

GLOBAL JOURNAL OF SCIENCE FRONTIER RESEARCH: D AGRICULTURE AND VETERINARY

Volume 16 Issue 3 Version 1.0 Year 2016

Type: Double Blind Peer Reviewed International Research Journal

Publisher: Global Journals Inc. (USA)

Online ISSN: 2249-4626 & Print ISSN: 0975-5896

GC-MS Analysis for Structural Identification and Bioactive Compounds in Methanolic Leaf Extract of *Mallotus Oppositifolius*

By Igwe K. K., Madubuike A. J., Amaku F. J., Chika Ikenga & Otuokere I. E.

Michael Okpara University of Agriculture, Nigeria

Abstract- The aim of the present study is to investigate the bioactive compounds from the leaf extract of *Mallotus oppositifolius* using GCMS analysis. The chromatogram showed nine peaks indicating the presence of nine compounds in the extract. The major phytocompounds in the leaf were Glutaconic anhydride with the highest concentration in the extract, 40.19 peak area %, RT 22.686 and molecular formula $C_5H_4O_3$; 2-Mercaptophenol with 18.23 peak area %, RT 22.068 and molecular formula C_6H_6 OS. Iso-Valreic and Valeric acids with 12.39,2.53 peak area %, RT 3.676,7.037 and the same molecular formula C_5H_{10} O_2 which had been proposed to have anticonvulsant effect in valerian and act as neurotransmitter. Oleamide with the least concentration of 2.15 peak area %, RT 27.959 and molecular formula $C_{18}H_{35}NO$ which could induce sleep in animals, being studied as a potential medical remedy for mood and sleep disorder and cannabinoid–regulated disorder. The phytochemicals in *Mallotus oppositifolius* could be of therapeutic importance.

Keywords: GCMS analysis, mallotus oppositifolius, anticonvulsant, neurotransmitter, sleep depressant.

GJSFR-D Classification: FOR Code: 079999



Strictly as per the compliance and regulations of:



© 2016. Igwe K. K., Madubuike A. J., Amaku F. J., Chika Ikenga & Otuokere I. E. This is a research/review paper, distributed under the terms of the Creative Commons Attribution-Noncommercial 3.0 Unported License http://creativecommons.org/licenses/by-nc/3.0/), permitting all non commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

GC-MS Analysis for Structural Identification and Bioactive Compounds in Methanolic Leaf Extract of Mallotus Oppositifolius

Igwe K. K. α, Madubuike A. J. σ, Amaku F. J. ρ, Chika Ikenga ω & Otuokere I. E. ¥

Abstract- The aim of the present study is to investigate the bioactive compounds from the leaf extract of Mallotus oppositifolius using GCMS analysis. The chromatogram showed nine peaks indicating the presence of nine compounds in the extract. The major phytocompounds in the leaf were Glutaconic anhydride with the highest concentration in the extract, 40.19 peak area %, RT 22.686 and molecular formula C₅H₄O₃; 2-Mercaptophenol with 18.23 peak area %, RT 22.068 and molecular formula C₆H₆ OS. Iso-Valreic and Valeric acids with 12.39,2.53 peak area %, RT 3.676,7.037 and the same molecular formula C5H10 O2 which had been proposed to have anticonvulsant effect in valerian and act as neurotransmitter. Oleamide with the least concentration of 2.15 peak area %, RT 27.959 and molecular formula C₁₈H₃₅NO which could induce sleep in animals, being studied as a potential medical remedy for mood and sleep disorder and cannabinoid-regulated disorder. The phytochemicals in Mallotus oppositifolius could be of therapeutic importance.

Keywords: GCMS analysis, mallotus oppositifolius, anticonvulsant, neurotransmitter, sleep depressant.

I. Introduction

he use of plants in the treatment of ailments has been long time immemorial [1]. oppositifolius(Geisel) is one the plant used by Nigerians for the treatment of skin diseases [2]. Mallotus oppositifoliusis an erect branching perennial shrub up to 3.6 m high when fully matured. The plant is commonly found in drier types of forest and grow throughout the West Africa region [3]. Ethnobotanically, *Mallotus* oppositifolius is used as chewing sticks for cleaning the teeth and the stem for yam stakes. The Ohafia people in Nigeria use the cold infusion to expel placenta blood clot after delivery, while the decoction is a vermifuge in Ivory Coast. In Ghana, the crushed leaves are applied to inflammation of the eye during an attack of small pox [3]. Rottlerin has also been found in its bark and leaves [4]. The aqueous and ethanol extracts of the plant show antifungal properties [5] and anti parasitic activity

Author α σ : Department of Veterinary Physiology, Pharmacology and Biochemistry, Michael Okpara University of Agriculture, Umudike, Nigeria. e-mail: kkigwe191@gmail.com

Author p: Physical Chemistry Research Lab, University of KwaZulu, Natal, Durban, South Africa.

