.e. u is a open set) . .

Now, Let $X = \{a, b, c, d, e\}$ be a non-empty set and

$\tau = \{\phi, X, \{a, b, c\}, \{d, e\}, \{c\}, \{d, e, c\}\}$ is a collection of subset of X

A. Show that τ is a Topology defined on X

(i) $\phi, X \in \tau$.

(ii) $\phi \cup X = X \in \tau$

$X \cup \{a, b, c\} = X \in \tau$

$\{a, b, c\} \cup \{d, e\} = \{a, b, c, d, e\} = X \in \tau$

$\{d, e\} \cup \{c\} = \{d, e, c\} \in \tau$

$\{c\} \cup \{d, e, c\} = \{d, e, c\} \in \tau$

(iii) $\phi \cap X = \phi \in \tau$

$X \cap \{a, b, c\} = \{a, b, c\} \in \tau$

$\{a, b, c\} \cap \{d, e\} = \phi \in \tau$

$\{d, e\} \cap \{c\} = \phi \in \tau$

$\{c\} \cap \{d, e, c\} = \{c\} \in \tau$

Here all three conditions for topology is satisfied, it means that τ is a topology on X .

Now we have all possible subsets of $X = 2^5 = 2 \times 2 \times 2 \times 2 \times 2 = 32$, which are given below.

$\phi, X, \{a\}, \{b\}, \{c\}, \{d\}, \{e\}, \{a, b\}, \{a, c\}, \{a, d\}, \{a, e\}, \{b, c\}, \{b, d\}, \{b, e\}, \{c, d\}, \{c, e\}, \{a, b, c\}, \{a, b, d\}, \{a, b, e\}, \{a, c, d\}, \{a, c, e\}, \{a, d, e\}, \{b, c, d\}, \{b, c, e\}, \{b, d, e\}, \{c, d, e\}, \{d, e, a\}, \{d, e, b\}, \{a, b, c, d\}, \{a, b, c, e\}, \{a, b, d, e\}, \{a, c, d, e\}, \{b, c, d, e\}, \{c, e, d, a\}, \{d, e, a, b\}$

B. Verifications for g-closed sets

As given, $X = \{a, b, c, d, e\}$

And, $\tau = \{\phi, X, \{a, b, c\}, \{d, e\}, \{c\}, \{d, e, c\}\}$

Now we have also

Open sets: $\phi, X, \{a, b, c\}, \{d, e\}, \{c\}, \{d, e, c\}$

Closed sets: $X, \phi, \{d, e\}, \{a, b, c\}, \{a, b, d, e\}, \{a, b\}$

Now as per definition of g-closed set, here we are verifying for all (32) subsets of X ;

(i) Let $A = \{a\}$, $u = \{a, b, c\}$ such that $A \subseteq u$
a. Then $\tau - \text{Cl}(A) = \{a\}^- = \{a, b\} \subseteq u$
(i.e. g-closed set)

(ii) Let $A = \{b\}$, $u = \{a, b, c\}$ such that $A \subseteq u$
a. Then $\tau - \text{Cl}(A) = \{b\}^- = \{a, b\} \subseteq u$
(i.e. g-closed set)

(iii) Let $A = \{c\}$, $u = \{a, b, c\}$ such that $A \subseteq u$
a. Then $\tau - \text{Cl}(A) = \{c\}^- = \{a, b\} \subseteq u$
(i.e. g-closed set)

(iv) Let $A = \{d\}$, $u = \{d, e\}$ such that $A \subseteq u$
a. Then $\tau - \text{Cl}(A) = \{d\}^- = \{d, e\} \subseteq u$
(i.e. g-closed set)

(v) Let $A = \{e\}$, $u = \{d, e\}$ such that $A \subseteq u$
a. Then $\tau - \text{Cl}(A) = \{d\}^- = \{d, e\} \subseteq u$
(i.e. g-closed set)

(vi) Let $A = \{a, b\}$, $u = \{a, b, c\}$ such that $A \subseteq u$
a. Then $\tau - \text{Cl}(A) = \{a, b\}^- = \{a, b\} \subseteq u$
(i.e. g-closed set)

(vii) Let $A = \{a, c\}$, $u = \{a, b, c\}$ such that $A \subseteq u$
a. Then $\tau - \text{Cl}(A) = \{a, c\}^- = \{a, b, c\} \subseteq u$
(i.e. g-closed set)

(viii) Let $A = \{c, d\}$, $u = \{d, e, c\}$ such that $A \subseteq u$
a. Then $\tau - \text{Cl}(A) = \{c, d\}^- = \{a, b\} \not\subseteq u$
(i.e. NOT g-closed set)

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(ix) Let $A = \{d, e\}$, $u = \{d, e, c\}$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{d, e\}^- = \{d, e\} \subseteq u$
 (i.e. g-closed set)

(x) Let $A = \{a, e\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{a, e\}^- = \{a, b, d, e\} \subseteq u$
 (i.e. g-closed set)

(xi) Let $A = \{a, c\}$, $u = \{a, b, c\}$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{a, c\}^- = \{a, b, c\} \subseteq u$
 (i.e. g-closed set)

(xii) Let $A = \{a, d\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{a, d\}^- = \{a, b, d, e\} \subseteq u$
 (i.e. g-closed set)

(xiii) Let $A = \{b, d\}$, $u = X$, such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{b, d\}^- = \{a, b, d, e\} \subseteq u$
 (i.e. g-closed set)

(xiv) Let $A = \{b, e\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{b, e\}^- = \{a, b, d, e\} \subseteq u$
 (i.e. g-closed set)

(xv) Let $A = \{c, e\}$, $u = \{d, e, c\}$ such that
 $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{c, e\}^- = X \not\subseteq u$
 (i.e. NOT g-closed set)

(xvi) Let $A = \{a, b, c\}$, $u = \{a, b, c\}$ such that
 $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{a, b, c\}^- = \{a, b, c\} \subseteq u$
 (i.e. g-closed set)

(xvii) Let $A = \{a, b, d\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{a, b, d\}^- = \{a, b, d, e\} \subseteq u$
 (i.e. g-closed set)

(xviii) Let $A = \{a, b, e\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{a, b, e\}^- = \{a, b, d, e\} \subseteq u$
 (i.e. g-closed set)

(xix) Let $A = \{a, c, d\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{a, c, d\}^- = X \subseteq u$
 (i.e. g-closed set)

(xx) Let $A = \{a, c, e\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{a, c, e\}^- = X \subseteq u$
 (i.e. g-closed set)

(xxi) Let $A = \{b, c, d\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{b, c, d\}^- = X \subseteq u$
 (i.e. g-closed set)

(xxii) Let $A = \{b, c, e\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{b, c, e\}^- = X \subseteq u$
 (i.e. g-closed set)

(xxiii) Let $A = \{c, d, e\}$, $u = \{d, e, c\}$ such that
 $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{c, d, e\}^- = X \not\subseteq u$
 (i.e. NOT g-closed set)

(xxiv) Let $A = \{d, e, a\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{d, e, a\}^- = \{d, e\} \subseteq u$
 (i.e. g-closed set)

(xxv) Let $A = \{d, e, b\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{d, e, b\}^- = \{a, b, d, e\} \subseteq u$
 (i.e. g-closed set)

(xxvi) Let $A = \{a, b, c, d\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{a, b, c, d\}^- = X \subseteq u$
 (i.e. g-closed set)

(xxvii) Let $A = \{a, b, c, e\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{a, b, c, e\}^- = X \subseteq u$
 (i.e. g-closed set)

(xxviii) Let $A = \{b, c, d, e\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{b, c, d, e\}^- = X \subseteq u$
 (i.e. g-closed set)

(xxix) Let $A = \{c, e, d, a\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{c, e, d, a\}^- = X \subseteq u$
 (i.e. g-closed set)

(xxx) Let $A = \{d, e, a, b\}$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \{d, e, a, b\}^- = \{a, b, d, e\} \subseteq u$
 (i.e. g-closed set)

(xxxi) Let $A = X$, $u = X$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = X^- = X \subseteq u$ (i.e. g-closed set)

(xxxii) Let $A = \phi$, $u = \phi$ such that $A \subseteq u$
 a. Then $\tau - \text{Cl}(A) = \phi^- = \phi \subseteq u$ (i.e. g-closed set)

Therefore, we have 29 g-closed sets, which are the subsets of the set $X = \{a, b, c, d, e\}$.

IV. CONCLUSION

Here we find 29 g-closed sets out of 32 subsets of $X = \{a, b, c, d, e\}$ with

$\tau = \{\phi, X, \{a, b, c\}, \{d, e\}, \{c\}, \{d, e, c\}\}$ as given below

$\phi, X, \{a\}, \{b\}, \{c\}, \{d\}, \{e\}, \{a, b\}, \{a, c\}, \{d, e\}, \{a, e\}, \{a, c\}, \{a, d\}, \{b, d\}, \{b, e\}, \{a, b, c\}, \{a, b, d\}, \{a, b, e\}, \{a, c, d\}, \{a, c, e\}, \{b, c, d\}, \{b, c, e\}, \{d, e, a\}, \{d, e, b\}, \{a, b, c, d\}, \{a, b, c, e\}, \{b, c, d, e\}, \{c, e, d, a\}, \{d, e, a, b\}$

Total : 29

Also following 3 subsets of X are NOT g-closed sets as given below:

{c, d}, {c, e}, {c, d, e}

Total : 03

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A Generalized Probability Distribution Pertaining To Product of Special Functions with Applications

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GJSFR Classification (FOR)

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Abstract- The aim of the present paper is to study a generalized probability distribution involving M-series and H-function. Here, we first obtain the distribution function for the probability density function and then apply the technique of convolution to obtain the distribution of sum of two independent random variables with p.d.f. involving the generalized hyper geometric function. The results obtained here are unified in nature and capable of yielding a very large number of corresponding results (new and known) involving simpler special functions and polynomials as special cases of our results.

Keywords- Probability density function, distribution function, H-function, M-series.

I. INTRODUCTION

In this paper we consider a general class of statistical probability distribution, having the probability density function

$$f(x) = \frac{x^{\lambda-1}}{C(1+\beta x)^\mu} {}_rM_s^{\alpha} \left[\frac{\omega x^\gamma}{(1+\beta x)^\eta} \right] \\ \times H_{p,q}^{m,n} \left[\frac{zx^\rho}{(1+\beta x)^\sigma} \left| \begin{matrix} (a_j, A_j)_{1,p} \\ (b_j, B_j)_{1,q} \end{matrix} \right. \right] \quad \dots(1)$$

for $0 < x < \infty$ and $f(x) = 0$ for other values of x and the constant C is given by

$$C = \beta^{-\lambda} \sum_{k=0}^{\infty} \frac{(u_1)_k \dots (u_r)_k}{(v_1)_k \dots (v_s)_k} \frac{w^k}{\Gamma(\alpha k + 1)} \beta^{-\gamma k} \\ \times H_{p+2,q+1}^{m,n+2} \left[\frac{z}{\beta^\rho} \left| \begin{matrix} (1-\lambda-\gamma k, \rho), (1-\mu-\eta k+\lambda+\gamma k, \sigma-\rho), (a_j, A_j)_{1,p} \\ (b_j, B_j)_{1,q}, (1-\mu-\eta k, \sigma) \end{matrix} \right. \right]. \quad \dots(2)$$

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The M-series introduced by Sharma (2008) is defined as

$${}_rM_s^{\alpha} (u_1, \dots, u_r; v_1, \dots, v_s, w) = \sum_{k=0}^{\infty} \frac{(u_1)_k \dots (u_r)_k}{(v_1)_k \dots (v_s)_k} \frac{w^k}{\Gamma(\alpha k + 1)}. \quad \dots(3)$$

For convergence conditions and other details of M-series, see Sharma (2008).

The H-function introduced by Fox (1961) is defined as

$$H_{p,q}^{m,n} \left[z \left| \begin{matrix} (a, A)_{1,p} \\ (b, B)_{1,q} \end{matrix} \right. \right] =$$

$$\frac{1}{2\pi i} \int_{-i\infty}^{+i\infty} \frac{\prod_{j=1}^m \Gamma(b_j - B_j \xi) \prod_{j=1}^n \Gamma(1 - a_j + A_j \xi)}{\prod_{j=m+1}^q \Gamma(1 - b_j + B_j \xi) \prod_{j=n+1}^p \Gamma(a_j - A_j \xi)} z^\xi d\xi. \quad \dots(4)$$

For convergence conditions and other details of H-function see Fox (1961).

The following conditions are assumed to be satisfied

(i) $\mu > \lambda + \gamma k > 0, \beta > 0, \sigma \geq \rho > 0,$

(ii) $\lambda + \gamma k + \rho \min_{i \leq j \leq m} \left(\frac{b_j}{B_j} \right) > 0,$

(iii) $(\lambda + \gamma k - \mu - \eta k) + (\rho - \sigma) \min_{i \leq j \leq n} \left(\frac{a_j - 1}{A_j} \right) < 0,$

(iv) $\Delta = \sum_{j=1}^m B_j - \sum_{j=m+1}^q B_j + \sum_{j=1}^n A_j - \sum_{j=n+1}^p A_j > 0,$

(v) $r \leq s, |w| < 1.$

(vi) The parameters involved in (1) are real and so restricted that $f(x)$ remains positive for $0 < x < \infty$ (5)

The p.d.f. $f(x)$ given by (1) is generalization of the generalized F-distribution defined by Malik (1967). It is also shown that the p.d.f. defined by Mathai and Saxena (1971) is particular case of the p.d.f. $f(x)$ given by (1).

