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Estimation of Liver Glycogen

Eggplant Solanum Melongena

Highlights

Rezent Zombie-Worms Osedax

Control Methods against Hard Ticks

Discovering Thoughts, Inventing Future

VOLUME 16 ISSUE 2 VERSION 1.0



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CONTENTS OF THE ISSUE

- i. Copyright Notice
 - ii. Editorial Board Members
 - iii. Chief Author and Dean
 - iv. Contents of the Issue
-
1. First Global Survey of Evidences the Ichnogenus *Osedacoides* and it Relates to the Rezent Zombie-Worms *Osedax*. **1-5**
 2. Fungi Associated with Pre-Harvest Deterioration of Egg Plant *Solanum Melongena* L. and their Control using Fruit Extract of *Tetrapleura Tetraptera*. **7-15**
 3. Estimation of Liver Glycogen in Normal Control, Diabetic Control and *Tinospora Cordifolia* Extract Treated Albino Rats. **17-20**
 4. *Parkia Biglobosa* Jacq (dawa-dawa): The Threatened Giant of the Guinea Savanna of Nigeria (The Cross River State Situation). **21-32**
 5. Overview of the Biology, Epidemiology and Control Methods Against Hard Ticks: A Review. **33-45**
-
- v. Fellows
 - vi. Auxiliary Memberships
 - vii. Process of Submission of Research Paper
 - viii. Preferred Author Guidelines
 - ix. Index



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First Global Survey of Evidences the Ichnogenus *Osedacoides* and it Relates to the Rezent Zombie-Worms *Osedax*

By Hans-Volker Karl

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Abstract- In this paper, I present the first global overview of the members of fossil ichnotaxon *Osedacoides* and it recent relatives of the morphogenus *Osedax*. Close on 15 years ago those worms were found on sea floor fallen whalebones that dissolve last recyclers in the food chain to the bone [25]. Before that time it was simply technically not possible to observe a whale carcass over a longer period of time and to study the dynamics of the submarine carcasse- societies. Probably the *Osedacoides* do not specialize in whales, but they did also in past geological ages already been feasting on carcasses, as boreholes show in fossils. The main focus of this work is on the fossil evidences.

Keywords: *ichnofossils, cladistic analysis, osedax like worms, osedacoides, overview.*

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First Global Survey of Evidences the Ichnogenus *Osedacoides* and it Relates to the Rezent Zombie-Worms *Osedax*

Hans-Volker Karl

Abstract- In this paper, I present the first global overview of the members of fossil ichnotaxon *Osedacoides* and it recent relatives of the morphogenus *Osedax*. Close on 15 years ago those worms were found on sea floor fallen whalebones that dissolve last recyclers in the food chain to the bone [25]. Before that time it was simply technically not possible to observe a whale carcass over a longer period of time and to study the dynamics of the submarine carcasse- societies. Probably the *Osedacoides* do not specialize in whales, but they did also in past geological ages already been feasting on carcasses, as boreholes show in fossils. The main focus of this work is on the fossil evidences.

Keywords: *ichnofossils, cladistic analysis, osedax like worms, osedacoides, overview.*

1. INTRODUCTION

The Recent osteophagous marine “Zombie- worm” morphogenus *Osedax* (Polychaeta) was originally discovered in 2002 with two species on the ocean bottom in a depth of approximately, 2.800 m in bones of whales where it produces borings, by now, a dozen are described. Up to now it was believed that it co-evolved with the whales, the source of its preferred diet [8,10]. The cetacean size did not determine the whale-fall communities [25]. In the meantime it could be also experimentally cultivated in cattle bones [13]. This fact as well as the reconstructed molecular clock back to at least the Cretaceous period led to the conclusion that *Osedax* originally might have infested fish or marine “reptiles” long before the marine Mammalia evolved [3,15]. In spite of the fact that the borings most probably were produced by an organism similar to *Osedax* there is no direct evidence that it was the genus *Osedax* itself. There are only ichnofossils known, but not body remains of the animal. A definite assignment of the borings to a particular body-based taxon (morphotaxon) is impossible. In agreement with Bromley [2] the name of the Recent morphospecies *Osedax* should no more be used for fossil borings in vertebrate bones [15], therefore they prefers to establish a new ichnotaxon *Osedacoides* for these fossils, and follows Bromley [2] who proposed not to use the names of the animals but to establish ichnotaxa for borings in hard substrates, even if the producers (like for example Bivalvia or

