

GLOBAL JOURNAL OF SCIENCE FRONTIER RESEARCH BIOLOGICAL SCIENCE Volume 13 Issue 3 Version 1.0 Year 2013 Type : Double Blind Peer Reviewed International Research Journal Publisher: Global Journals Inc. (USA) Online ISSN: 2249-4626 & Print ISSN: 0975-5896

Antifungal Activities of Essential Oils and Crude Extracts of Some Aromatic Plants against *Fusarium* Rot of *Trichosanthes Dioica*

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Abstract - This experiment was conducted for assessing the antifungal activities of essential oils and crude extracts of some aromatic plants against *Fusarium* rot of *Trichosanthes dioica* was at the Central Department of Botany. Pathogenicity test was taken for the confirmation of disease by transferring the inoculum from pure culture. For the control of the above fungus essential oils and extracts of five aromatic plants *Zanthoxylum armatum, Mentha arvensis, Amomum subulatum, Valeriana jatamansi* and *Cymbopogon flexuosus* were used. Each oil and extracts were diluted in different concentration in 80% acetone and in distilled water respectively. The value of minimum inhibitory concentration and percentage of mycelia growth inhibition of oil and extracts were obtained as, oil of *C. flexuosus* showed the higest fungitoxicity (100%) at the 5.0 and 50 μ ImI⁻¹ concentration followed by *Z.armatum, M. arvensis, A. subulatum* and *V. jatamansi* were found to be 10 and 100 μ ImI⁻¹ respectively also the higest percentage of mycelial growth inhibitors were found to be *C. flexuosus* followed by *A. subulatum, Z. armatum, M. arvensis* and *V. jatamansi*.

Keywords : inoculum, minimum inhibitory concentra-tion, mycelial growth, pathogenicity.

GJSFR-C Classification : FOR Code: 270499



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Antifungal Activities of Essential Oils and Crude Extracts of Some Aromatic Plants against *Fusarium* Rot of *Trichosanthes Dioica*

Bina Wagle ^a & Usha Budathoki ^o

Abstract - This experiment was conducted for assessing the antifungal activities of essential oils and crude extracts of some aromatic plants against Fusarium rot of Trichosanthes dioica was at the Central Department of Botany. Pathogenicity test was taken for the confirmation of disease by transferring the inoculum from pure culture. For the control of the above fungus essential oils and extracts of five aromatic plants Zanthoxylum armatum, Mentha arvensis, Amomum subulatum, Valeriana jatamansi and Cymbopogon flexuosus were used. Each oil and extracts were diluted in different concentration in 80% acetone and in distilled water respectively. The value of minimum inhibitory concentration and percentage of mycelia growth inhibition of oil and extracts were obtained as, oil of C. flexuosus showed the higest fungitoxicity (100%) at the 5.0 concentration followed by Z.armatum, M. and 50 μ lml⁻¹ arvensis, A. subulatum and V. jatamansi were found to be 10 and 100 μ lml⁻¹ respectively also the higest percentage of mycelial growth inhibitors were found to be C. flexuosus followed by A. subulatum, Z. armatum, M. arvensis and V. jatamansi . Similarly extracts of C. flexuosus followed by Z. armatum, A. subulatum, V. jatamansi and M. arvensis respectively.

Keywords : inoculum, minimum inhibitory concentration, mycelial growth, pathogenicity.

I. INTRODUCTION

Dointed gourd (*Trichosanthes dioica* Roxb) is a tropical vegetable crop, commonly found in Terai region of Nepal.

a) Post Harvest Damage

The fruits of this plant suffer a lot of damage from fungal disease during summer season. The disease forms a luxuriant wooly mycelium on the affected fruit which appears as if wrapped in absorbent cotton. The tissue in the interior of the fruit becomes watery, soft and the decaying matter emits a bad odour. The species of *Fusarium* are the commonest and most widely distributed in soil and on organic substances, which cause fruit rot on different plants. The rot of pointed gourd is carried out by *Fusarium solani* (Mart.) Sacc, Synder & Hansen.

The aim of the present study was to isolate the pathogen from infected pointed gourd and test its

Author α σ : Central Department of Botany, Tribhuvan University, Kirtipur, Kathmandu. pathogenicity for studing the antifungal activities of oils and extracts of different aromatic plants against the *F. solani*.