II. THE DISTRIBUTION FUNCTION

$$\begin{aligned}
 F(x) &= \frac{1}{C \beta^\lambda} \sum_{k=0}^{\infty} \frac{(u_1)_k \dots (u_r)_k}{(v_1)_k \dots (v_s)_k} \frac{w^k}{\Gamma(\alpha k + 1)} \\
 &\times \frac{1}{\beta^{\gamma k}} \frac{1}{2\pi i} \int_{-i\infty}^{+i\infty} \left(\frac{z}{\beta^\rho} \right)^\xi \phi(\xi) d\xi \\
 &\times \int_0^{\frac{\beta}{1+\beta x}} y^{\lambda + \gamma k + \rho \xi - 1} (1-y)^{\mu + \eta k + (\sigma - \rho) \xi - \lambda - \gamma k - 1} dy \\
 , \dots
 \end{aligned} \tag{7}$$

where C is given by (2) and

$$\phi(\xi) = \frac{\prod_{j=1}^m \Gamma(b_j - B_j \xi) \prod_{j=1}^n \Gamma(1 - a_j + A_j \xi)}{\prod_{j=m+1}^q \Gamma(1 - b_j + B_j \xi) \prod_{j=n+1}^p \Gamma(a_j - A_j \xi)} \dots \tag{8}$$

Further, writing the incomplete beta function occurring in (7) in terms of Gauss hyper geometric function using a result (Erdélyi et al. (1954, p.87)), applying Euler's transformation formula (Erdélyi et al. (1954, p.64, eq.(23)))

$$F(x) = \frac{x^\lambda}{C(1+\beta x)^\mu} \sum_{k=0}^{\infty} \frac{(u_1)_k \dots (u_r)_k}{(v_1)_k \dots (v_s)_k} \frac{w^k}{\Gamma(\alpha k + 1)} \frac{x^{\gamma k}}{(1+\beta x)^{\eta k}}$$

The distribution function $F(x)$ for p.d.f. $f(x)$ is given by

$$F(x) = \int_0^\infty f(t) dt \tag{6}$$

Now, substituting the value of $f(t)$ from (1), expressing the M-series in series form and the H-function in the form given by (4), interchanging the order of summation and integration and putting $y = \beta t/(1+\beta t)$ in the resulting integral, we get

and expressing the result thus obtained in terms of H-function of two variables with the help of (Srivastava, Gupta and Goyal (1982, p.84, eq.(6.2.1))), we arrive at the following result after a little simplification

$$\times H_{1,1;p+1,q+1:1,1}^{0,1;m,n+1:1,1} \left[\begin{array}{c} \frac{zx^\rho}{(1+\beta x)^\sigma} \\ \frac{-\beta x}{1+\beta x} \end{array} \middle| \begin{array}{c} (1-\mu-\eta k:\sigma,1):(1-\lambda-\gamma k,\rho),(a_j,A_j)_{1,p};(0,1) \\ (-\lambda-\gamma k:\rho,1):(b_j,B_j)_{1,q},(1-\mu-\eta k,\sigma);(0,1) \end{array} \right] \dots \quad (9)$$

III. PARTICULAR CASES

(1) If we set $k = 0$, $\rho = \sigma$ and $\beta = 0$, we get the result obtained by Mathai and Saxena (1971, p.201, eq. (132)) and also by Srivastava and Singhal (1972, p.6, eq. (14)) after a little simplification.

$$f(x) = \begin{cases} \frac{x^{\lambda-1}}{C_1 (1+\beta x)^\mu} {}_pF_q \left[(a_p); (b_q); \frac{\beta x}{1+\beta x} \right] \\ 0, \text{ otherwise} \end{cases}$$

...

where

$$C_1 = B[\lambda, \mu - \lambda] \beta^{-\lambda} {}_{p+1}F_{q+1}[\lambda, (a_p); \mu, (b_q); 1], \quad \dots (11)$$

and the corresponding distribution function as obtained from (9) is given by

$$F(x) = \frac{x^\lambda}{C_1 \lambda (1+\beta x)^\lambda} {}_{1:p;1}F_{1:q;0} \left[\begin{array}{c} \lambda: a_1, \dots, a_p; \lambda - \mu; \frac{\beta x}{1+\beta x} \\ 1+\lambda: b_1, \dots, b_q; \dots; \frac{\beta x}{1+\beta x} \end{array} \right], \dots \quad (12)$$

provided that $|\beta x| < 1$ and the conditions easily obtainable from those stated in (5) are satisfied.

(3) On taking $\alpha = 1$ in (1), the M-series reduces to generalized hypergeometric function ${}_rF_s$ (Sharma (2008, p.189, eq. (5))), we get

$$f(x) = \begin{cases} \frac{x^{\lambda-1}}{C_2 (1+\beta x)^\mu} {}_rF_s \left[\frac{w x^\gamma}{(1+\beta x)^n} \right] \\ \times H_{p,q}^{m,n} \left[\frac{zx^\rho}{(1+\beta x)^\sigma} \middle| \begin{array}{c} (a_j, A_j)_{1,p} \\ (b_j, B_j)_{1,q} \end{array} \right], x > 0 \\ 0, \text{ otherwise} \end{cases}$$

...

(13)

where

$$\begin{aligned}
 C_2 &= \beta^{-\lambda} \sum_{k=0}^{\infty} \frac{(u_1)_k \dots (u_r)_k}{(v_1)_k \dots (v_s)_k} \frac{w^k}{k!} \beta^{-\gamma k} \\
 &\times H_{p+2,q+1}^{m,n+2} \left[\frac{z}{\beta^\rho} \left| \begin{matrix} (1-\lambda-\gamma k, \rho), (1-\mu-\eta k+\lambda+\gamma k, \sigma-\rho), (a_j, A_j)_{1,p} \\ (b_j, B_j)_{1,q}, (1-\mu-\eta k, \sigma) \end{matrix} \right. \right] \\
 , \quad &\dots(14)
 \end{aligned}$$

and the corresponding distribution function as obtained from (9).

$$\begin{aligned}
 F(x) &= \frac{x^\lambda}{C_2(1+\beta x)^\mu} \sum_{k=0}^{\infty} \frac{(u_1)_k \dots (u_r)_k}{(v_1)_k \dots (v_s)_k} \frac{w^k}{k!} \frac{x^{\gamma k}}{(1+\beta x)^{\eta k}} \\
 &\times H_{1,1;p+1,q+1:1,1}^{0,1; m,n+1 : 1,1} \left[\begin{matrix} \frac{zx^\rho}{(1+\beta x)^\sigma} \\ \frac{-\beta x}{1+\beta x} \end{matrix} \left| \begin{matrix} (1-\mu-\eta k, \sigma, 1); (1-\lambda-\gamma k, \rho), (a_j, A_j)_{1,p}; (0,1) \\ (-\lambda-\gamma k, \rho, 1); (b_j, B_j)_{1,q}, (1-\mu-\eta k, \sigma); (0,1) \end{matrix} \right. \right] \\
 , \quad &\dots(15)
 \end{aligned}$$

(4) If we set $r = s = 0$ in (1), the M-series reduces to Mittag-Leffler function (Sharma (2008, p.188, eq.(4))), we get

$$\begin{aligned}
 f(x) &= \left[\begin{matrix} \frac{x^{\lambda-1}}{C_3 (1+\beta x)^\mu} E_\alpha \left[\frac{w x^\gamma}{(1+\beta x)^\eta} \right] \\ \times H_{p,q}^{m,n} \left[\frac{zx^\rho}{(1+\beta x)^\sigma} \left| \begin{matrix} (a_j, A_j)_{1,p} \\ (b_j, B_j)_{1,q} \end{matrix} \right. \right] \end{matrix} \right], x > 0 \\
 &\quad [0, \text{ otherwise}]
 \end{aligned}$$

where

$$C_3 = \beta^{-\lambda} \sum_{k=0}^{\infty} \frac{w^k}{\Gamma(\alpha k + 1)} \beta^{-\gamma k}$$

... (16)

$$\times H_{p+2,q+1}^{m,n+2} \left[\frac{Z}{\beta^\rho} \right]_{(b_j, B_j)_{1,q}, (1-\mu-\eta k, \sigma)}^{(1-\lambda-\gamma k, \rho), (1-\mu-\eta k+\lambda+\gamma k, \sigma-\rho), (a_j, A_j)_{1,p}} , \dots (17)$$

and the corresponding distribution function is given by

$$F(x) = \frac{x^\lambda}{C_3(1+\beta x)^\mu} \sum_{k=0}^{\infty} \frac{w^k}{\Gamma(\alpha k + 1)} \frac{x^{\gamma k}}{(1+\beta x)^{\eta k}} \\ \times H_{1,1;p+1,q+1:1,1}^{0,1;m,n+1:1,1} \left[\begin{array}{c} \frac{zx^\rho}{(1+\beta x)^\sigma} \\ \frac{-\beta x}{1+\beta x} \end{array} \right]_{(-\lambda-\gamma k:\rho,1):(b_j, B_j)_{1,q}, (1-\mu-\eta k, \sigma);(0,1)}^{(1-\mu-\eta k:\sigma,1):(1-\lambda-\gamma k, \rho), (a_j, A_j)_{1,p};(0,1)} . \dots (18)$$

IV. THE DISTRIBUTION OF THE SUM OF TWO INDEPENDENT RANDOM VARIABLES

In this section, we shall obtain the distribution of the sum of two independent random variables with the p.d.f. as given by (10). By convolution formula, the distribution function $G(y)$ of $Y = X_1 + X_2$ is given by

$$G(y) = \int_0^y F_1(y - x_2) f_2(x_2) dx_2 , \dots (19)$$

where $F_2(x_2)$ is the p.d.f. of the variable X_2 and $F_1(y - x_2)$ is the distribution function of the variate X_1 . The result can be expressed in the form of following theorem:

Theorem 1. Let X_i ($i = 1, 2$) be two independent random variables with p.d.f. defined by

$$f_i(x_i) = \begin{cases} \frac{x_i^{\lambda_i-1}}{L_i (1+\beta_i x_i)^{\mu_i}} F_{p_i}^{q_i} \left[(a_{p_i}^{(i)}); (b_{q_i}^{(i)}); \frac{\beta_i x_i}{1+\beta_i x_i} \right], & x > 0 \\ 0, & \text{otherwise} \end{cases} \dots (20)$$

Where for $i = 1, 2$.

$$L_i = B[\lambda_i, \mu_i - \lambda_i] \beta_i^{-\lambda_i} p_i^{p_i} F_{q_i+1} \left[\lambda_i, (a_{p_i}^{(i)}); u_i(p_{q_i}^{(i)}); 1 \right] , \dots (21)$$

and the following conditions are assumed to be satisfied, for $i = 1, 2$

(i) $\mu_i > \lambda_i > 0, \beta_i > 0$,

(ii) $p_i \leq q_i$ or $p_i = q_i + 1$ and

$$(\mu_i - \lambda_i) + \sum_{j=1}^{q_i} (b_j^{(i)}) - \sum_{j=1}^{p_i} (a_j^{(i)}) > 0 ,$$

(iii) the parameters involved in (20) are so restricted that $f_i(x_i)$ remain positive for $0 < x_i < \infty$. Then the p.d.f. $g(y)$ of $Y = X_1 + X_2$ is given by

$$g(y) = \frac{y^{\lambda_1 + \lambda_2 - 1}}{L_1 L_2} B(\lambda_1, \lambda_2) F_{1:q_1;0;q_2+1;0;0}^{4:p_1;1;p_2;0;0}$$

$$\begin{bmatrix} \beta_1 y \\ \beta_1 y \\ \beta_2 y \\ -\beta_2 y \\ -\beta_1 y \end{bmatrix} \left| \begin{array}{l} (\lambda_1; 1, 1, 0, 0, 1), \quad (\lambda_2; 0, 0, 1, 1, 0), (\mu_2, 0, 0, 1, 1, 0), \\ \dots \end{array} \right.$$

$$(1 + \lambda_1; 1, 1, 0, 0, 1), ((a_{p_1}^{(1)}), 1), (\lambda_1 - \mu_1, 1), ((a_{p_2}^{(2)}), 1), \dots, \dots, \\ \dots, ((b_{q_1}^{(1)}), 1), \dots, ((b_{q_2}^{(2)}), 1), (\mu_2, 1), \dots, \dots \Bigg],$$

...(22)

$$|\beta_i y| < 1 \quad (i=1,2).$$

Proof. On substituting the values of $F_1(y-x_2)$ and $f_2(x_2)$ as obtained from the equations (18) and (20) respectively in equation (19), we get

$$G(y) = \frac{1}{\lambda_1 L_1 L_2} \int_0^y \frac{(y - x_2)^{\lambda_1} x_2^{\lambda_2 - 1}}{(1 + \beta_1(y - x_2))^{\lambda_1} (1 + \beta_2 x_2)^{\mu_2}}$$

$$F_{1:p_1;1}^{1:p_1;1} \left[\begin{array}{c} \lambda_1: a_1^{(1)}, \dots, a_{p_1}^{(1)}; \lambda_1 - \mu_1; \frac{\beta_1(y-x_2)}{1+\beta_1(y-x_2)} \\ 1+\lambda_1: b_1^{(1)}, \dots, b_{q_1}^{(1)}; -; \frac{\beta_1(y-x_2)}{1+\beta_1(y-x_2)} \end{array} \right]_{p_2} F_{q_2} \left[(a_{p_2}^{(2)}); (b_{q_2}^{(2)}); \frac{\beta_2 x_2}{1+\beta_2 x_2} \right] dx_2$$

... (23)

Now, expressing the generalized Kampé de Fériet function and the generalized hypergeometric function in series form using the results (Bryson (1974, p.27, eq. (28) and p.19, eq.

(23) and interchanging the order of integration and summations, then substituting $x_2 = yz$ in the resulting integral, and after a little simplification, we get

$$G(y) = \frac{y^{\lambda_1 + \lambda_2}}{\lambda_1 L_1 L_2} \sum_{r_1, r_2, r=0}^{\infty}$$

$$\begin{aligned}
 & \times \frac{(\lambda_1)_{r_1+r_2} \prod_{j=1}^{p_1} (a_j^{(1)})_{r_1} (\lambda_1 - \mu_1)_{r_2} \prod_{j=1}^{p_2} (a_j^{(2)})_r \beta_1^{r_1+r_2} \beta_2^{r_1+r_2+r}}{r_1! r_2! r! (1+\lambda_1)_{r_1+r_2} \prod_{j=1}^{q_1} (b_j^{(1)})_{r_1} \prod_{j=1}^{q_2} (b_j^{(2)})_r} \\
 & \times \int_0^1 z^{\lambda_2+r-1} (1-z)^{\lambda_1+r_1+r_2} [1+\beta_1 y(1-z)]^{-(\lambda_1+r_1+r_2)} (1+\beta_2 yz)^{-(\mu_2+r)} dz
 \end{aligned} \quad \dots(24)$$

Further, writing the integral occurring in right hand side of (24) in terms of Appell's function F_3 using a result (Srivastava and Karlsson (1985, p.279, eq. (18))), expressing the Appell's function F_3 in series form (Exton (1976, p.24,

eq.(1.4.3))) and after a little simplification, the distribution function $G(y)$ can be expressed in terms of generalized Lauricella function (Srivastava and Daoust (1969, p.454))) as follows

...(25)

Since $G(y)$ is the distribution function of the random variable Y , so the p.d.f. $g(y)$ of random variable Y is obtained by differentiating the expression $G(y)$ with respect to y and we get the desired result (22).