Bryozoa) can be largely ascertained by their typical morphology. Bromley [2] used the ichnogenus *Trypanites* Mägdefrau, [20] as an example, which he defined as simple shaft- or pocket-like borings with a single opening whose producer is not incontestably known. Based upon his revised diagnosis, and calibrated by its type ichnospecies *Trypanites weisei* Mägdefrau, (“Simple, unbranched borings in hard substrate with a single opening to the surface”) *Trypanites* can now be generally used in a wider sense for such borings. The principal difference between the two ichnotaxa is: *Trypanites* is mostly in inorganic substrates, *Osedacoides* exclusively in organic substrates (Fig. 1-C). According Muniz et al. [24] the morphology of *Trypanites ionasi* exhibits a proximal, cylindrical, smooth-walled portion of the fossil boring with the deeper, enlarged and more irregular portion may have accommodated the bulbous root-structure. These are features of *Osedacoides* [14]. In compare, Furlong et al. [5] describes a high diversity of biota and low diversity, but high abundance, of borings is present along a modern *Trypanites*-type ichnofacies (Fig. 1-A). Species richness reaches 37 organisms within the study area, and two boring bivalves (*Petricola pholadiformis* and *Zirfaea pilsbryi*), which produce *Gastrochaenolites*-like traces (Fig. 1-B). *Trypanites* isp. seems to descend from cemented strata directly to crocodilian remains also, which occur in massive sandstone bodies [9,28].

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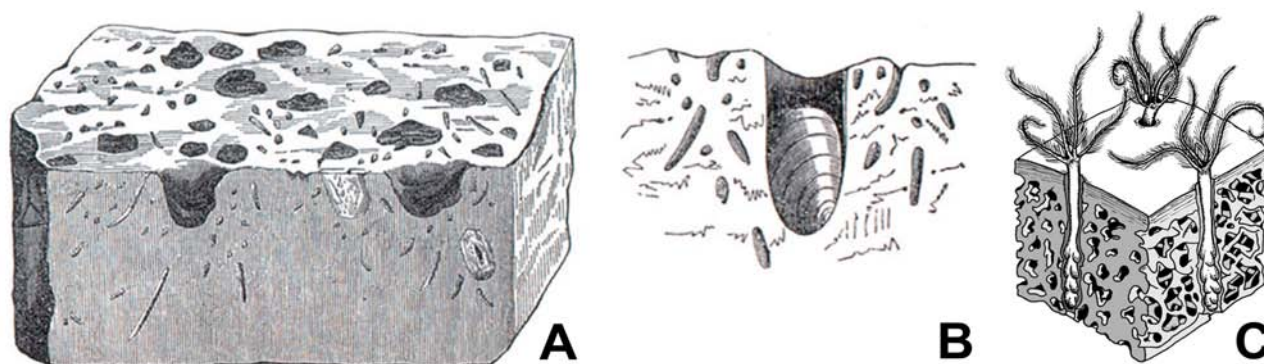


Figure 1 : A - *Trypanites*, B - *Gastrochaenolites*, Jurassic rocky-shore trace fossils from England according De La Beche [5], C - *Osedacoides*, Pliocene whalebone according Muniz et al. [23]