Author W: Recare Natural Products, Lagos, Nigeria.

Author ¥: Department of Chemistry, Michael Okpara University of Agriculture, Umudike, Nigeria.

against blastocystishominis [6] The bioassay-guided fractionation of an ethanol extract of the leaves and inflorescence of *Mallotu* oppositifolius collected in Madagascar led to the isolation of the two new bioactive dimeric phloroglucinols mallotojaponins B and C, together with mallotophenone. These compounds show antiproliferative and antiplasmodial activities [7]. The crude extracts of Mallotus oppositiformis possess antifungal activity on most of the fungi and inhibits the growth of Aspergillus flavus, Candida albicans, Microsporium audouinii, Penicillium spp, Trichophyton mentagrophytes, Trichoderma spp and Trichosporon cutaneum [5]. The leaves are ingredients of common anti-malaria and anti-inflammatory remedies Phytochemical screening of Mallotus oppositiformis revealed the presence of secondary metabolites such as alkaloids, phenols, flavonoids, anthroquinones and cardenolides. A higher concentration of these resides in the leaves than in the root [9]. Hydroalcoholic extract of leaves of Mallotus oppositifolius plant is used for CNS conditions in Ghana, which exhibits antidepressant effects mediated by enhancement of serotoninergic neurotransmission and inhibition of glycine receptor activation [10]. There is increase in fungal related cases for the last decade. Fungal related diseases may not be common as other microbial infections but, when present, they are difficult to treat especially if patients immunity is low [11]. Therefore traditional doctor who tries to cure an ailment using plant may use the whole plant or extract from leave, stem, root, and seed or mix all together. This type of treatment is wrong so there is need to scientifically analyze the medicinal plant. GCMS analysis has been employed in this research to identify phytochemicals responsible for bioactivities associated with Mallotus oppositifolius.

II. MATERIAL AND METHODS

a) Plant Materials

Fresh leaves of *Mallotus oppositifolius* was harvested at Ohafia town in Abia State, Nigeria. The plant leaves were identified by Prof M C Dike at the Taxonomy section of College of Natural and Environmental Management, Michael Okpara University of Agriculture, Umudike, Nigeria.

b) Preparation of Plant Extract

The plant material of *Mallotus oppositifolius* was collected from wild, shade dried for 10 days and pulverized to powder using mechanical grinder. The plant extract was prepared using Soxhlet method described by [12]. Thirty five grams (35 g) of powdered sample was introduced into the extraction chamber of the Soxhlet extractor using methanol as solvent. Temperature was maintained at 70o C throughout the extraction period of 48 hrs. At the end of the extraction period, the extract was concentrated using oven at 35o C to obtain dried extract which was sent for GCMS analysis.

c) GCMS analysis of Mallotus oppositifolius

The characterization of the Phytochemicals in *Mallotus oppositifolius* was done using GC-MS QP2010 Plus (Shimadzu, Japan). The identification of the phytochemicals in the sample was carried out using a QP2010 gas chromatography with Thermal Desorption System, TD 20 coupled with Mass Spectroscopy (Shimadzu). The ionization voltage was 70eV. Gas Chromatography was conducted in the temperature programming mode with a Restek column (0.25 mm, 60 m, XTI-5). The initial column temperature was 80oC for 1min, and then increased linearly at 70oC min-1 to 220oC, held for 3 min followed by linear increased temperature 10oC min-1 to 290oC for 10 min. The temperature of the injection port was 290oC and the GC-MS interface was maintained at 290oC. The sample

was introduced via an all-glass injector working in the split mode, with helium carrier gas low rate of 1.2 ml min-1. The identification of compounds was accomplished by comparison of retention time and fragmentation pattern, as well as with mass spectra of the GC-MS.

d) Identification of Phytocompoments in Mallotus oppositifolius

GC-MS Chromatogram Mallotus of oppositifolius revealed nine peaks showing that nine compounds different were present. Identity of the active components in the extract was done by comparison of their retention indices, peak area percentage and mass spectra fragmentation pattern with those stored in the database of National Institute of Standards and Technology (NIST) and also with published literature, NIST08.LIB [13], WILEY8.LIB [14], PESTEI-3.LIB and FA-ME.LIB library sources were used for matching the identified components from the plant material. The name, molecular weight, formula, structure and bioactivities of the compounds were ascertained.