The result obtained here is quite general in nature and is capable of yielding a large number of corresponding results merely by specializing the parameters involved in it. To illustrate we give the following known special case of our result.

If we take the variables X_1 and X_2 have generalized F-distribution obtained by Malik (1967) then the density function of random variable Y can be easily obtained from equation (22). Further on integrating the result thus obtained from 0 to y with respect to y , we get the distribution function for the random variable Y , which is recently obtained by Dyer (1982, p.185, eq.(8.7)) in a slightly different form.

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Determination of copper (II) by simple extraction procedure prior to differential pulse polarography using 4-(2-hydroxy phenylethaminodiol) benzene-1,3-diol

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Abstract-Outline of the present paper proposes the determination of copper in biological samples, plants and pharmaceutical preparations using a newly synthesized analytical reagent 4-(2-hydroxy phenyl ethaminodiol), benzene-1,3-diol (4-2-HPEDB-1,3,D) to form a complex at pH 4.0 (acetate Buffer), which is extracted in to chloroform and the electrochemical behavior of chloroform extract was studied by differential pulse paleography under the optimum conditions. The linearity was maintained at the concentration range of 0.05 to 200 μ g/mL at pH 4.0 with correlation factor of 0.9997. The influence of sample matrix was investigated thoroughly which made the method more sensitive and selective. The present method was successfully applied for the determination of copper in biological samples, plants and pharmaceutical preparations and the results obtained from the proposed method show good agreement with reported method which is exist in literature. The precision, accuracy and validity of the method was checked by the investigating the Standard Reference Material (SRM) which is distributed by the National Institute of Standard and Technology (NIST).

Keywords- Copper, Differential pulse polarography, (4-2-HPEDB-1,3,D), biological samples, Plant material and pharmaceutical preparations.

I. INTRODUCTION

Generally copper is widely distributed in environment, food material and animal origins and more over it is exist in +2 oxidation state in several compounds. It plays important role in carbohydrates and lipid metabolism, due to this reason it is fact that many of the higher plants and animal life needs the trace amounts of copper as nutrient to survive. Copper has both essential and toxic element to human beings, however the high dosage of copper moderately causes toxicity and it will be responsible for the pointer to various diseases. For example the willson's disease arising in human beings by intake of high concentration of copper. Copper enters in to the humans, plants and animal bodies are mainly through food, air, water, rain, snow, irrigation water and fertilizers etc.

The other application of copper is used as thermal conductors, electrical conductors, building material and an important constituent of various metal alloys, in addition to

that copper has 29 distinct isotopes ranging from the atomic mass number from 52 to 80. Among this ^{63}Cu , and ^{65}Cu are stable and naturally occurring isotopes of copper remaining are radioactive isotopes. It also functions as a co-factor in various enzymes and in copper based pigments. In view of that it is necessary to monitoring the copper in different field areas. Several analytical reagents have been synthesized for the determination of copper in various samples which is shown in Table 1.

In early days various analytical methods are reported for the monitoring of copper like Spectrophotometry [1-6], Voltammetry [7] Atomic absorption spectrometry [8-10], Inductively coupled plasma atomic emission spectrometry [11-14] and Inductively coupled plasma mass spectrometry [15]. The above said methods have suffers some disadvantages like time consuming, and it require more sophisticated laboratory conditions, well expensive instrumentation.

The aim of the present work is to determined the copper in various samples at low concentration levels using differential pulse polarographic technique is to over come the drawbacks in the above said methods usually having insufficient for very low concentration level determination in samples.

Therefore authors proposed a novel, simple, selective, sensitive and new analytical method, differential pulse polarography for the determination of copper and it is a alternative method for the determination of copper in biological samples, plant and pharmaceutical preparations with newly synthesized analytical reagent which is very low expensive and synthesized at ordinary laboratory conditions.

II. Experimental

A. Apparatus

An Elico CL-362 model polarographic system is used for DPP measurements and Elico Li-129 Model glass-calomel combined electrode was employed for measuring pH values. Ag/AgCl (salt KCl) was used as a reference electrode which provide a reversible half reaction with nernstian behavior be constant over time and easy to assemble and maintain and a platinum wire as an auxiliary electrode which displays negative potential range.

B. Reagents

All reagents used were of analytical reagent grade. Double distilled water was used throughout the experiment. A stock solution of copper (II) was prepared by dissolving appropriate amount of copper sulphate in double distilled water in volumetric flask. Working standard solution was freshly prepared by diluting the stock solution with double distilled water. 0.1 M concentration of 4-2-HPEDB-1,3,D was prepared by dissolving 2.48 g of 4-2-HPEDB-1,3,D in 100 mL of methanol

C. Synthesis of 4-2-hydroxy phenyl ethaminodiol benzene-1,3-diol

(4-2-HPEDB-1,3,D)

Equimolar ratio of 2,4-dihydroxy acetophenone and 2-aminophenol in methanol mixture was refluxed for 3-4 hours and the contents were cooled at room temperature, it gives orange-red color precipitate. The precipitate was filtered and washed with methanol to pure Schiff base. (M.P-115 °C, yield 97%) as shown in Scheme I. I.R -(KBr), 1601.8cm⁻¹(C=N), 3304.0 cm⁻¹ (N-H), Free (OH) 3375.3 cm⁻¹ 1465.9 cm⁻¹(O-H) and the spectrum was shown in Figure 1.

D. Recommended analytical Procedure for the determination of metal ions

An aliquot of working standard solution containing 1-100 μ L of metal ion is taken in to 25 mL volumetric flask. To this 5 mL of acetate buffer solution (pH 4.5), 2mL of reagent solution were added. The mixture was shaken with 5.0 mL portion of chloroform for 30s and allowed to stand for 5-10 min. The organic phases curve collected and transferred in to polarographic cell and diluted with 9 mL of supporting electrolyte and then deoxygenated with nitrogen gas for ten min. After recording polarogram small increment (0.2mL) of standard solution is added to the cell treated for 1min, and polarogram is again recorded under similar conditions. In the same manner, 10 polarograms are recorded for 10 additions.. In the present study the best precision was obtained at pH 4.0 with a drop time 2 sec, pulse amplitude of 50 mV and an applied potential of -420.0 mV. The relative standard deviation and correlation coefficients were found to be 5.45% and 0.9997 respectively for 5 replicants.

E. Analysis of pharmaceutical preparations

A tablet or an appropriate aliquate of each (Supradyn and Multivitamin Iron Mineral tablets.) were grounded well and added 2-3 mL of nitric acid for dissolution and transfer in to a 100 mL calibrated flask, diluted to the mark with distilled water and mixed well and analyzed for copper by the above said general procedure. The results were given in the Table 2.

F. Analysis of biological samples

The hair samples were washed with acetone 2-3 times in a beaker with continuous stirring. Then they were dried in an electric oven at 70° C for 4 h. Two grams of the sample was weighed and taken in a beaker. To this a (1:1) mixture of

nitric acid and perchloric acid was added, and the mixture was heated on a hot plate. The solution was evaporated to near dryness. The ash was taken up with 5 mL of HCl (1+9) and evaporated to dryness. The residue was taken up in 2 mL conc. HCl, filtered and made up to 25 mL with water. Suitable volumes of these solutions are taken for the determination of copper as described in above said procedure and the results were shown in Table 2.

G. Analysis of plant material

The cabbage and banana samples (5 g each) were placed in a 250 mL beaker, and a solution of concentrated H₂SO₄ / HNO₃ 1:1 (v/v) (10 mL) was added. This mixture was heated until the solution is clear. The solution was filtered off and concentrated to 5 mL then cooled and diluted to 50 mL with deionised double-distilled water then the general procedure was applied to 1 mL of this solution and results obtained were shown in Table 2.

The accuracy and precision of the present method was validated by taking the Standard Reference Material SRM-1573a of tomato leaves, which is distributed by National Institute of Standard and Technology. Inter calibration was performed by using tomato leaves-SRM 1573a and it's certified value is 4.7 ppm. The analytical results obtained from the present method (4.68±0.05) shows the good agreement with certified value (4.7 ppm) of the tomato leaves.

III. RESULT AND DISCUSSION

DIFFERENTIAL PULSE POLAROGRAPHIC STUDIES

A. Effect of pH

The effect of pH on the peak potential E_p and current intensity i_p, using differential pulse polarography was examined for [Cu-(4-2-HPEDB-1,3,D)]. The pH was varied in the whole pH range 2.5 to 10.5 for [Cu-(4-2-HPEDB-1,3,D)] complex. It can be observed from Figure 2, - 460.0 mV that the maximum peak current obtained with pH 4.0. When the pH has been increased from 2.5 to 10.5 the peak potentials have been shifted towards more negative values, indicating proton participation in the reduction process and the results were shown in Figure 3.

B. Effect of Pulse amplitude and Scan Rate

The influence of the pulse amplitude was investigated. The results suggested that DPP peak current reached the maximum value when the pulse amplitude was 50 mV. As for the scan rate; the current response with increasing the scan rate of 12 mVs⁻¹ gave the maximum response. Accordingly, the optimum conditions for recording a maximum developed and sharper DPP peak for 0.05 mM [Cu-(4-2-HPEDB-1,3,D)] are scan rate : 12 mVs⁻¹ and pulse amplitude : 50 mV.

Other experimental parameters such as temperature and ionic strength were optimized. The stripping peak currents were not modified when the temperature varied between 20-50°C. The value chosen was 25°C.

C. Effect of Solvent

The extraction of [Cu-(4-2-HPEDB-1,3,D)] complex was carried out with different organic solvents like dimethyl formaldehyde, CCl₄, Cyclohexane, chloroform, xylene,

toluene, n-butanol, 1-pentanol, 1-amyl alcohol and nitrobenzene. Among these solvents the extraction of [Cu-(4-2-HPEDB-1,3,D)] complex efficiency more in chloroform when compare to other organic solvents. Therefore chloroform is chosen as solvent for extraction of [Cu-(4-2-HPEDB-1,3,D)] complex for further studies.

D.Calibration

The detection limit and the relative standard deviation obtained as 0.45 $\mu\text{g/mL}$, 5.45% respectively. The linearity is maintained in the concentration range of Cu 0.05 to 200 $\mu\text{g/mL}$ with correlation coefficient of 0.9997 and calibration curve is prepared according to the general procedure under the optimized conditions as shown in Figure 4.

The Method of Quantification (MOQ) value was calculated based on calibration curve of analytical procedure. The MOQ values found to be 6.920 $\mu\text{g/mL}$ for copper determination.

E .Stoichiometry of the complex

The composition of the complex was found to be 1:1 = Cu^{2+} : 4-2-HPEDB-1,3,D . The Stoichiometry of the complex was verified by Mole ratio method and Job's continuous variation method and their data was shown in Figure 5 and 6.

F. Effect of Foreign ions

The effect of interfering ions on the determination of copper in biological samples, Plant material and pharmaceutical preparations was investigated and the results were shown in Table 3 which is individually added to the copper having appropriate concentration and the general procedure was applied. The tolerable limits of various foreign ions are masked using suitable masking agents and recovery ranges (<2%) are shown in Table 3. The results are almost quantitative in the presence of interfering ions to evaluate the feasibility and sensitivity of the present method.

IV. CONCLUSION

The present method was successfully applied for the determination of copper in biological samples, plant material and pharmaceutical preparations. The Differential pulse polarographic method, after chloroform extraction procedure, for the assay of the copper determination using newly synthesized analytical reagent was reported and the results were compared with reference method with good agreement. The method has added advantages over reported methods i.e.

1. The organic reagent was less expensive, economical and it was synthesised at ordinary laboratory atmospheric conditions.
2. The organic reagent synthesised for the present work is very distinct in terms of selectivity, sensitivity towards metal ions.
3. The risk of contamination is very low and foreign ions do not interfere in the present method during the analysis of copper.

4. Electrodes used in the present work for the analysis of metal in samples is very sensitive and selective.
5. Statistical analysis and eliminating of time taking process, lengthy extraction steps makes the methods more sensitive and selective one for the determination of copper in biological samples, Plant material and pharmaceutical preparations.
6. The validity of proposed method was checked by standard reference material (SRM) tomato leaves 1573a which is given by National Institute of Standard and Technology which show the method have more accuracy and precision.