II. METHODOLOGY

The research was carried out with the following steps:

a) Collection of Host and Isolation of Pathogen

The infected pointed gourd was collected from local market of Kathmandu valley. A piece of fungal colony was transferred aseptically on a petri plate containing PDA media and was incubated in inverted position in an incubator at 25±2°C for one week. After one week, the mycelial growth was observed under a compound microscope.

b) Identification and Pathogenicity Test

After studying and observing the characteristics features of the pathogen, it was identified as *F. solani*. The characteristics features of the fungus were identified with the help of standard literature (Booth, 1977). For pathogenicity test, the inoculum from the pure culture of *F. solani* was transferred to the healthy fruit. When incubated at $25\pm2^{\circ}$ C for 7 days, the characteristics symptoms were produced, which were found to be similar with the symptoms on fruits previously collected.

c) Maintenance of the Pure Culture

The pure culture of *F. solani* was preserved by sub-culturing in several slants and plates containing PDA media.

d) Preparation of one week old culture

The fungus from pure culture was inoculated into PDA media containing petriplates, and incubated. After seven days the inoculum disc was taken from the culture for further experiment.

e) Measurement of Spore

Measurement of spore was done by the help of ocular micrometer and stage micrometer. Ocular micrometer was calibrated by the help of stage micrometer.

Calculation of calibration factor applying the formula by:

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No. of division on stage micrometer

One ocular division = $\frac{1}{No. of division of ocular micrometer} \times 10 \mu m$

Collection and extraction of essential oils and extracts of aromatic plants

Plant samples of *V. jatamansi*, *A. subulatum*, *Z. armatum*, *M. arvensis* and *C. flexuosus* each 50 gm.were collected out of these *V. jatamansi* and *A. subulatum* were hydro- distilled for 6-8 hours in Clevenger's apparatus in 500 ml water for extraction of oiland rest were taken from HPPCL (Herb Production and Processing Company Limited). Similarly the crude extract of aromatic plants were made by the help of mortar with pistal.

g) Determination of yield of essential oils

The volume of oil extracted was noted down. The yield of the essential oil was calculated in terms of % using the following formula.

h) Dilution of essential oil and crude extract

Essential oils were diluted into different concentration of 0.625, 1.25, 2.5, 5.0, 10.0, μ lml⁻¹ with 80 % acetone and each crude extracts was diluted to different concentrations, 0.625, 1.25, 2.5, 5.0, 10.0, 20, 30, 40, 50 & 100 μ lml⁻¹ with distilled water.

i) Assessment of toxicity of oil and extract against fungal pathogen

The toxicity of the oils was assessed by using the poisoned food technique 0.5ml of each concentration of each oil and extract was taken in presterilized cooled petriplate and 9.5 ml of PDA media was poured on that with gently swirling to mix the contents thoroughly. In control set the essential oil was replaced by equal volume of acetone (80%) and the control set of crude extract was replaced by equal volume of distilled water. The inoculum disc (4mm diameter) taken from the 7 days old culture of the pathogen, was placed aseptically at the centre of each plate and turned upside down in its position. The plates were then incubated at $25\pm2^{\circ}$ C for 7 days. All the experiments were revised thrice. The percentage inhibition of mycelial growth of test fungus was calculated separately.

j) Calculation of percentage of mycelial growth inhibition

Fungitoxicity was assessed in terms of percentage inhibition of mycelial growth of test fungus.

% inhibition of mycelial growth =
$$\frac{g_c - g_t}{\frac{g_c}{g_c}}$$

Where,

 $g_{c}=\mbox{growth}$ of mycelial colony after incubation in control set i.e. diameter of colony in control set - diameter of inoculum disc

 g_t = growth of mycelial colony after incubation period in treatment set i.e. diameter of the colony in treatment set - diameter of inoculum disc.

k) Determination of minimum inhibitory concentration of essential oil and extract

The minimum inhibitory concentration (MIC) is the concentration of any substance in the external medium which just inhibits cell division of a normal cell.

MIC has been expressed as the minimum dose of the essential oil and extract required for complete (100%) inhibition of mycelial growth of the test fungus.

III. Result and Disscussion

a) Assessment of fungitoxicity of essential oils and extracts

Essential oils and extracts of different plant species showed different efficacies in the inhibition of the mycelial growth by measuring the colony size of culture plates.