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2

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II. RELATIONS BETWEEN *OSEDAX* LIKE BORINGS AND ICHNOFOSSILS

Systematics: Kingdom Animalia, Phylum Annelida, Class Polychaeta, Order Sabellida, Family Siboglinidae, Morphogenus *Osedax* Rouse et al., 2004, Type species *Osedax rubiplumus* Rouse, Goffredi & Vrijenhoek, 2004. *Osedax rubiplumus* is a species of bathypelagic Polychaetes that is reported to sustain itself on the bones of falling whales. Their paedomorphic males are 0.4–1.1 millimeters (0.016–0.043 in), and have an incomplete prototroch with a posterior hooked chaete [26]. The species have 16 hooks with 6–8 capitulum teeth, which have handles that are 18–23 micrometers (0.00071–0.00091 in). The female ovisac is measured 8 mm by 4 mm by 0.3 mm, with four posterior roots, which have spherical lobes. They also have a trunk, which is 3.8 centimeters (1.5 in) in length and 2 millimeters (0.079 in) wide with the crown plumes, which are 2.1 centimeters (0.83 in) in length. The species is found in East North Pacific where it is abundant. They are used in Calmodulin (CaM, an abbreviation for calcium-modulated protein), a calcium-binding messenger protein expressed in all eukaryotic cells [27]. Higgs et al. [10] published modern *Osedax* traces and also used CT scans to construct 3d models of the borings, and reported borings that were roughly similar to that reported by Kiel et al. [15,18,19,20]. Higgs et al. [10] further found that the borings were mostly restricted to dense cortical bone, generally avoiding lipid-rich cancellous zones. Apparently some isotopic evidence suggests that *Osedax* synthesizes collagen rather than lipids, although other studies have documented *Osedax* in Japanese waters that subsist on blubber in spermaceti [10,11,12]. The hitherto described and named *Osedacoides* ichnofossils are:

a) *Ichnogenus Osedacoides* Karl, Brauckmann & Groening, 2012

LSIDurn:lsid:zoobank.org:act:527977AE-F51F-436D-81AC-A7F75E924913

Type ichnospecies: *Osedacoides jurassicus* Karl, Brauckmann & Groening, 2012

Etymology: *Osedacoides* = *Osedax*-like.

Diagnosis: Simple, basally thickened to branched borings in marine vertebrate bones with a single opening to the surface.

b) *Osedacoides jurassicus* Karl, Groening & Brauckmann 2012

LSIDurn:lsid:zoobank.org:act:371B736C-4E88-4F88-9CDA-E01C46143FB4

Holotype: Geozentrum Göttingen-GZG.V.773-34: Borings in hyoplastron sin. (MAACK [21]: plate 35, fig. 35 = original of *Stylemys lindenensis*, now: *Tropidemys seebachi*, pl. 2 figs. 3-4, pl. 3-4) [19].

Etymology: *jurassicus* = Jurassic, the type stratum.

Locality: Lindener Berg, Hanover, Lower Saxony, Germany.

Horizon: Middle Kimmeridgian, Late Jurassic.

Description: Average diameter of openings 1 - 3, 5 mm, mode of life in groups.

c) *Osedacoides cretaceous* Karl & Niehuys 2012

LSIDurn:lsid:zoobank.org:act:377D8C46-93CF-4451-A57E-1C6D1BF70C0C

Holotype: BGR- Federal Institute for Geology and Minerals- LBEG Hanover, Germany, leg. and coll. Mosbach, 1915, old no. Gr.A.24 no. 8: One boring in peripheral plate of *Ctenochelys stenopurus* described by Karl & Niehuys [14] at their page 174 (4) and illustrated at plate 4, fig. 1- 4).

Etymology: *cretaceous* = Cretaceous, the type stratum.

Locality: Alsen quarry in Lägerdorf near Itzehoe, 30 m (TK 25: 2123 Lägerdorf), Germany.

Horizon: Lägerdorf-Formation, Chalk Group (Schreibkreide- Gruppe), Unter campanian-Obersantonian, Upper Cretaceous (Lithostratigraphic units of Germany ID: 2008074).