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Table 1. Comparison of the reagents for the determination of copper with other electro analytical techniques

S. No	Reagents	Electro analytical technique	Concentration	Samples	Reference
1.	Mercapto acid	Voltammetry	8×10^{-7} to 1×10^{-5} mol/L	Real environmental samples	[7]
2.	1-(2-thiazolylazo)-2-naphthol.	Differential pulse polarography	0.18–13.5 and 0.30–17.3	Various samples	[16]
3.	crown ethers	Voltammetry	Up to 100 ppb	Alcoholic beverages	[17]
4.	SSA	Adsorptive stripping voltammetry	$3 - 23 \mu\text{g L}^{-1}$	Crude oil, crude oil tank button sludge	[18]
5.	Nitroso-R dopant anion	Anodic stripping voltammetry	1.2 to 243.9 ng mL ⁻¹	Water and human hair samples	[19]
6.	-	Anodic stripping voltammetry	100 to 300 ng	Beer	[20]
7.	Alizarin red S (ARS)	Catalytic-Adsorptive Stripping Voltammetry	1.7×10^{-3} mol/L	Real samples	[21]
8..	4-2-HPEDB-1,3,D	Extractive differential pulse polarography	0.05 to 200 $\mu\text{g/mL}$	Biological samples, pharmaceutical and plant material	Present method

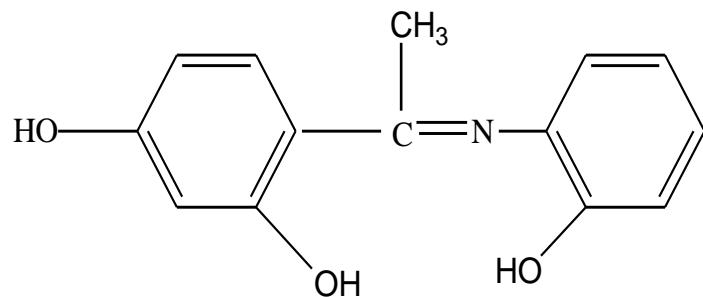
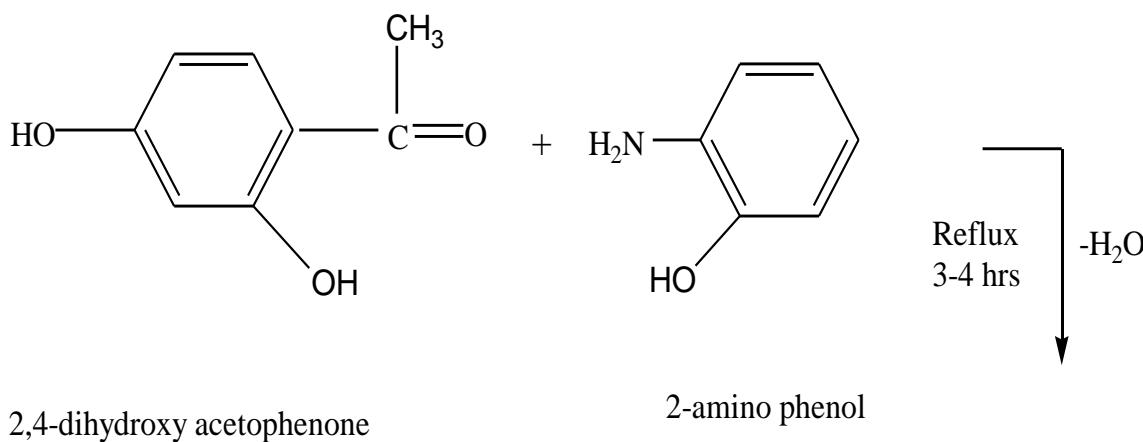
Table 2. Determination of copper in Biological samples, pharmaceutical preparation and plant material

S.NO	Samples	Present method	Reported method [6]	R.S.D
1.	Pharmaceutical preparations			
	Supradyn Certified value(0.86)	0.86.5	0.86	0.35
	Multivitamin Iron minerals Certified value(0.25)	0.27	0.26	0.16
2.	Human hair			
	A	46.52	45.11	0.04
	B	69.12	68.98	0.04
	C	63.99	61.81	0.02
3.	Plant material			
	Cabbage	38.75	37.66	0.11
	Banana	16.05	14.11	0.57

Table 3. Effect of Foreign ions in the determination of Copper

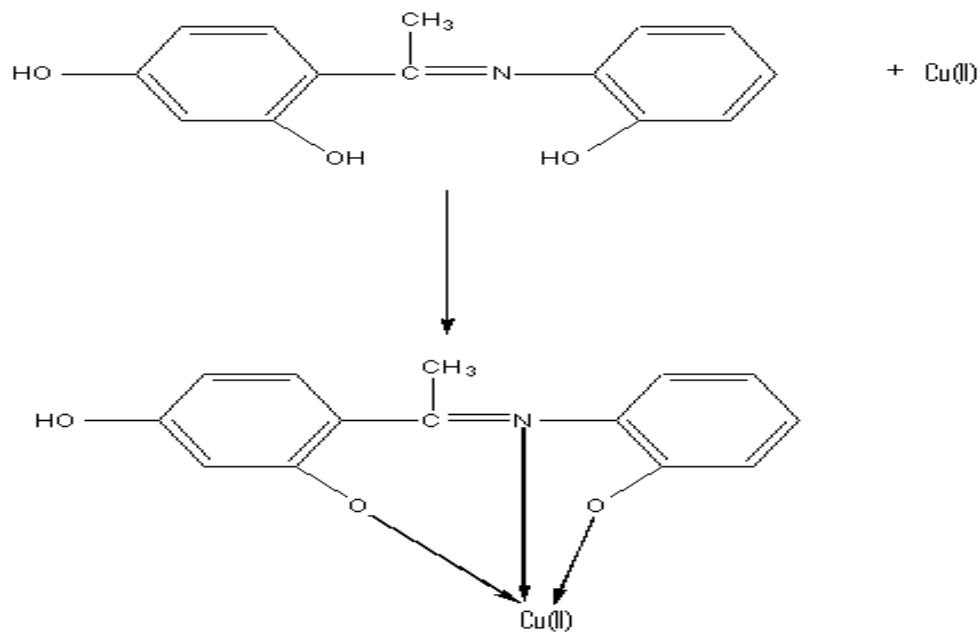
S.No	Ions	Concentration $\mu\text{g/mL}^{\text{a}}$
1	SCN^- , $\text{C}_2\text{O}_4^{2-}$, Cd^{2+} , Mg^{2+} , Cr^{3+} ,	300
2.	Zn^{2+} , Tartarate ion	100
3	Al^{3+} , Fe^{3+} , CN^-	50
4	Co^{2+} , Ni^{2+} , NO_3^- , Pb^{3+}	10

^a It can be masked by using 2mL of 2% sulphuric acid.



4-(2-hydroxy phenyl ethaminodiol), benzene-1.3-diol

Synthesis of 4-(2-hydroxy phenyl ethaminodiol) benzene-1,3-diol (4-2-HPEDB-1,3,D)



Complexation of 4-2-HPEDB-1,3,D with Cu

Scheme I. Synthesis and complexation of 4-2-HPEDB-1, 3,D with Cu

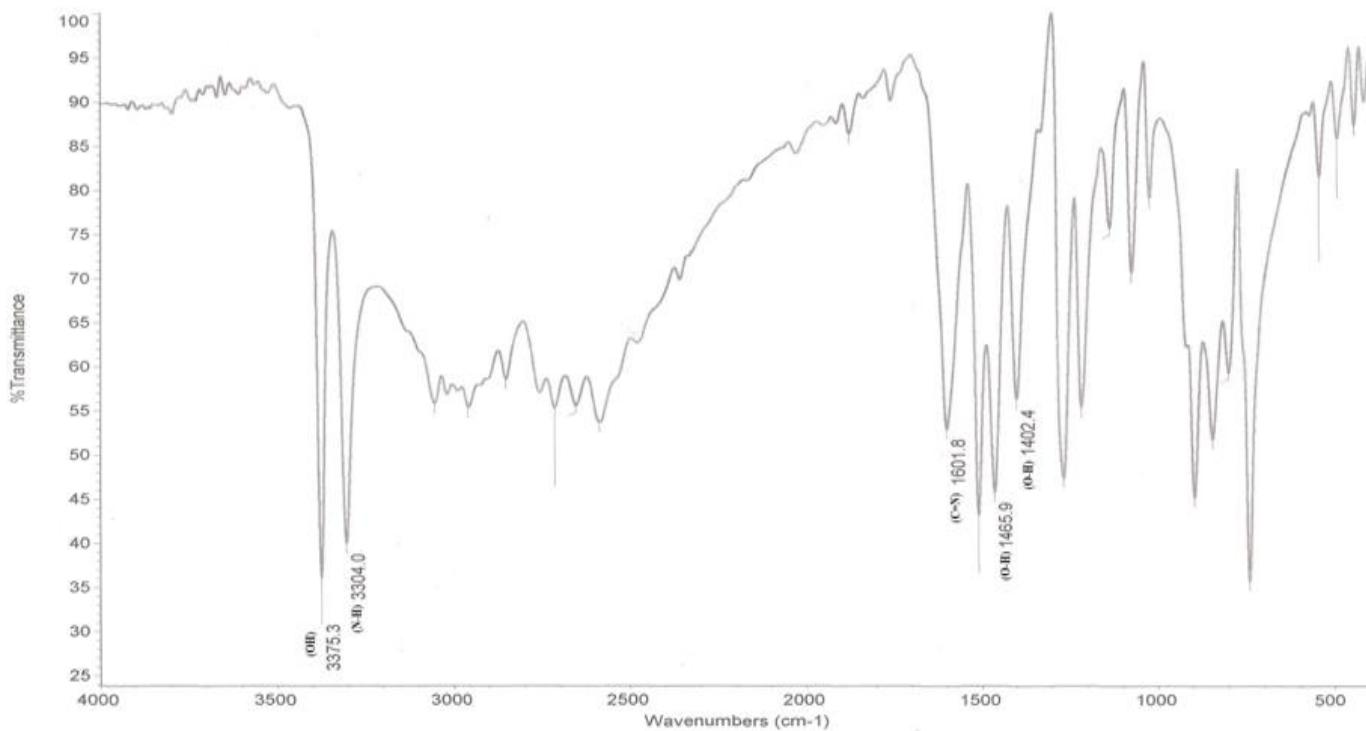
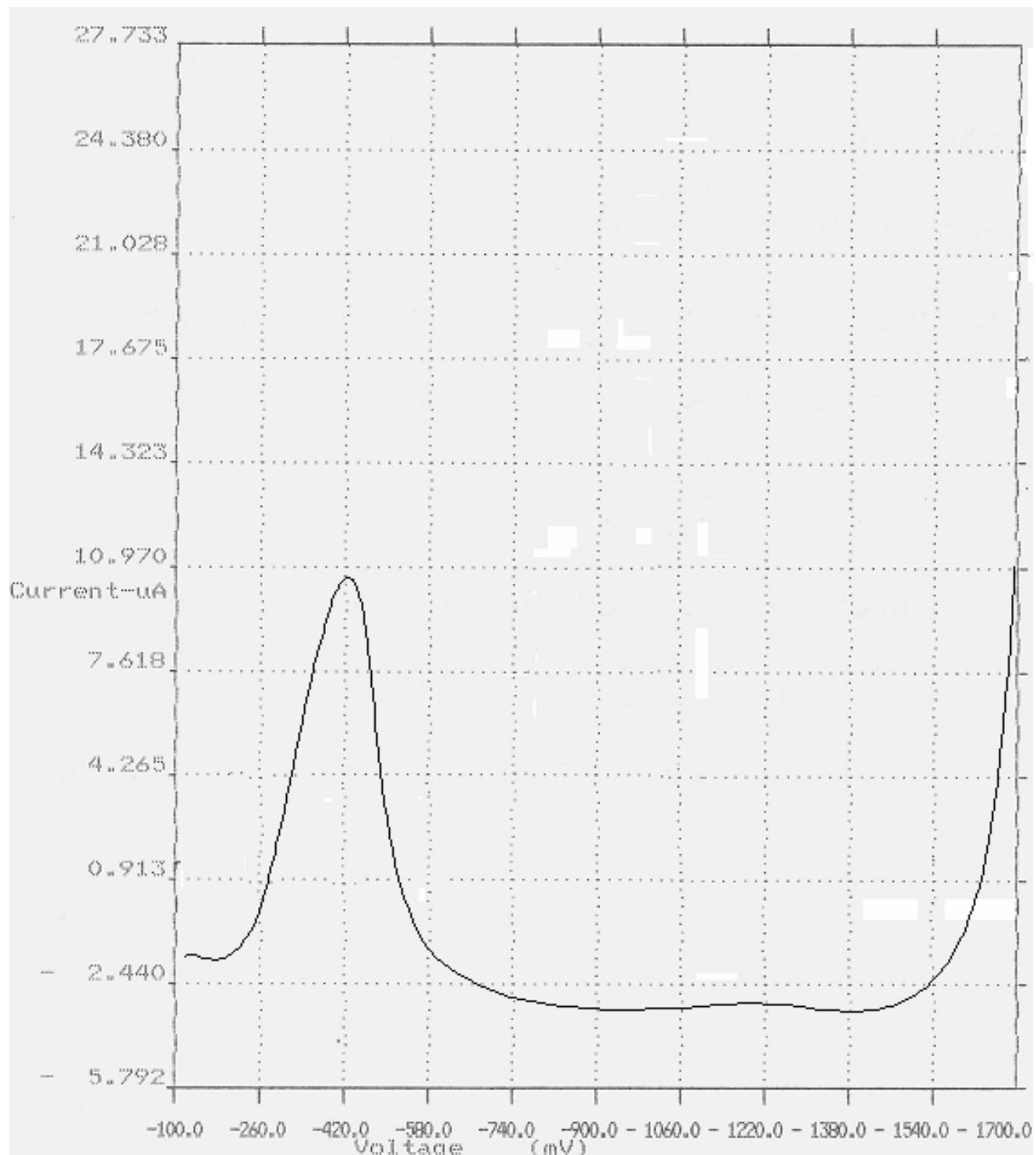


Fig 1. I.R spectrum for 4-(2-hydroxy phenyl ethaminodiol) benzene-1,3-diol (4-2- HPEDB-1,3,D)



**Fig 2. Differential pulse polarogram of Cu (II). Peak at - 420.0 mV
at pH 4.0; Cu-(4-2-HPEDB-1,3,D) concentration :1% of 1 ml
Scan rate : 12 mVs⁻¹ pulse amplitude :50 mV**

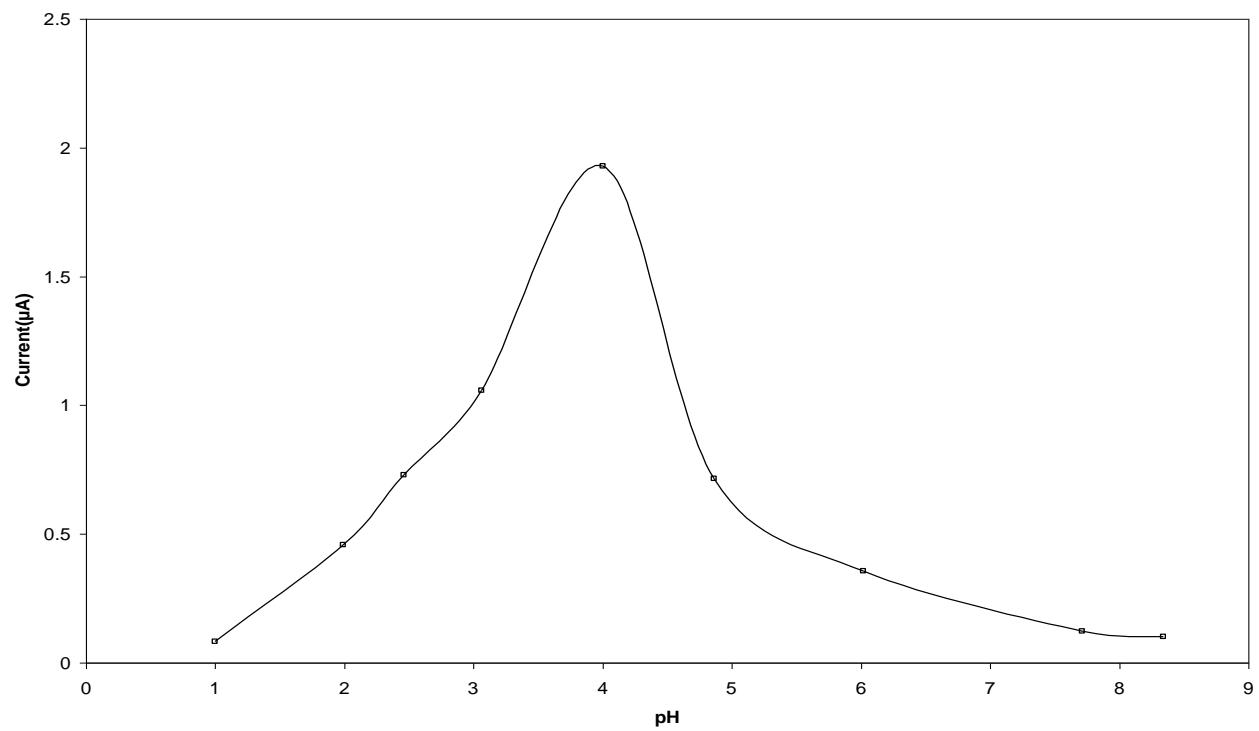


Fig 3. Effect of pH on determination of Cu (II)