S.N	Concent-ration	Inoculum size (mm)	Colony size		Mean	Mycelial	% of inhibition	
	of oil		(mm)			colony size	growth	of mycelial
	μ lml $^{-1}$		I	П		(mm)	(mm)	growth
1	0	4	42	42	42	42.00	38.00	0.00
2	0.62	4	19	18	19	18.67	14.67	61.40
3	1.25	4	12	11	12	11.67	7.67	79.82
4	2.50	4	8	7	8	7.67	3.67	90.35
5	5	4	4	4	4	4.00	0.00	100.00
6	10	4	4	4	4	4.00	0.00	100.00

Table 1: Percentage of inhibition of *F. solani* mycelial growth using essential oils of *C. flexuosus*

Minimum inhibitory concentration (MIC) =5 μ ImI⁻¹

Essential oil of *C. flexuosus* showed mycelial inhibition as 0%, 61.40%, 79.82%, 90.35%, 100% & 100% at 0, 0.625, 1.25, 2.5, 5.0, 10.0 μ lml⁻¹ oil concentration against *F. solani* respectively.

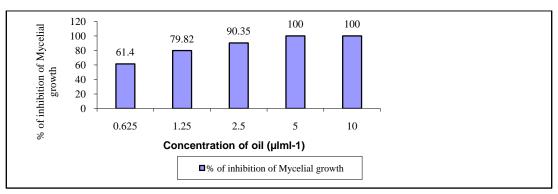


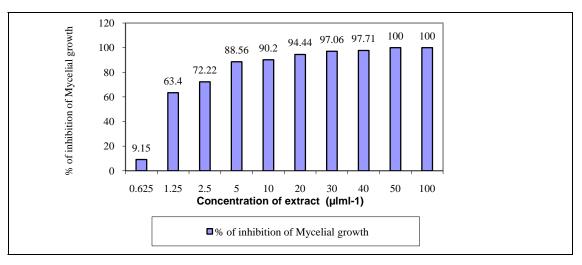
Figure 1 : Percentage of antifungal activity of C. flexuosus oil against F. solani

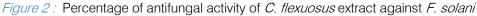
Table 2 : Percentage of inhibition of F. solani mycelial growth using extract of C.flexuosus

	Concent-	Incoulum	Colony size (mm)			Mean	Mycelial	% of
S.N	ration of extract µlml⁻¹	act size (mm) colony			growth (mm)	inhibition of mycelial growth		
1	0	4	55	55	55	55.00	51.00	0.00
2	0.62	4	50	50	51	50.33	46.33	9.15
3	1.25	4	23	23	22	22.67	18.67	63.40
4	2.50	4	18	18	18.5	18.17	14.17	72.22
5	5	4	10	10	9.5	9.83	5.83	88.56
6	10	4	9	9	9	9.00	5.00	90.20
7	20	4	7	7	6.5	6.83	2.83	94.44
8	30	4	6	5	5.5	5.50	1.50	97.06
9	40	4	5	5.5	5	5.17	1.17	97.71
10	50	4	4	4	4	4.00	0.00	100.00
11	100	4	4	4	4	4.00	0.00	100.00

Minimum inhibitory concentration (MIC) =50 μ ImI⁻¹

Extract of *C. flexuosus* showed mycelial inhibition as 0%, 9.15%, 63.40%, 72.22%, 88.56%, 90.20%, 94.44%, 97.06%, 97.71%, 100% & 100% at 0, 0.625, 1.25, 2.5, 5.0, 10.0, 20, 30, 40, 50, $100 \,\mu$ lml⁻¹ oil concentration against *F. solani* respectively.



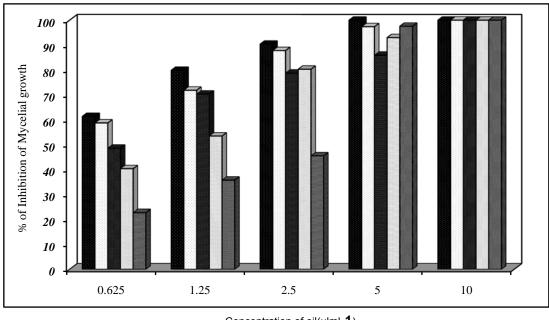


Minimum Inhibitory Concentration (MIC) value of Aromatic plants oils and extracts of *C. flexuosus* was 5.0 & 50.0 μ lml⁻¹ similarly 10.0 & 100.0 μ lml⁻¹ for *M. arvensis, V. jatamansi, A. subulatum* and *Z. armatum* respectively.