III. RESULTS AND DISCUSSION

a) Results

GCMS chromatogram of the methanolic extract of *Mallotus oppositifolius* (Figure 1) showed nine peaks which indicated the presence of nine phytochemicals constituents.

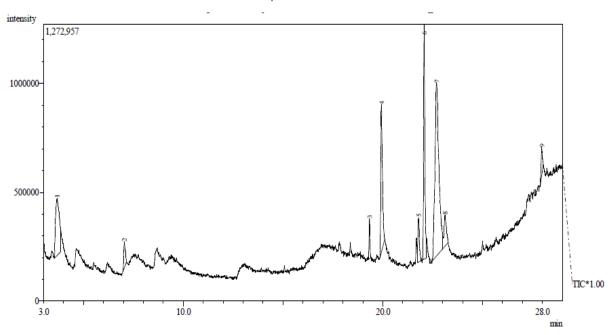


Figure 1: Shows the chromatogram of Mallotus oppositifolius

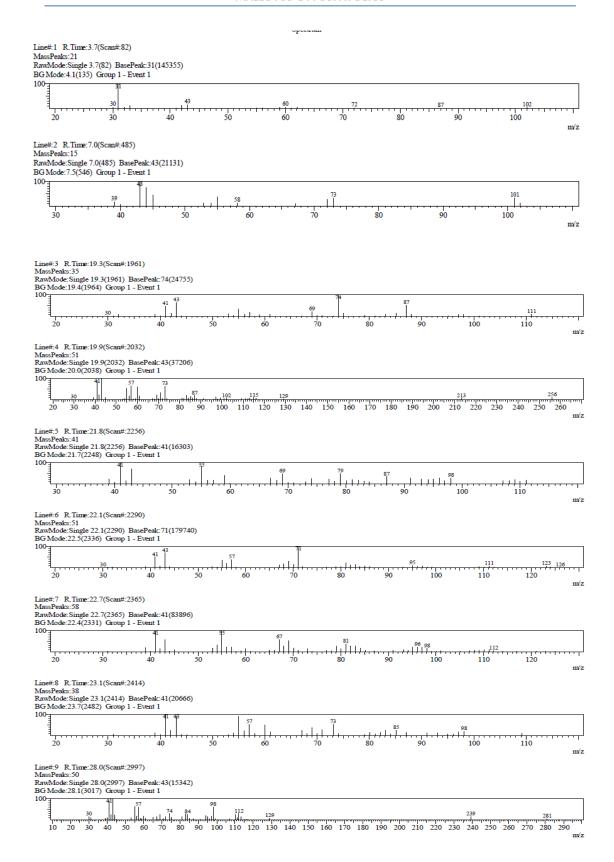


Figure 2 : Shows the mass spectra of the nine phytocompounds in Mallotus oppositifolius identified by GCMS analysis

Table 1: Shows the names, retention time, peak area percentage, molecular weight, molecular formula and bioactivity of compounds identified in *Mallotus oppositifolius* by GCMS analysis