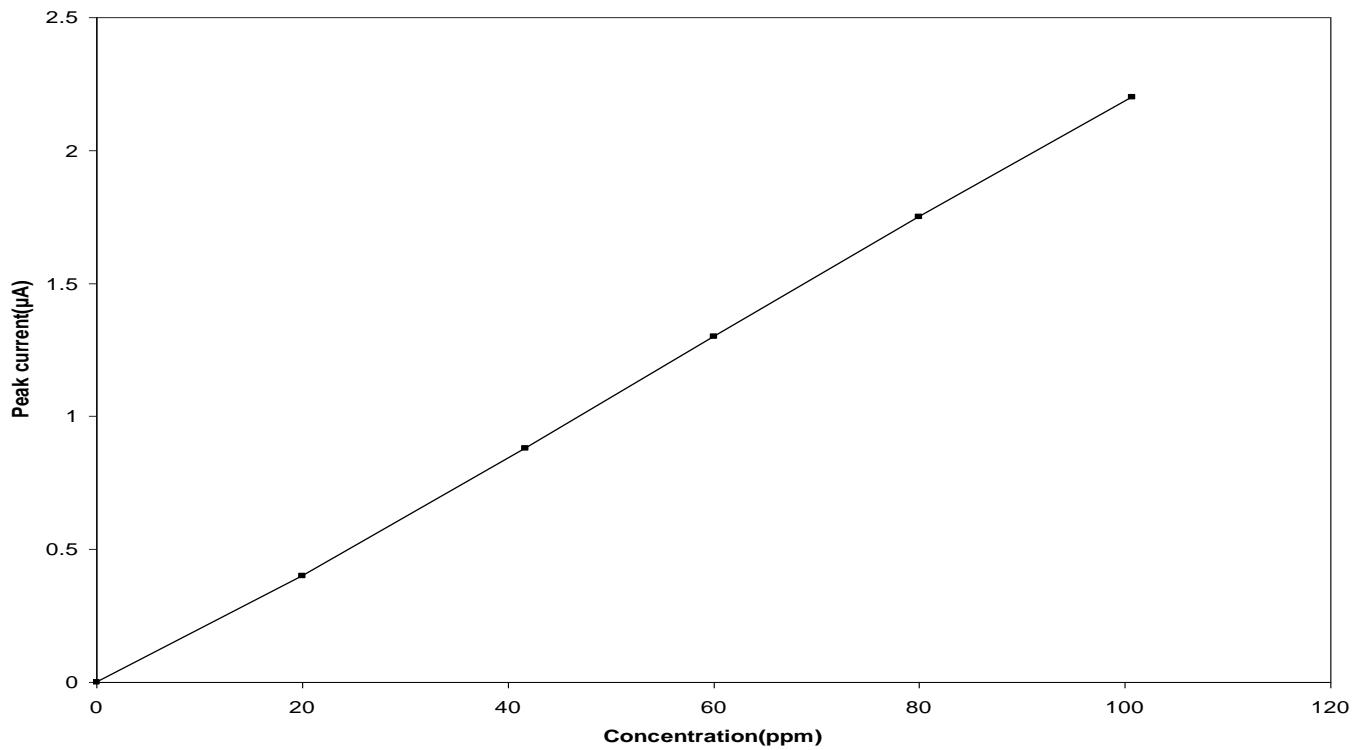


Fig 4. Calibration curve for Cu(II)

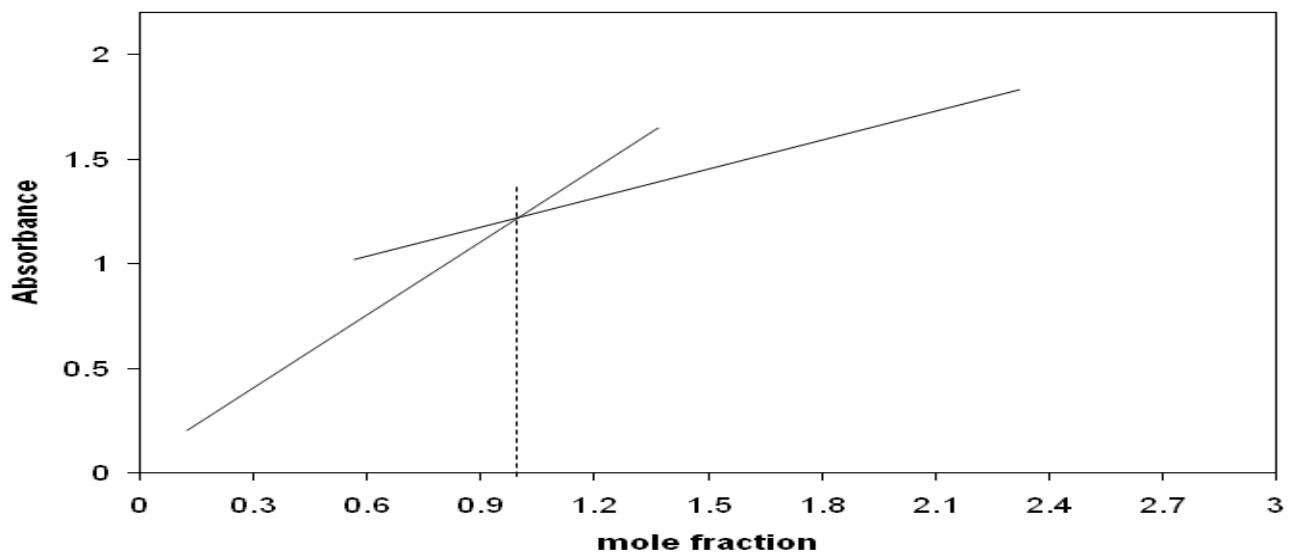


Fig.5. Mole ratio method for the Cu(II) - 4-2-HPEDB-1,3,D

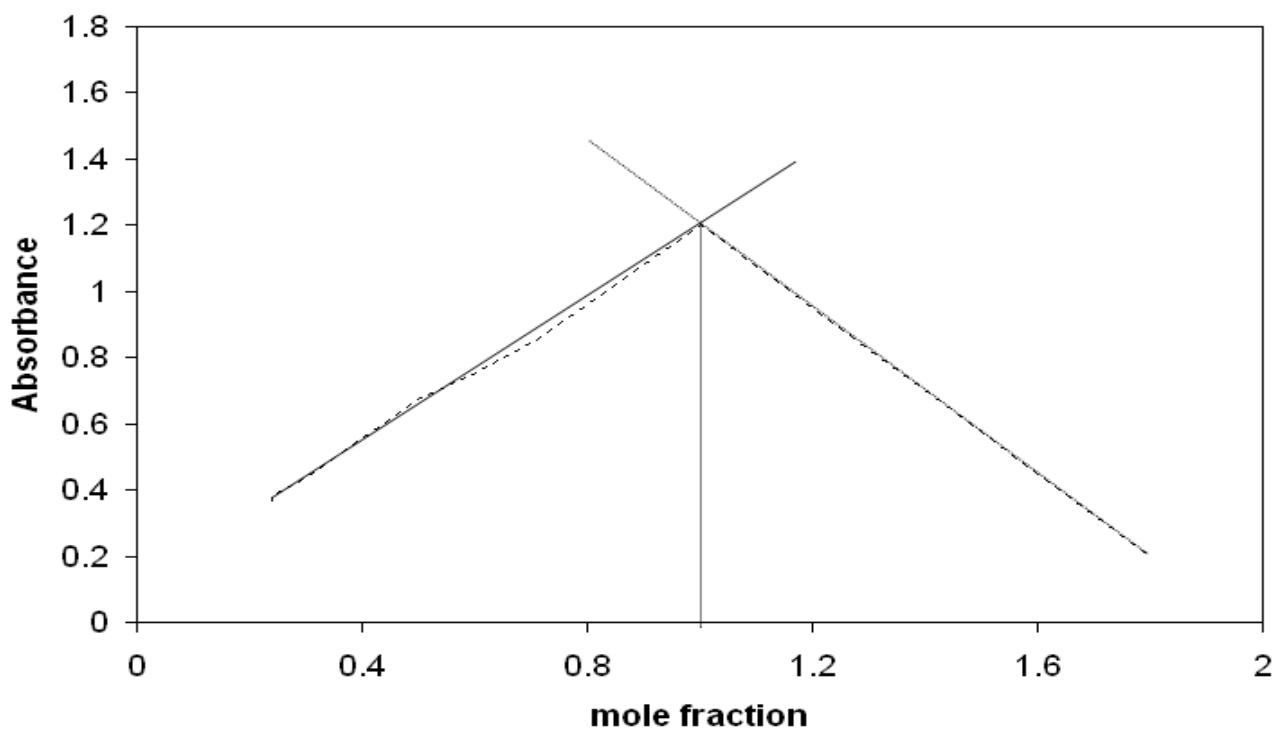


Fig.6. Job continuous variation method for the Cu(II) - 4-2-HPEDB-1,3,D

Refraction as A Function of Atomic Gravitation

Fresh Insight into the Nature of Space and A New Possibility In The Creation Of Metamaterials

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GJSFR Classification (FOR)
240301.240203.240504

Abstract- The paper states and illustrates what it claims as a new discovery, namely, that the refractive index of a material is directly proportional to the material's atomic gravitation. Then there follows three revolutionary implications of this discovery: Refraction is explained as a simple relativistic phenomenon; space (the fabric of the universe) is now seen to consist of the gravitational and the electromagnetic fields moving at c relative to each other, i.e., these fields are not in space, they are space; and, finally, atomic gravitation (rather than relative permittivity and relative permeability) is offered as the guiding principle in the construction of meta materials of different refractive indices.

I. INRODUCTION

Having discovered a direct proportionality between the refraction of electromagnetic waves through a medium and the gravitational field of the medium's atoms, I now present not only this discovery but also my own humble interpretation of it in the context of the theory of general relativity and the creation of meta materials in the rapidly expanding realm of nanotechnology

II. DISCOVERED

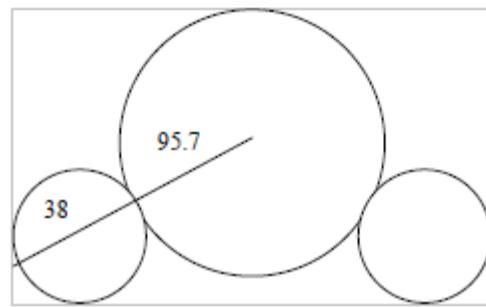
For a given wavelength, the refractive index of a medium, n, is directly proportional to the gravitational field at r where r is the radius of the representative particle of the medium's constituent building blocks. Thus $n = kGm/r^2$ where k is about 1.25×10^{16} ; G = universal gravitational constant; m and r are the mass and radius of the representative particle.

III. ILLUSTRATION

A. Water

Disregarding, for simplicity, the continual combining and separating of water molecules due to intermolecular hydrogen bonding, we may take the water molecules the representative particle of the medium.

Now in the water molecule we find: H-O bond length = 95.7pm., H – O – H angle is 104.5° [1]; the covalent radii of O and H are, respectively, 73pm and 38 pm [2]; and the atomic weights of O and H are, respectively, 16 and 1 [3]. We can therefore regard the water molecule as a sphere of $r = 95.7 + 38 = 133.7$ pm. (see diagram below where we take the nucleus of oxygen as a good approximation to the molecular center of mass)



So $n =$

$$kGm/r^2 = (1.25 \times 10^{16} \times 6.67 \times 10^{-11} \times 18 \times 1.66 \times 10^{-27}) / (1.3372 \times 10^4 \times 10^{-24})$$

= 1.39. This is quite close to the expected 1.33 [4] considering the simplifications above.

B. Sodium Chloride (table salt)

In this crystal the radii of Na and Cl ions are respectively, 98 pm and 181 pm [2]; atomic weights of Na and Cl are respectively 23 and 35.5 pm [3]; further, each Na ion is surrounded by six Cl ions (as each Cl ion is neighbored by six Na ions) [5]. So we may take, as our representative particle of this medium, the Na ion with its coordinate six Cl ions. This particle with its center in the Na nucleus extends through a radius $r = 98 + 181 + 181 = 460$ pm.

So, $n = (1.25 \times 10^{16} \times 6.67 \times 10^{-11} \times (6 \times 35.5 + 23) \times 1.66 \times 10^{-27}) / (4.62 \times 10^4 \times 10^{-24}) = 1.54$. This is the value expected! [4].

C. Diamond

Diamond affords such a close packing of carbon atoms that, for a first approximation, we can take the single carbon atom as our representative particle. Given the covalent radius of carbon as 77 pm [2] and its atomic weight as 12 [3], we have:

$$n = (1.25 \times 10^{16} \times 6.67 \times 10^{-11} \times 12 \times 1.66 \times 10^{-27}) / (7.72 \times 10^2 \times 10^{-24})n = 2.80 - \text{which, for a first approximation, is reasonably close to the expected value of 2.42}$$

[4]. For a more accurate value, let us observe the following: Each carbon atom in diamond is the centre of a tetrahedron whose vertices are four carbon atoms covalently bonded to the central one with a bond length of 154 pm [6]; the atom at each vertex is also the centre of a tetrahedron. In this way

these tetrahedral units join up into a huge diamond crystal. Now it so happens that the representative particle for diamond is a tetrahedron with its first generation tetrahedra, i.e., a tetrahedron whose four vertices are centres of four more tetrahedra. Therefore this unit has $(1+4\times 5)$ atoms and a radius of $154+154+77=385$ pm. This gives us $n=(1.25\times 1016\times 6.67\times 10^{-11}\times 21\times 12\times 1.66\times 10^{-27})\div(385\times 10^{-24})=2.35$. The expected value is 2.42 [6].