Description: Average diameter of opening = 10, 5x 10, 1 mm, inner diameter = 15, 5 mm and deep = 19, 5 mm (pl. 4 fig. 4), single mode of life.

d) *Osedacoides ionasi* (Muniz, de Gibert & Esperante 2010)

LSID urn:lsid:zoobank.org:pub:A3A8A74F-4C5F-40DF-B3BD-F2A3B76CD19C (under work)

Original ichnospecies: *Trypanites ionasi* Muniz, de Gibert & Esperante 2010

Holotype: Muniz et al. 2010: 271, figure 2: Arrows in A and B indicate the specimen selected as the holotype.

Etymology: *ionasi* = from the name Jonas.

Locality: Almería, Spain.

Horizon: Middle platform, lower Pliocene.

Description: Average diameter of opening 0.9 mm, inner diameter 1.9 mm and deep 40 mm, mode of life in groups.

III. CLADISTIC ANALYSIS

Character coding

1 - life in anorganic substrate= 0, or organic substrate= 1; 2 - single mode of life= 0, or mode of life

in groups= 1; 3 - diameter of openings and inner diameter lesser than 10 mm= 0, or more than 10 mm= 1; 4 - inner diameter slightly larger than opening= 0, or significantly larger than opening= 1; 5 - deep equal to or slightly greater than the greatest diameter= 0, or more times greater= 1.

Data matrix

*T.weisei*00001, *O. jurassicus*11001, *O. cretaceous*10110, *O. ionasi*11011

The character differentiation with DOLMOVE (Interactive Dollo and Polymorphism Parsimony by JOSEPH FELSENSTEIN, 1986a) shows a simple tree: (*O. ionasi*, (*O. cretaceous*, *O. jurassicus*, *T. weisei*))) this calculated with PARS (Discrete character parsimony algorithm, version 3.6a3 by JOSEPH FELSENSTEIN, 1986b) is conform with that. Two most parsimonious trees found: (*O. cretaceous*:2.50, (*O. ionasi*:0.50, *O. jurassicus*:0.50):1.00, *T. weisei*:1.50)[0.5000];

((*O. ionasi*:0.50, *O. cretaceous*:2.50):1.00, *O. jurassicus*:0.50, *T. weisei*:1.50)[0.5000]; that are clear specific differentiation in the genus *Osedacoides*. Two most parsimonious tree found and requires a total of 6.000 in tree A (fig. 2-A)

between	and	length
1	<i>cretaceous</i>	2.50
1	2	1.00
2	<i>ionasi</i>	0.50
2	<i>jurassicus</i>	0.50
1	<i>weisei</i>	1.50
Also requires a total of 6.000 in tree B (fig. 2-B)		
between	and	length
1	2	1.00
2	<i>ionasi</i>	0.50
2	<i>cretaceous</i>	2.50
1	<i>jurassicus</i>	0.50
1	<i>weisei</i>	1.50