		, 0. 00	.,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,		104 111	manorae			GCMS analysis
Bioactivity	It has been proposed that it is the anticonvulsant agent in valerian.	Neurotransmitter	antibacterial drug fungicide	Mild antioxidant and anti-atherosclerotic activity [15]			peroxisome proliferator activated receptor	estrogen receptor; agonist	It accumulates in the cerebrospinal fluid during sleep deprivation and induces sleep in animals. It is being studied as a potential medical treatment for mood and sleep disorders, and cannabinoid-regulated depression [16].
Molecular structure	HO CH ₃	HO CH ₃	ено ОНО	CH ₃		HS-OH		HO	H ₂ N H ₃ C
Molecular formular	$C_5H_{10}O_2$	C ₅ H ₁₀ O ₂	C ₆ H ₈ O ₂	$C_{16}H_{32}O_2$	C2H14	SO ⁹ H ⁹ O	C ₅ H ₄ O ₃	$C_5H_6O_2$	C ₁₈ H ₃₅ NO
Molecular weight	102.13	102.13	112.12	256.42	98.18	126.17	112.08	98.09	281.47
Peak area %	12.39	2.53	2.08	14.22	3.87	18.23	40.19	4.33	2.15
Retention time	3.676	7.037	19.329	19.926	21.791	22.068	22.686	23.115	27.959
Name of Compound	3-Methylbutanoic acid or more commonly isovaleric acid	Valeric acid or pentanoic acid	Sorbic acid	n-Hexadecanoic acid or Palmitic acid	Surfactant	2-Mercaptophenol	Glutaconic anhydride	2-Hydroxy-2-cyclopenten-1-one	Oleamide
S.No	-	۷	ε	4	⁸ НОСН	O	7	8	O

b) Discussion

The chromatogram of Mallotus oppositifolius indicated the presence of nine phytocompounds. The compound 3-Methylbutanoic acid demonstrated anticonvulsion activity. Convulsion is produced by a number of metabolic disorder such as hypoglycemia, hypocalcaemia and hormonal imbalances [17]. This compound identified by GCMS analysis could counter the effect of convulsion and therefore could be used as a therapeutic remedy for idiopathic seizures. A seizure represents the abnormal behaviour caused by an electrical discharge from neurons in the cerebral cortex clinical signs and symptoms that vary presents according to the site of neuronal discharge in the brain[18]. Manifestations of seizure generally include sensory, motor, autonomic or psychic phenomena. A convulsion refers to the specific seizure type of a motor seizure involving the entire body was neurotransmitter effect demonstrated by phytocompound, Valeric acid or Pentanoic acid as was identified by GCMS analysis. Neurotransmitters carry nerve impulses across synapse and are small molecules that incorporate a positively charged nitrogen atom. They include several amino acids, peptides and monoamines. The amines acid, glutamine, glycine and gamma amino butyric acid (GABA) serve neurotransmitters at most CNS synapse [19].

The compound Glutaconic anhydride with a retention time of 22.686 and a peak area percentage of 40.19% had a peroxisome proliferator activated receptor activity. Peroxisomes play important role in B-oxidation leading to the formation of acetyl Co A and hydrogen peroxide which is broken down by catalase. [20]. The peroxisome system facilates the oxidation of very long fatty acids example C_{20} and C_{22} . Peroxisomes shorten the side chain of cholesterol in the bile acid formation and also takes part in the synthesis of etherglycerolipids [21]. Therefore this compound Glutaconic anhydride could play a biochemical role of facilitating B-oxidation in the cell. Oleamide was identified also with GCMS at retention time of 27.959 with 2.15% peak area percentage. The compound exhibits a bioactivity of influencing mood and sleep disorder especially if it accumulates in the cerebrospinal fluid during sleep deprivation. The compound also induces sleep in animals [16].

IV. CONCLUSION

The result of this analysis showed the presence of various phytocompounds in methanolic extract of *Mallotus oppositifolius*. Glutaconic anhydride which had the highest concentration in the extract (40.19%; $C_5H_4O_3$) and n-Hexadecanoic acid (14.22%; $C_{16}H_{32}O_2$) showed peroxisome proliferetor receptor activation and antioxidantion, anti-atherosclerotic activity respectively. The compound 3-Methylbutanoic acid ($C_5H_{10}O_2$)

commonly known as isovaleric acid and its isomer Valeric acid ($C_5H_{10}O_2$) which is also known as pantanoic acid were found to be anticovulsants and nuerotrasmitters respectively. Sorbic acid $C_6H_8O_2$ and Oleamide $C_{18}H_{35}NO$ with close range concentration of (2.08%; 2.15%) in the *Mallotus oppositifolius* extract exhibited activity of antibacterial,fungicidal, and sleep inducer respectively. The phytocompound,oleamide could therefore be pharmacologically useful as preanaesthetic agent. The plant has a wide array of medicinal usage and those compound identified by Gas Chromatography-Mass Spectrometry could undergo molecular docking to creat new roadmap for drug modelling.