D. Carbon disulphid

The data for this molecule is as follows : carbon-sulphur bond length = 155 pm [7]; covalent radii of carbon and sulphur are, respectively, 77pm and 102 pm[2]; atomic weights for carbon and sulphur are 12 and 32 [3].

We can therefore consider this molecule as a sphere of $r = 155 + 102 = 257$ pm. This gives us $n = 1.25 \times 1016 \times 6.67 \times 10^{-11} \times 76 \times 1.66 \times 10^{-27} \div (2.572 \times 104 \times 10^{-24}) = 1.60$. The expected value is 1.62 [4].

Of course another way of illustrating all this is to plot a graph of n against Gm/r^2 for a chosen number of substances; however, the above presentation was deliberately chosen to demonstrate the procedure of identifying the representative particle in a medium. What we must emphasize, in addition to these few examples, is that the chemistry as well as the atomic, molecular or ionic geometry resulting from this chemistry must be given particular attention before we can discern the representative particle and its gravitation. When dealing with ionic compounds, for instance, we must realize that pure ionic bonds do not exist [8] and that the degree of covalency in the compound under investigation may warrant consideration; and when we come to examining the refractive index of metals in this light we have to take into account the peculiarities of metallic bonding as we try to establish the appropriate representative particle of the medium. Especially in connection with atomic radii, consideration must be given to the environment of an atom or molecule: for example, due to large intermolecular spaces in air, the atoms in its molecules will tend to assume the irrespective van der waals radius values.

IV. INTERPRETATION

$$OF\ n = kGm/r^2$$

A. Explaining refraction

This has been the subject of heated scientific and philosophical debate from ancient times down to this day. Reviewing the debate – before offering another problematic explanation – an expert in this field commented “Every explanation of refraction has some problems that have not yet been overcome” [9].

To claim, as we do by the above equation, that the refraction of electromagnetic waves through different materials is the function of gravitation is to remind us of the general relativity theorists who employ the idea of gravity causing space time-curvature to explain the bending of stellar light by massive bodies .We interpret the equation as claiming that refraction is an entirely relativistic phenomenon.

However, we shall not use this idea of gravitational space-time curvature; we only need one axiom of relativity, namely, that gravity causes *time dilation* and *length contraction*. Since a gravitational field dilates time while contracting length, it is easy to see why an observer will find the speed of light in glass to be less than the speed of light in air: If light travels at 3×10^8 metres in 1 second in air, this distance of 3×10^8 metres undergoes gravitational contraction in glass to a lower value (due to the gravitational field set up by glass atoms). This value turns out to be 3×10^8 metres divided by n , where n is the refractive index of glass. Since this 1 second is measured in air outside the glass slab the speed is found to be $(3 \times 10^8$ metres divided by n) per second. Of course if the 1 second is measured inside the glass slab where there is gravitational dilation (time ticks slower) the speed of light would be found to be c (or 3×10^8 m/s). Let us explain how: When 1 second elapses outside the glass slab, it is still (1 divided by n) seconds in the glass slab. So the speed of light for an observer in the glass slab (were this possible) is $(3 \times 10^8 / n) / (1/n) = 3 \times 10^8$ metres per second.

B. Fresh insight into the inside of atoms and molecules

The equation $n = kGm/r^2$ suggests that for the phenomenon of refraction, the only region that matters is the circumference of the representative particle. In the case of diamond, for example, electromagnetic radiation appears to suffer refraction only as it crosses the circumference of the carbon atom. What happens to the radiation as it traverses regions of greater gravity at points between the nucleus and the circumference? The only way to explain this is to postulate regions in atoms and molecules where the gravitational field = 0 so that $n = 0$ so that radiation in these regions travels at infinite velocity. Infinite speed of electromagnetic radiation means that, in such regions, a photon simply finds itself everywhere *in no time*; for a photon, in other words, the whole of such a region is simply a point! Now a photon is an electromagnetic field and an electromagnetic field is nothing but an electric field in motion. It therefore seems plausible to conclude that, for a photon, an electric field (in which it finds itself) is a single point. So the region in the atom where this happens (where $n = 0$ and gravity = 0) must be the electric field between the positively charged nucleus and the negatively charged electrons at the circumference.

So when a photon arrives at a point on the circumference of the representative particle it instantaneously finds itself on the opposite side of the circumference. (Remember that the atomic sphere or its outermost shell is a fuzzy band of electrons. A medium consisting of many atoms will offer many such bands to cross; these all add up to one medium of that particular refractive index- like many very thin glass slabs held tightly together).

C. Where to find “gravitons”

As said above, for a photon the electric field is simply one point and that, strictly speaking, the photon cannot be said to propagate ‘through’ an electromagnetic field. It is only in a gravitational field where (according to an observer outside

this field) the photons move with a speed governed by $v = c/n$ where $n = k E$. (E being the gravitational field forming the medium) Similarly, we now postulate, a graviton can only be observed propagating through an electromagnetic field, not through a gravitational field. Experiments with gravitons, so far, have not been as successful as expected and we think this failure is to be attributed to the fact that scientists have been trying to see the propagation of these particles at the wrong place, namely, in a gravitational field. The emerging picture of the interaction between the two fields is this: either we have photons flying through the gravitational field at c or gravitons flying through the electromagnetic field at c : where the gravitational field is zero the electromagnetic field reduces to a point as far as photons are concerned; but for gravitons the point is a universe in which they are flying at c . Where the electromagnetic field is zero the gravitational field reduces to a point as far as gravitons are concerned; but for photons this point is a universe in which they fly at c .

D. A new view of the 'fabric of the universe'

The equation $n = k Gm/r^2$ allows us to speak about electromagnetic radiation propagating *not through space-time* but through a gravitational field; in deed the equation goes further to suggest that apart from gravitation, electromagnetic radiation cannot propagate (i.e. move from one point to another). We are therefore led to think of the gravitational field (with the electric magnetic field in motion relative to it) not as an entity in space but as space itself! This means that if we were to wipe out these two fields – e.g. by ‘freezing’ all mass and charge into a singularity we would be left not with a singularity in space but a singularity surrounded by a realm – perhaps that much coveted spirit world – of spacelessness (and timelessness, see below) Scientists believe there are four fields in the universe: the gravitational, the electromagnetic, the strong and the weak force. But already some have suggested that the weak field is ultimately electromagnetic [10]. Physicists need the strong force to explain how the particles in the nucleus can hold together against the repulsion due to the positive charge of the protons. If we now propose, for instance, that the positive charge resides on the outside of the nucleic ‘shell’ then we do not need a force any stronger than gravitation to hold the particles together in the nucleus. This view of space as fields together with all that we have said above must compel us to replace the space-time curvature metaphor with space density in explaining relativistic phenomena.

For we realize that time is not a dimension of space but a mere consequence of the fact that these fields are in finite relative motion. We may try to demonstrate this by observing the following:

11 Every object in the universe can and must move only at c relative to light or any other electromagnetic field. If nature had set these two fields to move at infinite velocity relative to each other then every motion would be instantaneous and time – as well as space – as we know it would be nonexistent; and if the fields were fixedly stationary relative to each

other no material object would move and we would be visited with omnitemporality where one second would be an eternity. In other words, what time is depends on the relative motion of these fields. The concept space density is already in use whenever we speak of one medium as being optically denser than another; and naturally therefore the refractive index of a medium, n , is the measure of this density. $n = kGm/r^2$ and the creation of metamaterials. Technology, in the construction of negative index metamaterials, has been guided by an equation which gives n in terms of relative permittivity and relative permeability [11]. The challenge has been to get that material whose permittivity and permeability are simultaneously negative. This has proved very difficult demanding complex and tedious methods of construction. Realizing that $n = kGm/r^2$ (which means that permittivity and permeability are ultimately dependent on atomic gravity) should significantly reduce the problem of design and construction.

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Fixed-Dome Biogester Construction and Determination of Effects of Temperature on the Performance

GJSFR Classification (FOR)
240202,240503,249901

Dr. Ugwoke D .U¹ and Ekpe E.O²

Abstract - Construction of a 0.459m³ fixed-dome galvanized biogester with objectives to determine the effects of temperature on its performance at 100 days retention period was carried out at Abakaliki area of high moisture content cow dung wastes at 1:2 ration. The measurements of the ambient and slurry temperatures and the biogas volumes using mercury- in -glass thermometer (-10.0OC – 100.0OC) and downwards displacement techniques of water in a 25-litre calibrated white jerry can respectively were conducted at 9.30 am, 12.00noon and 2.30pm and corresponding mean values computed for each day. Results showed 25.0OC -41.0OC (ie 25.0OC – 38.5OC ambient and 29.0OC – 41.0OC slurry temperatures) range which agree with earlier results. Two ranges of temperature fluctuations existed in the slurry range. The mild case between 1st and 57th day gave rise to early enhanced quantity and quality biogas production while the later range gave rise to reduced the volume of biogas. The highly fluctuated ambient temperature favoured the activation of the microorganisms for better performance of the digester. The value produced is quite impressive and this is as a result of the favourable micro-organisms activities within the temperature range in the leakage and corrosion free biogester.

Keywords: mesophiles, slurry, digestion, cubic and regression.

I. INTRODUCTION

Energy as capacity for doing work has been an essential input to all aspect of human Endeavour and needs no surface. Energy radiated to the earth is far more that we can ever use. The use of solar energy is expanding generally and particularly in biomass energy form which is one type of renewable energy and a good alternative to the just depleting and polluting energy source.

distortion in its chain – supply at any point in time because it will cause a serious economic and social hardship Okeke, (2004) and Garba, (1996). All energy sources emanated from the sun – a giants star that radiates energy in all directions with only a small portion reaching the earth's The anaerobic production of biofuel known as biogas from a biogester which captures the methane which would have gone into the atmosphere and would have added to global warning met with diverse technologies in the modern time. The process of this methane production passes through four biological and chemical processes of hydrolysis, acidogenesis, acetogenesis and methanogenesis, represented in figure 1.0 below

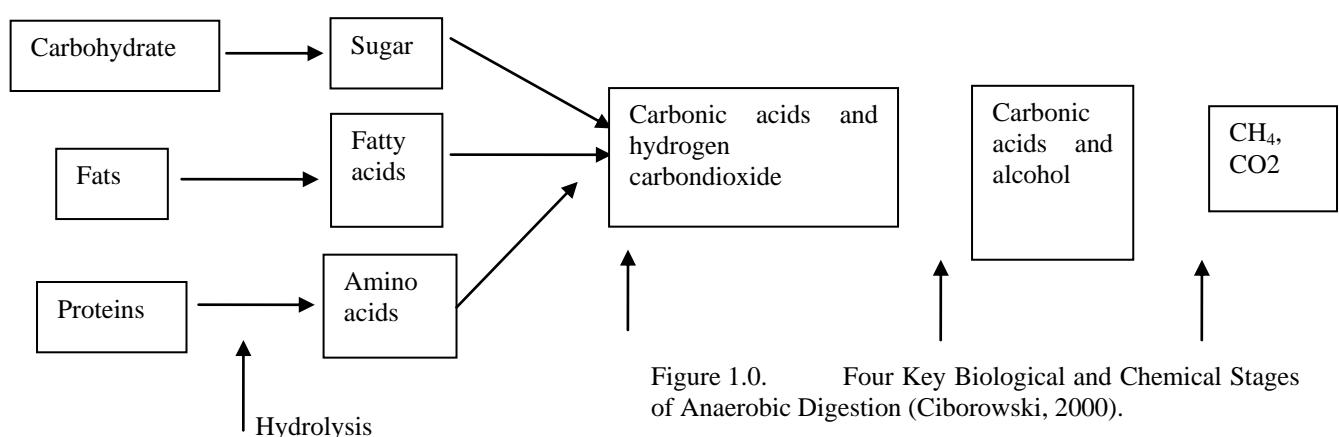


Figure 1.0. Four Key Biological and Chemical Stages of Anaerobic Digestion (Ciborowski, 2000).

Several factors affect these technologies in respect to the production of biogas-(Porkhared, et. al. 1991). These factors include pH- values of the slurry, the nature and type of feed stocks, the C/N ratio, loading rate, the size of the digester and more importantly the temperature of the digester which exists as ambient and slurry temperatures (Dioha el. al.2003).

By the time the radiation reaches the earth, most of the ultraviolet components have been filtered out remaining virtually the visible region with a small portion covering the

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short infrared region. It is this small portion that is converted to heat energy quantified in terms of temperature. Other factors such as the distance of the sun from the earth, length of day, latitude and longitude of the location on the earth, affect the intensities of solar radiations on the earth's surface (Okpani and Nnabuchi, 2008). Also air – mass which is the radiative length of the direct beam path through the atmosphere affects solar radiation (Tomas, 2000).

Therefore temperature as a function of solar radiations affected the content of our digesters. Three zones of this temperature are identified with biogas production through anaerobic degradation of organic materials. These are mesophilic, thermophilic and psychrophilic zones which exist within the wider temperature range corresponding to three different sets of bacteria. They are mesophiles; those that operate best at 20.0 °C – 40.0 °C, the thermophiles; which operate at 40.0 °C – 60.0 °C and the psychrophiles that operate at 4.0 °C – 15.0 °C. Marchaim in 1992 identified that the optimum biogas generation existed at the mesophilic zone of ambient temperature while the methanogenic bacteria are inactive in extreme low zone of psychrophilic zone and high temperature zone of thermophilic zone. Experiences in China, Anonymous reported in 1992, indicated that a rapid change of more than 5°C will slow down biogas production noticeably. This implies that biogas yield is greatly affected by seasons since ambient and fermentative temperature values are influenced by the earth temperature which is related to the atmosphere.