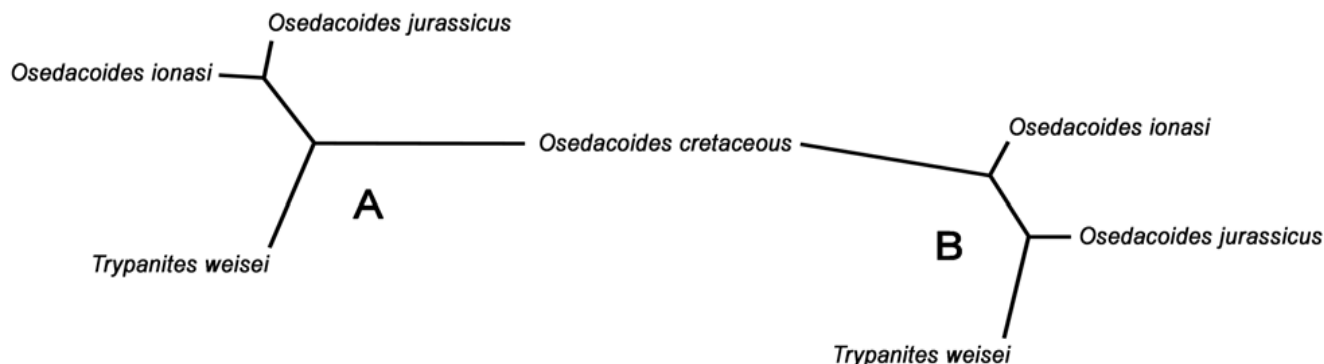


Figure 2 : Cladogram of the hitherto described *Osedacoides* ichnospecies) made with TreeView©Roderic Page. A - Unrooted tree A shows a larger distance of *O. ionasi* and *O. jurassicus* to *O. cretaceous*; B - Tree B construction of the same calculation by PARS shows a larger distance of *O. ionasi* and *O. cretaceous* to *O. jurassicus*.

IV. RESULTS

A shell remain of the quite recently re-discovered type material of the upper Jurassic turtle *Tropidemys seebachi* Portis, 1878 was covered by borings of presumed marine "worms" similar to the Recent *Osedax* for which the new ichnotaxon *Osedacoides jurassicus* was introduced. One another

turtle shell remain, a peripheral plate of the upper Cretaceous *Ctenochelys stenoporus* shows a boring of *Osedacoides cretaceous*, which differs to the type species *Osedacoides jurassicus* with the larger dimensions and the single mode of life. An overview of the corresponding fossil ichnofossils hitherto known is given here:

Age	Horizon	Ichnotaxon	Source	Material
Neogene	Pliocene	<i>Osedacoides ionasi</i>	[14,23]	whalebones
		undescribed	[12]	whalebones
	Miocene	undescribed	[1]	whalebones
Palaeogene	Oligocene	undescribed	[19]	whalebone and teeth fishbone
		undescribed	[18]	whalebones
		undescribed	[19]	birdbones
Upper Cretaceous	Cenomanian	<i>Osedacoides cretaceous</i>	[14,16]	sea turtlebones
		undescribed	[3]	sea turtlebones
Early Cretaceous	Albian	undescribed	[3]	plesiosaurbones
Late Jurassic	Kimmeridgian	<i>Osedacoides jurassicus</i>	[14,15]	sea turtlebones
	Oxfordian	undescribed	[3]	ichthyosaurbones

The examination of the Late Jurassic *Osedacoides jurassicus* by Karl et al. [15] shows that *Osedax*-like marine animals lived even much earlier. This would be another strong reference to the fact that the specialization on whalebones evolved secondarily and a long time later. Additionally, the occurrence of *Osedax*-like animals in the Late Jurassic epicontinental sea in North Germany shows that such organisms were originally not restricted to the deep-sea. This is supported by the discovery of a third recent species in 2005, which fed on whalebones in a depth of about 120 m. As osteophagous polychaetes *Osedax* belongs to the decomposing animals among the carcass feeders. As a detritus feeder *Osedax* might not have been closely adapted to particular hosts like a true parasite. *Osedacoides ionasi* from the Pliocene of Spain may represent the first trace-fossil evidence of an *Osedax*-like behavior in whalebones. The first record in bones of birds [19] proves that *Osedax* evidently feed upon penguin-like diving birds. As already mentioned the environmental reconstruction of the Lägerdorf Formation shows pelagic sediments of an open epicontinental sea with 100-150 m water depth. Recent *Osedax*-species fed on whalebones in a depth of about 120 m, but the occurrence of *Osedax*-like animals in the Late Jurassic epicontinental sea in North Germany shows that such organisms were originally not restricted to the deep-sea [15]. The deeper marine areas were not the habitat of the turtles alive. These are only post mortem dropped to the ground. The bones were fragmented before their embedding. The surfaces of all bones showing erosions, which are typical for necrophages animals, such as cancers, echinoderms, worms, snails, mussels or lampreys. One bone fragment shows bite marks. Also fossil traces of *Osedax* like borings from Late Cretaceous plesiosaur and sea turtle bones reports of

Danise & Higgs [3], and from early Cretaceous by Danise et al. [6] from ichthyosaur bones.