b) Comparative fungi toxicities of different essential oils

S.N rati	Concent-	% of Mycelial growth inhibition							
	ration of oil μ lml ⁻¹	Cymbopogon flexuosus	Zanthoxylum armatum	Valeriana jatamansi	Mentha arvensis	Amomum subulatum			
1	0	0	0	0	0	0			
2	0.625	61.4	58.77	48.61	40.35	22.76			
3	1.25	79.82	71.93	70.37	53.51	35.77			
4	2.5	90.35	87.77	78.7	80.26	45.53			
5	5	100	97.37	85.9	92.98	97.56			
6	10	100	100.00	100	100	100			

Table 3: Comparative fungitoxicities of different essential oils in different concentrations



Concentration of oil(μ ImI-1)

Cymbopogon flexuosus	□Zanthoxylum armatum	Valeriana jatamansi
■ <i>Mentha arvensis</i>	Amomum subulatum	

Figure 3 : Fungitoxicities of different essential oils in different concentrations

S.N	Concent-	% of Mycelial growth inhibition							
	ration of extract μlml ⁻¹	Amomum subulatum	Zanthoxylu-m armatum	Valeriana jatamansi	Mentha arvensis	Cymbopogon flexuosus			
1	0	0	0	0	0	0			
2	0.625	30.72	26.47	18.95	14.38	9.15			
3	1.25	40.22	44.44	48.69	20.92	63.4			

4	0.5	04.07	77 45	E2 E0	06.00	70.00
4	2.5	84.97	77.45	53.59	26.80.	72.22
5	5	87.91	80.64	58.17	59.48	88.56
6	10	89.87	86.27	62.75	68.28	90.2
7	20	91.83	90.20	77.78	73.53	94.44
8	30	94.77	90.52	84.97	77.78	97.06
9	40	96.73	96.08	87.58	83.33	97.71
10	50	98.37	99.02	98.37	90.52	100
11	100	100	100.00	100	100	100

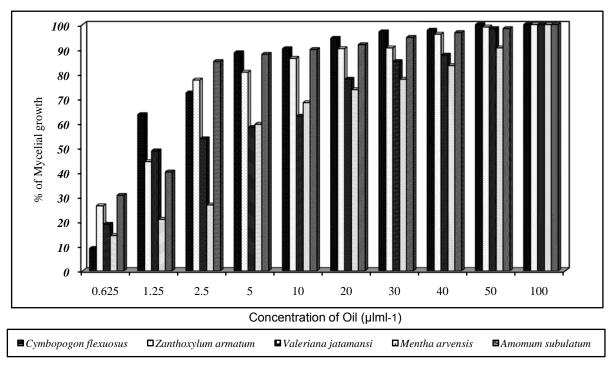


Figure 4 : Fungitoxicities of different extracts in different concentrations

Out of five test plants, essential oil and extract of *C. flexuosus* was found more effective in its fungitoxic properties and showed 100% inhibition of mycelial growth of the test fungus at MIC of 5.0μ ImI⁻¹ and 50μ ImI⁻¹ respectively. Similarly Z. armatum, V. jatamansi, M. arvensis, A.subulatum were found effective and showed 100% mycelial growth inhibition at 10μ lml⁻¹ and 100μ lml⁻¹ respectively. Also the percentage of mycelial growth inhibition were found to be different, according to their different concentration of each essential oil and extracts, the highest percentage of mycelial growth inhibition were found to be oil of C. flexuosus, Z. armatum, M. arvensis, A. subulatum and V. jatamansi respectively and similarly extracts of C. flexuosus, Z. armatum, A. subulatum, V. jatamansi М. arvensis respectively. and Thus comparative fungitoxicities of five different essential oils and extracts were observed against F. solani.

A perusal of literature showed that, similar experiments for the control of *F. solani* using essential

oils and extracts has not yet been done in Nepal. So this is the first study to assess fungitoxicities of essential oils and extract against *F. solani*. This research work can be applicable for commercial scale of inhibitory activities.

IV. Acknowledgements

First of all, I would like to express my sincere gratitude and appreciation to my supervisor Professor Dr. Usha Budathoki, of Central Department of Botany, T.U. for her kind instructions, valuable suggestions, comments, guidance, encouragement and sound cooperation throughout this research work.

I am grateful to Professor Dr. Pramod Kumar Jha, Head of the Central Department of Botany, T.U. for his administrative and laboratory support. I would like to extend my thanks for all the friends who helped me to carry out this research in concrete form. Moreover, I am grateful to Er. Sunil Dhakal for his kind cooperation. Lastly, I would like to express deepest appreciation to my Family and all of my relatives for their constant suggestions, co- operations and best wishes.

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