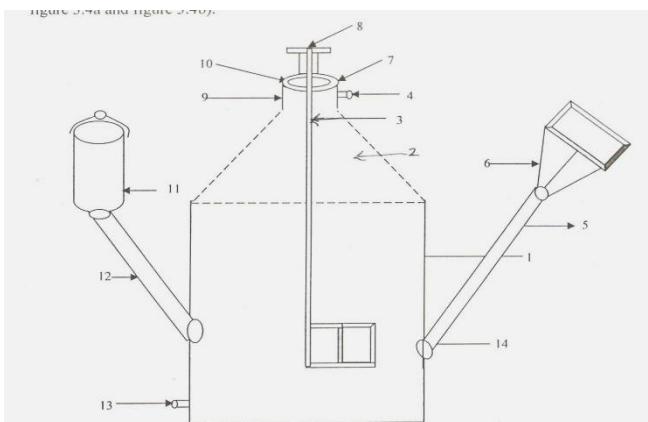
Song et. al. (2004) maintained also that mesophilic digestion takes place optimally at 20.0 °C -45.0 °C and 50.0 °C -52.0 °C for thermophilic digestion. Other investigation reports by Itodo et. al. (1995), Ezeonu et. al. (2005), Nwokoye et. al. (2008), Dioha et. al. (2003) and Dioha et. al. (2006) indicated mesophilic temperature ranges of 20.0°C- 40.0°C, 28.0°C – 35.0°C and 35.00C – 50.00C, 30.00C – 40.00C and 20.00C – 45.00C for cow dung respectively.

It means that all technologies involving anaerobic digestion in order to produce biogas in the past and at present must be within the above ranges. In Nigeria and overseas the existing biogas plants are made of concrete, cement, bricks, plastics and metals. In some cases, though locally, we have digesters fixed with separate or integral gasholders with attended difficulties in costs and operations. Nigeria is rich in manpower and material resources such as quality iron and steels which are potential materials for the construction of improved biodigester (ie galvanized fixed – dome biodigester) with outweighed advantages over the floating types. Also cow dung are readily available in Nigeria.

Therefore, the researcher wishes to re-examine the effect of temperature on the performance of a constructed galvanized fixed – dome biodigester using cow dung from a high moisture content areas of the country – Abakaliki, within a reasonable length of time ie. 8th April, 2009 -17th July, 2009.

II. MATERIALS AND METHOD

The materials used for the construction include: galvanized iron sheets, sockets, bearings, air valve, shaft to stir, bolts and nut; 18mm gauge of galvanized iron plate for cover plate, washers, flat iron bar for stirrer blade, 16mm gauge galvanized iron sheets to form the inlet and outlet pipes, 25-litre calibrated white jerry can and hoses. The researcher adopted a three stage approach of making a detailed master plan of the dimensions of the parts, the construction details and evaluation of the digester using cow dung. Some standard parts such as the bearings, the iron sheet, oil seal, nipple and sockets, bolts and nuts, hoses and black paints were purchased from material markets using the dimensions specified. Some other parts such as top plate for cover, stirring handle, Paddle (blade), tank with inlet and outlet pipes were welded according while shaft, locknuts, etc were machined to fit in using lathe machine in the engineering workshop of National Centre for Energy, Research and Development, NCERD, University of Nigeria Nsukka. The constructed digester consists of the cylindrical compartment, and a hemispherical dome with small cylindrical neck and a metal cover, the inlet and outlet pipes and gas pipes to utility. The capacity is 0.45900 m³ and thickness of 1.5mm to withstand pressure of the gas produced (see figure 2.0 below).



Part no = part name: 1 = Fermentation unit, 2 = gas storage unit, 3 = stirrer shaft, 4 = gas outlet pipe, 5 = slurry inlet pipe, 6 = funnel-like mouth, 7 = bolts hole, 8 = stirrer handle, 9 = short metal eck, 10 = metal reim/ring, 11 = cylindrical outlet mouth, 12 = slurry pipe, 13 = Nipple and socket, 14 = shaft blade.

Figure 2.0: Schematic Diagram of the bilodigester

After positioning the biodigester in a shade and intercepting sun light free environment, it was tested for leakage and was batch fed with fresh cow dung (slurry) collected from Abakaliki Abattoir after sorting to remove non-degradable materials such as stone, cellophane, feather, bones, pebbles etc.

The slurry was obtained by using 1:2 radio of the waste: water. The weighting was carried out by using the “five goat

model" Z051299 weighing balance graduated in imperial and metric scales of 0-110 lb and 0-50.0kg respectively after correcting the zero errors. Then the Slurry was thoroughly mixed and introduced into the digester while it was open. The mass ratio of 90:180) totaling 270kg and equivalent to one-third of the entire volume was used and the container tightly covered with rubber seals, iron plate cover, bolts and nuts, and air valve.

Starting from the next day, the measurements and readings of the ambient and slurry temperatures, and biogas volumes were carried out at 9.30am, 12.00noon and 2.30pm with stirring to break the scum formed on the surface of the slurry and redistribute the temperatures evenly within the contents. The mean values for the ambient temperature, slurry temperature and biogas volumes were computed for 0).

each day after the respective last measurements and readings.

The slurry and ambient temperatures were monitored daily using a mercury – in – glass thermometer graduated in Celsius scale. The range of the scale of the instrument is – 10.0OC – 110.00C. After positioning the digester in convenient place, the ambient and the slurry temperatures were measured after proper stirring of the slurry giving rise to slurry turbling in the cylindrical outlet top showing that these temperatures are the replica of the temperatures inside the biodigester. The volume of the biogas produced was determined by subsequent downward displacements of water in the 25 – litre calibrated jerry can until the biogas produced was exhausted and the total volume taken (see figure 3.0 and table 1.



Table 1.0: Values of Ambient and Slurry Temperature (°C), Biogas Volume (l) and Retention Time (days).

S/N	DATE	RETN. TIME (Days)	Ambient Temp (°C)	Slurry TEMP. (°c)	VOL. (litres)	CUM. GAS VOLUME (litres)
	8/4/09	Charging the Biogester				
1	9/4/09	1	33.0	37.0	13.0	13.0
2	10/4/09	2	34.0	39.0	15.5	28.5
3	11/4/09	3	34.5	36.0	52.5	81.0
4	12/4/09	4	36.0	37.0	49.0	120.0
5	13/4/09	5	34.0	38.0	40.0	170.0
6	14/4/09	6	36.0	37.0	47.0	217.0
7	15/4/09	7	34.0	37.0	41.0	258.0
8	16/4/09	8	35.0	36.5	46.0	304.0
9	17/4/09	9	34.0	36.0	46.5	350.0
10	18/4/09	10	29.0	34.0	36.5	387.0
11	19/4/09	11	35.9	36.0	36.5	423.5
12	20/4/09	12	32.0	35.0	45.5	469.0
13	21/4/09	13	32.5	38.0	34.0	503.0
14	22/4/09	14	35.0	36.0	46.5	549.5
15	23/4/09	15	33.0	37.0	46.0	595.5
16	24/4/09	16	32.0	36.0	46.5	642.0
17	25/4/09	17	35.0	39.0	45.5	687.5
18	26/4/09	18	29.5	36.0	35.5	723.0
19	27/4/09	19	36.0	37.0	31.5	754.5
20	28/4/09	20	30.0	35.0	46.0	800.5
21	29/4/09	21	31.0	37.5	14.0	814.5
22	30/4/09	22	39.0	39.5	18.0	832.5
23	01/5/09	23	26.0	39.0	11.0	843.5
24	02/5/09	24	37.0	35.5	14.5	868.0
25	03/5/09	25	27.0	37.0	32.0	900.0
26	04/5/09	26	37.5	38.0	28.0	928.0
27	05/5/09	27	31.0	41.0	28.5	956.5
28	06/5/09	28	28.5	40.0	24.0	980.5
29	07/5/09	29	30.0	41.0	35.5	1026.0
30	08/5/09	30	32.0	40.0	29.0	1055.0
31	09/5/09	31	33.5	38.0	39.5	1094.5
32	10/5/09	32	34.5	37.0	28.0	1122.5
33	11/5/09	33	32.0	38.0	31.0	1153.5
34	12/5/09	34	38.0	40.5	29.0	1182.5
35	13/5/09	35	32.0	40.0	32.0	1215.0
36	14/5/09	36	33.5	38.0	26.0	1241.0
37	15/5/09	37	33.0	39.0	25.0	1266.0
38	16/5/09	38	35.0	37.0	27.0	1293.0
39	17/5/09	39	27.0	35.5	26.0	1318.0
40	18/5/09	40	31.5	38.0	35.0	1353.0
41	19/5/09	41	35.0	39.0	27.0	1380.0
42	20/5/09	42	32.0	38.0	29.5	1410.0
43	21/5/09	43	30.0	37.0	35.5	1445.5
44	22/5/09	44	38.0	40.5	25.5	1471.0
45	23/5/09	45	32.0	37.0	25.0	1496.0
46	24/5/09	46	38.0	41.0	24.5	1520.5
47	25/5/09	47	38.5	39.0	24.5	1545.0
48	26/5/09	48	34.0	38.0	26.5	1571.0
49	27/5/09	49	33.0	40.0	27.0	1598.5
50	28/5/09	50	36.0	39.0	22.5	1621.0
51	29/5/09	51	36.5	41.0	23.5	1644.5

52	30/5/09	52	35.0	39.0	17.5	1662.0
53	31/5/09	53	32.0	37.0	19.5	1681.5
54	01/6/09	54	32.0	38.0	17.0	1698.5
55	02/6/09	55	32.0	38.0	29.5	1728.0
56	03/6/09	56	33.0	40.0	32.0	1760.0
57	04/6/09	57	30.0	37.0	13.0	1773.0
58	05/6/09	58	37.0	39.5	16.0	1789.0
59	06/6/09	59	37.0	38.5	17.0	1806.0
60	07/6/09	60	27.0	31.5	23.0	1829.0
61	08/6/09	61	34.0	39.0	19.5	1848.5
62	09/6/09	62	35.0	38.0	15.5	1864.0
63	10/6/09	63	33	37	23.5	1887.5
64	11/6/09	64	29	32	17	1904.5
65	12/6/09	65	33	37	14	1918.5
66	13/6/09	66	35	39	12	1930.5
67	14/6/09	67	32	37	12.5	1943.0
68	15/6/09	68	31	35	11	1954.0
69	16/6/09	69	31	34	16.0	1970.6
70	17/6/09	70	30	33.5	14	1984.6
71	18/6/09	71	32	35.5	16	2000.6
72	19/6/09	72	33	38	13	2013.6
73	20/6/09	73	34.5	39.5	12	2025.6
74	21/6/09	74	35	39	10	2035.6
75	22/6/09	75	38	41	11	2046.6
76	23/6/09	76	35.5	37	11.5	2058.1
77	24/6/09	77	33	35	11.5	2269.6
78	25/6/09	78	34	38	12	2081.6
79	26/6/09	79	32	36	17	2098.6
80	27/6/09	80	32	37	16.5	2115.1
81	28/6/09	81	34	38	15.5	2130.6
82	29/6/09	82	32.5	38	13	2143.6
83	30/6/09	83	31	36	11.5	2155.0
84	01/7/09	84	33	36	16	2171.0
85	02/7/09	85	34	38	11	2182.0
86	03/7/09	86	30	35	14	2196.0
87	04/7/09	87	33	36	11	2207.0
88	05/7/09	88	34	40	5.5	2212.5
89	06/7/09	89	30	35	3.5	2216.0
90	07/7/09	90	31	36	4.5	2220.5
91	08/7/09	91	30	36	10.5	2241.0
92	09/7/09	92	31	40	4.0	2245.0
93	10/7/09	93	28	32	4.5	2250.0
94	11/7/09	94	25.6	29	3.0	2253.5
95	12/7/09	95	33	40	5.5	2259.0
96	13/7/09	96	32	38	5.0	2264.0
97	14/7/09	97	33.5	39	4.2	2268.2
98	15/7/09	98	32.5	37.0	4.0	2272.2
99	16/7/09	99	30.5	39.0	3.5	2275.7
100	17/7/09	100	30.0	38.0	3.1	2278.8

III. RESULTS AND DISCUSSION

Observed in figures 4.0 and 5.0 are the consistent fluctuations of the ambient and slurry temperatures with retention time. These give rise to 25.0°C - 38.50°C ambient temperature and 29.0°C - 41.0°C slurry temperature ranges all within the 25.0°C - 41.0°C mesophillic temperature range only.

The novel cubic regression approximation profile of biogas volumes with the same retention time, figure 6.0, gave the minimum and maximum biogas volumes of 13.08 liters on the 99th day -37.67 liters on the 4th day. The stable flammability started on the 4th day.

The consistent fluctuations of temperature were more within the ambient range than in slurry range. This feature might be due to the the lower absorption of heat by more compressed

particles of the slurry unlike in the air. Within the slurry temperature range mild fluctuations exist between the 1st day to the 57th day. After 57th day the fluctuations were high. Within the low fluctuation range the micro-organisms in the slurry were stabilized and active. This leads to high productions of biogas. The maximum value of biogas produced was on the 4th day which is within the period of moderate sustainance of the stability of the organisms. As from 57th day of the retention time of intense absorption of heat by the organisms, the stability was disrupted and as such biogas production decreased. This condition became more pronounced due to insufficient nutrients in the medium.

Figure 7.0 showed the normal cumulative biogas volume of 2198.0 litres within 100 days. This shows that 25.0⁰C – 41.0⁰C mesophilic temperature range has mean favourable effects on the development, growth and stability of the medium micro organisms.

The maintenance of the organisms with the observed mesophilic temperature range tallies consistently with the results of investigations of earlier researchers such as Itodo, et. al. (2005) 20.0⁰- 40.0⁰C, Nwokoye, et. al. (2008) 28.0⁰C – 35.0⁰C, Dioha, et. al. (2003) 30.0⁰C – 40.0⁰C and Dioha, et. al. (2006) 20.0⁰ – 45.0⁰C.

It implies that above 41.0⁰C and below 25.0⁰ the biogas productions will be drastically reduced due to micro organisms inability to survive in the environment.

IV. CONCLUSION

In this work, the novel cubic regression approximation of biogas volumes, cumulative biogas volumes, ambient and slurry temperatures plots with the same retention period indicate that optimum biogas production existed within 25.0⁰ C-41.0⁰ C mesophilic temperature range which is ideal compared with earlier and similar investigation results of Itodo et. al. (2005), Nwokoye et. al (2008) and Dioha et. al (2006).

High ambient temperature variations with the retention time imply direct activation of the particles in the system. Although, we have harsh slurry temperature variations as from 57th day of the period, the fair temperature variations within the temperature range (ie from the 1st day to the 57th day) provided the favourable mean development, growth and stability of the micro-organisms which lead to enhanced biogas production at early stage.