V. ACKNOWLEDGEMENT

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VI. PHYLOGENETIC PROGRAMS

- I. Felsenstein, J. (1986a): PHYLIP/ DOLMOVE-Interactive Dollo and Polymorphism Parsimony © Copyright 1986-2002 by the University of Washington.
- II. Felsenstein, J. (1986b): PHYLIP/ PARS-Discrete character parsimony © Copyright 1986-2000 by the University of Washington.
- III. TreeView©Roderic Page

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Fungi Associated with Pre-Harvest Deterioration of Egg Plant *Solanum melongena* L. and their Control using Fruit Extract of *Tetrapleura tetraaptera*

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Abstract- This study was carried out to investigate the fungi associated with pre-harvest fruit rot of eggplant (*Solanum melongena* L.), their effect on fruit nutritional content and their control using fruit extracts of *Tetrapleura tetraaptera* in- vitro. The fungal pathogens isolated as the causative agents of fruit rot in this study were *Phomopsis melongena* and *Collectotrichum melongena*. The result of proximate analysis of fungal infected and non-infected eggplant carried out showed that there was an increase in the moisture and protein content of the fungal infected eggplant as compared to healthy ones (control), while there was a decrease in the crude fibre, fat, ash, and carbohydrate contents of the fungal infected eggplant fruits as compared to the healthy ones (control).

GJSFR-C Classification : FOR Code: 069999



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Fungi Associated with Pre-Harvest Deterioration of Egg Plant *Solanum melongena* L. and their Control using Fruit Extract of *Tetrapleura tetraaptera*

Akwaji, Patrick Ishoro ^α, Okon, Ekeng Ita ^σ, Markson, Aniedi-Abasi Akpan ^ρ & Iso, Usang Ekong ^ω

Abstract- This study was carried out to investigate the fungi associated with pre-harvest fruit rot of eggplant (*Solanum melongena* L.), their effect on fruit nutritional content and their control using fruit extracts of *Tetrapleura tetraaptera* *in-vitro*. The fungal pathogens isolated as the causative agents of fruit rot in this study were *Phomopsis melongenae* and *Collectotrichum melongenae*. The result of proximate analysis of fungal infected and non-infected eggplant carried out showed that there was an increase in the moisture and protein content of the fungal infected eggplant as compared to healthy ones (control), while there was a decrease in the crude fibre, fat, ash, and carbohydrate contents of the fungal infected eggplant fruits as compared to the healthy ones (control). It was observed that moisture content increased from 42.90 ± 0.12 in the non-infected eggplant fruit to 58.03 ± 0.20 in the eggplant fruit infected with *Phomopsis melongenae* and 42.90 ± 0.12 in the non-infected eggplant to 60.12 ± 0.40 when infected with *Collectotrichum melongenae* while the following parameters viz; protein 5.60 ± 0.10 , fat 5.37 ± 0.17 , crude fibre 5.12 ± 0.09 , carbohydrate 3.64 ± 0.19 and ash content 0.50 ± 0.15 of non-infected eggplant were depleted when infected with *Phomopsis melongenae* to 1.40 ± 0.18 , 5.60 ± 0.10 , 2.98 ± 0.27 , 5.37 ± 0.17 , and 0.25 ± 0.16 and protein 1.60 ± 1.11 , fat 3.0 ± 0.31 , crude fibre 3.30 ± 0.7 , carbohydrate 1.09 ± 1.21 , and ash content 0.30 ± 0.21 of non-infected eggplant were depleted to 5.60 ± 0.10 , 5.37 ± 0.17 , 5.12 ± 0.09 , 3.64 ± 0.19 respectively when infected with *Collectotrichum melongenae*. Results of the *in vitro* antifungal assay carried out showed that the ethanolic extracts of *Tetrapleura tetraaptera* had a significant effect ($p < 0.05$) in inhibiting the radial growth of the fungal pathogens at the different concentrations (5g/100ml, 10g/100ml and 15g/100ml) tested than the aqueous extracts. Phytochemical test carried out showed that tannin, flavonoid, saponin and cyanogenic glycosides were present in aqueous extract while alkaloids, steroids, triterpens, tannin, flavonoid and saponin were present in the ethanolic extracts.