References Références Referencias

- Sofowora A.E.,(1993) Medicinal Plants and Traditional Medicine in Africa. Vol 2. Spectrum Books Ltd, Ibadan; 288.
- Adekunle A.A., (2001). Ethnobotanical Studies of some Medicinal Plants from Lagos State of Nigeria. Nigerian Journal of Botany. 14: 71–9.
- 3. Burkill H.M., (1997).The Useful Plants of the West Tropical Africa. Vol 2. *Royal Botanic Garden*, *Kew.*pp 969
- Oliver B.E.P., (1960). Medicinal Plants in Nigeria. Nigerian College of Arts, Science and Technology, Lagos. pp 70.
- Adekunle, A.A., and Ikumapayi, A.M., (2006). Antifungal property and phytochemical screening of the crude extracts of Funtumiaelastica and *Mallotus* oppositifolius. The West Indian Medical Journal 55 (4): 219–23.
- Bremer Christensen, C., Soelberg, J., Stensvold, CR., and Jäger, A.K., (2015). Activity of Medicinal Plants from Ghana Against the Parasitic Gut Protist Blastocystis. *J Ethnopharmacol*. doi:10.1016/j.jep 2015.03.006.PMID25773490.
- Harinantenaina Liva, Bowman Jessica D., Brodie Peggy J., Slebodnick Carla, Callmander Martin W., Rakotobe Etienne, Randrianaivo Richard, Rasamison Vincent E., Gorka Alexander, Roepe Paul D., Cassera Maria B., and Kingston David G. I. Antiproliferative and Antiplasmodial DimericPhloroglucinols from *Mallotus* oppositifolius from the Madagascar Dry Forest (1). Journal of Natural **Products** 76 (3): 388-93. doi:10.1021/np300750q.PMC: 3606680. PMID2328 6240
- 8. Burkhill, H.M (1994). The Useful Plants of West Tropical Africa. *Royal Botanic Gardens,Kew.*2.144-150.
- Farombi, E.O., Ogundipe, O.O., and Moody, J.O.(2001). Antioxidant and Anti-inflammatory Activities of *Mallotus oppositifolius* in Model Systems. *Afr.J.Med.Sci.* 30:213-215.

- 10. Kennedy K. E. Kukuia, Priscilla K. Mante, Eric Woode, Elvis O. Ameyaw, and Donatus W. Adongo, Antidepressant Effects of Mallotus oppositifolius in Acute Murine Models. Article ID 324063, 12 pages http://dx.doi.org/10.1155/2014 /324063
- 11. Bryce K.(1992).The Fifth kingdom. Mycologue Publications, Ontario, 451.
- 12. Jensen W.B., (2007). The origin of Soxhlex Extraction. Journal Clinical Education. 84 (12), 1913-1914, 2007.
- 13. Stein S. E., (1990). National Institute of Standards and Technology (NIST) Mass Spectral Database and Software. Version 3.02, USA.
- 14. Mc Lafferty F. W., (1986). Registry of mass spectral data. Fourth electronic ed. Wiley New York.
- 15. Cho K.H., (2015). Monoacylglycerol oleic acid has Stronger Antioxidant, Anti-atherosclerotic and Protein glycation Inhibitory Activities than MAGpalmitic acid. Journal of Medicinal Food, 13(1): 99 -107.
- 16. Salvador Huitron Resendiz, Lhys Gombart, Benjamin F. Cravatt, and Steven J. Henriksen (2001). Effect of Oleamide on Sleep and its Rlationship to Blood pressure, Body temperature and Locomotor activity in Rats. Experimental Neurology 172 (1): 235 - 243.
- 17. V. P. Studdert, C. C. Gay., and D. C. Blood. (2012). Saunders Comprehensive Veterinary Dictionary. Fourth Edition Pg. 263.
- 18. Mosenich R. K., and So E. L., (1996). The Clinical Approach to Classification of Seizures and Epileptic Syndromes. Mayo Clinic Proceeding 71, 405-441.
- 19. Squire L. R., Bloom F. F., and Mc Connell S. K., (2003). Fundamental Neuroscience (2nd ed., pp 188-189, 391 – 416). New York: Academic Press.
- 20. Reddy J. K., and Mannaerts G. (1994).Peroxisomal Lipid Metabolism. Annu Rev Nur, 14: 343.
- 21. Robert K. Murray., Daryly K. Granner., and Victor W. Rodwell (1998). Harpers Illustrated Biochemistry. 27th Edition pg 189.