The temperature variations within and outside the digester generally maintain the mean favourable development, growth and stability of the organisms from high moisture areas of the country, which favours biogas production.

Cow dungs are vital raw materials for very early and enhanced biogas productions since the performance of the 0.4590 m³ capacity galvanized fixed-dome biodigester is quite impressive.

Therefore, the researcher recommends the biodigesters, though in much improved forms, and cow dungs, as potential digester and raw material respectively, for more enhanced biogas production both in quantity and quality.

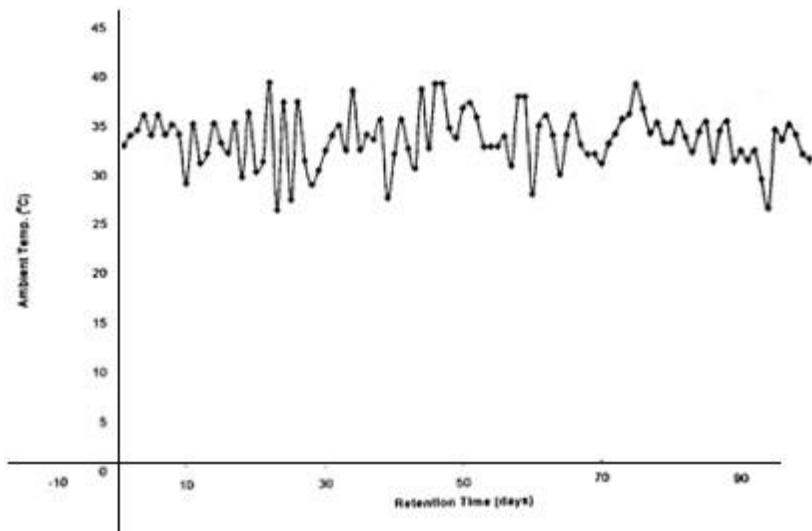


Figure 5.0: Graph of Slurry Temperature (°C) Against Retention time (days)

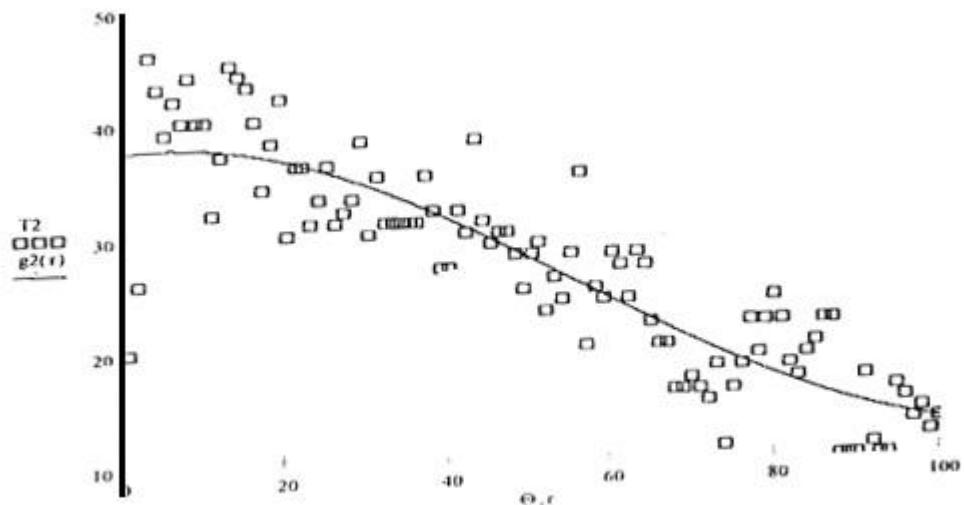


Figure 6.0. Graph of cubic Regression Approximation of Biogas Volume (l) Against Retention time (days)

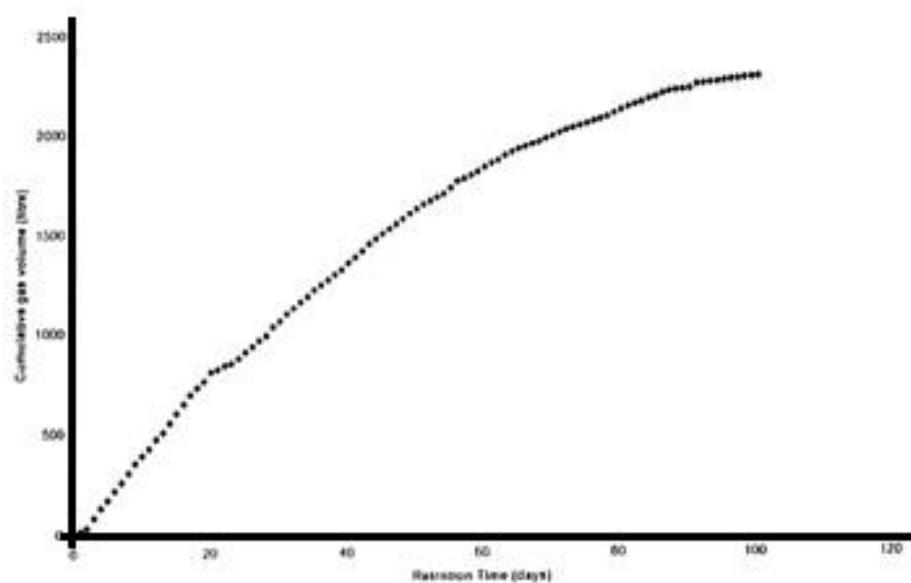


Figure 7.0. Plot of Cumulative Biogas Volume (l) versus Retention time (days).

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- Paper Title should be of Font Size 24 with one Column section.
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- Main Text: Font size 10 with justified two columns section
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- Large Images must be in One Column
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Author Guidelines:

1. General,
2. Ethical Guidelines,
3. Submission of Manuscripts,
4. Manuscript's Category,
5. Structure and Format of Manuscript,
6. After Acceptance.

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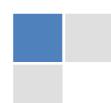
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- One should avoid outdated words.

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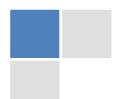
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- Present a justification. Status your particular theory (es) or aim(s), and describe the logic that led you to choose them.
- Very for a short time explain the tentative propose and how it skilled the declared objectives.



Approach:

- Use past tense except for when referring to recognized facts. After all, the manuscript will be submitted after the entire job is done.
- Sort out your thoughts; manufacture one key point with every section. If you make the four points listed above, you will need a least of four paragraphs.
- Present surroundings information only as desirable in order hold up a situation. The reviewer does not desire to read the whole thing you know about a topic.
- Shape the theory/purpose specifically - do not take a broad view.
- As always, give awareness to spelling, simplicity and correctness of sentences and phrases.

Procedures (Methods and Materials):

This part is supposed to be the easiest to carve if you have good skills. A sound written Procedures segment allows a capable scientist to replacement your results. Present precise information about your supplies. The suppliers and clarity of reagents can be helpful bits of information. Present methods in sequential order but linked methodologies can be grouped as a segment. Be concise when relating the protocols. Attempt for the least amount of information that would permit another capable scientist to spare your outcome but be cautious that vital information is integrated. The use of subheadings is suggested and ought to be synchronized with the results section. When a technique is used that has been well described in another object, mention the specific item describing a way but draw the basic principle while stating the situation. The purpose is to text all particular resources and broad procedures, so that another person may use some or all of the methods in one more study or referee the scientific value of your work. It is not to be a step by step report of the whole thing you did, nor is a methods section a set of orders.

Materials:

- Explain materials individually only if the study is so complex that it saves liberty this way.
- Embrace particular materials, and any tools or provisions that are not frequently found in laboratories.
- Do not take in frequently found.
- If use of a definite type of tools.
- Materials may be reported in a part section or else they may be recognized along with your measures.

Methods:

- Report the method (not particulars of each process that engaged the same methodology)
- Describe the method entirely
- To be succinct, present methods under headings dedicated to specific dealings or groups of measures
- Simplify - details how procedures were completed not how they were exclusively performed on a particular day.
- If well known procedures were used, account the procedure by name, possibly with reference, and that's all.

Approach:

- It is embarrassed or not possible to use vigorous voice when documenting methods with no using first person, which would focus the reviewer's interest on the researcher rather than the job. As a result when script up the methods most authors use third person passive voice.
- Use standard style in this and in every other part of the paper - avoid familiar lists, and use full sentences.

What to keep away from

- Resources and methods are not a set of information.
- Skip all descriptive information and surroundings - save it for the argument.



- Leave out information that is immaterial to a third party.

Results:

The principle of a results segment is to present and demonstrate your conclusion. Create this part a entirely objective details of the outcome, and save all understanding for the discussion.

The page length of this segment is set by the sum and types of data to be reported. Carry on to be to the point, by means of statistics and tables, if suitable, to present consequences most efficiently.

You must obviously differentiate material that would usually be incorporated in a study editorial from any unprocessed data or additional appendix matter that would not be available. In fact, such matter should not be submitted at all except requested by the instructor.

Content

- Sum up your conclusion in text and demonstrate them, if suitable, with figures and tables.
- In manuscript, explain each of your consequences, point the reader to remarks that are most appropriate.
- Present a background, such as by describing the question that was addressed by creation an exacting study.
- Explain results of control experiments and comprise remarks that are not accessible in a prescribed figure or table, if appropriate.
- Examine your data, then prepare the analyzed (transformed) data in the form of a figure (graph), table, or in manuscript form.

What to stay away from

- Do not discuss or infer your outcome, report surroundings information, or try to explain anything.
- Not at all take in raw data or intermediate calculations in a research manuscript.
- Do not present the similar data more than once.
- Manuscript should complement any figures or tables, not duplicate the identical information.
- Never confuse figures with tables - there is a difference.

Approach

- As forever, use past tense when you submit to your results, and put the whole thing in a reasonable order.
- Put figures and tables, appropriately numbered, in order at the end of the report
- If you desire, you may place your figures and tables properly within the text of your results part.

Figures and tables

- If you put figures and tables at the end of the details, make certain that they are visibly distinguished from any attach appendix materials, such as raw facts
- Despite of position, each figure must be numbered one after the other and complete with subtitle
- In spite of position, each table must be titled, numbered one after the other and complete with heading
- All figure and table must be adequately complete that it could situate on its own, divide from text

Discussion:

The Discussion is expected the trickiest segment to write and describe. A lot of papers submitted for journal are discarded based on problems with the Discussion. There is no head of state for how long a argument should be. Position your understanding of the outcome visibly to lead the reviewer through your conclusions, and then finish the paper with a summing up of the implication of the study. The



purpose here is to offer an understanding of your results and hold up for all of your conclusions, using facts from your research and generally accepted information, if suitable. The implication of result should be visibly described.

Infer your data in the conversation in suitable depth. This means that when you clarify an observable fact you must explain mechanisms that may account for the observation. If your results vary from your prospect, make clear why that may have happened. If your results agree, then explain the theory that the proof supported. It is never suitable to just state that the data approved with prospect, and let it drop at that.

- Make a decision if each premise is supported, discarded, or if you cannot make a conclusion with assurance. Do not just dismiss a study or part of a study as "uncertain."
- Research papers are not acknowledged if the work is imperfect. Draw what conclusions you can based upon the results that you have, and take care of the study as a finished work
- You may propose future guidelines, such as how the experiment might be personalized to accomplish a new idea.
- Give details all of your remarks as much as possible, focus on mechanisms.
- Make a decision if the tentative design sufficiently addressed the theory, and whether or not it was correctly restricted.
- Try to present substitute explanations if sensible alternatives be present.
- One research will not counter an overall question, so maintain the large picture in mind, where do you go next? The best studies unlock new avenues of study. What questions remain?
- Recommendations for detailed papers will offer supplementary suggestions.

Approach:

- When you refer to information, differentiate data generated by your own studies from available information
- Submit to work done by specific persons (including you) in past tense.
- Submit to generally acknowledged facts and main beliefs in present tense.

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	Containing all background details with clear goal and appropriate details, flow specification, no grammar and spelling mistake, well organized sentence and paragraph, reference cited	Unclear and confusing data, appropriate format, grammar and spelling errors with unorganized matter	Out of place depth and content, hazy format
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	Well organized, Clear and specific, Correct units with precision, correct data, well structuring of paragraph, no grammar and spelling mistake	Complete and embarrassed text, difficult to comprehend	Irregular format with wrong facts and figures
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<i>Result</i>	Well organized, Clear and specific, Correct units with precision, correct data, well structuring of paragraph, no grammar and spelling mistake	Complete and embarrassed text, difficult to comprehend	Irregular format with wrong facts and figures
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Index

A

alternating · 2, 6, 7
associated · 3, 2, 20, X
atmosphere · 33, 34

C

carbohydrates · 21
concentration · 21, 22, 23, 27, VII
Copper enters · 21

D

Determination · 3, 21, 24, 33
Determination Of Effects · 3, 33
Differential · 21, 22, 23, 27
Dirichlet · 2, 5, 6
distribution function · 13, 14, 15, 16, 17, 19, 20
Distributions · 20

E

efficiency · 23
electromagnetic · 30, 31, 32

F

Function Of Atomic · 3, 30

G

general · 13, 20, 22, 23, 30, 31, III, XIII
generalized · 3, 2, 9, 12, 13, 14, 15, 18, 19, 20
Generalized · 3, 12, 13

H

H-function, M-series · 13
hypergeometric · 3, 2, 6, 9, 13, 14, 15, 18

I

integrating · 20
investigation · 3, 7, 31, 34, 38, VI

L

longitude · 34

M

measurements · 21, 33, 35, VII, XII
mesophiles · 33, 34
Metamaterials · 3, 30

N

negative · 21, 22, 32

P

parameters · 14, 17, 20, 22
Performance · 3, 33
persistent · VII
pharmaceutical · 21, 22, 23, 24
photonsimply · 31
polarography · 3, 21, 22
polynomials · 9, 13
Possibility · 3, 30
Probability · 3, 13
Process · 3, III
production · 33, 34, 38, IX

R

refraction · 30, 31
Refraction · 3, 30
representation · 2, 5, 6, 9
representative · 30, 31

S

Search · VII
special · 2, 9, 13, 20

T

Temperature · 3, 33, 36, 39, 40
thatscientists · 32

thisdiscovery · 30
topological · 10, 12

U

understandable · 10

V

volumetric · 22

W

Whittaker · 9



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