1. INTRODUCTION

Eggplant (*Solanum melongena* L.), family Solanaceae is a popular vegetable also known as African eggplant which contains numerous small soft seeds which although edible, taste better because, the plant related to tobacco, and contains nicotinoids

and alkaloids and commonly served and eaten as desert mostly with groundnut in this part of the world (Giuliani and Smale, 2000).

It is one of the top ten vegetables in the world and it is grown on more than two million hectare with a production of nearly thirty three million tones (FAO, 2007). The plant had been cultivated in India for the past four thousand years and is one of the most important vegetable of Solanaceae family. The global area under eggplant cultivation has been estimated to be at 1.85 million hectare with total production of about 32 million metric tons, it is grown on nearly 550,000 hectares in India, making the country as the second largest producers after China. India accounts for about 8.7 million metric tons with an area of about 0.53 million hectares under cultivation (Anon, 1998, Sidhu, 1998).

The northern and southern parts of Nigeria are involved in the cultivation of eggplant which comes in different varieties and they vary in fruit colour, shapes and sizes (Chinedu *et al.*, 2011).

Egg plant is well adapted to high rainfall and high temperatures, and is among the few vegetables capable of high yields in hot- wet environments (Hanson *et al.*, 2006). Eggplant contains nutrients such as dietary fiber, folate, ascorbic acids, vitamin K, niacin, vitamin B₆, pantothenic acids, potassium, iron, magnesium, manganese, phosphorus and copper (USDA, 2009); and the nutrients contribute to the diet of the poor and mostly important during time when other vegetables are in short supply.

Eggplant is a delicate tropical perennial, often cultivated as a tender or half-hardy annual in temperate climates (Grubben and Denton, 2004). It grows 40 – 150 cm tall, with large coarsely lobed leaves that are 10 – 20 cm long and 5 – 10 cm broad (Grubben and Denton, 2004.) semi wide types can grow much larger, up to 255 cm with larger leaves, over 30 cm long and 15 cm broad, the stem is often spiny and the flowers are white to purple with a five lobed corolla, yellow stamens and the egg shaped glossy purple fruit has white fleshy with a meaty texture, and the cut surface of the flesh rapidly turns brown when the fruit is cut open (Hanson *et al.*, 2006).

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Eggplant fruit and leaves are both eaten as vegetables or used in traditional medicine (Bonsuet *al.*, 2008). Eggplants are highly valued constituents of Nigerian foods and indigenous medicine that are either eaten raw or cooked. Also very popular in rice dishes such as stew and soup of different kind, and also prepared as sauces that are consumed with yam and plantain (Edemetal., 2009).

In most advanced countries, such as China, India, Philippines eggplant has been used in their indigenous medicine, the medicinal uses ranges from weight reduction to treatment of several ailments which include asthma, constipation and skin infections, diabetes, leprosy, gonorrhea, dysuria, dysentery, asthenia and hemorrhoids (Gill, 1992).

Diseases and fungal pathogens of eggplant include damping-off caused by (*Pythium*spp, *Phytophthora*spp: and *Fusarium*spp) root rot (*Rhizoctonia*spp and *Sclerotium*spp). Blight (*Phomopsis*spp); fruit rot (*Phomopsisvexans* and *Rhizopusstolonifor*) and wilt (*Verticillium*spp; *Fusarium*spp) are noteworthy and take a considerable proportion of the produce annually. Eggplant wilt complex is known to be caused by number of fungi genera such as *Fusarium*, *Verticillium*, *Rhizoctonia*, *Sclerotium* and *Phytophthora* in different parts of the world (Rangas-wami 2000).

In view of the adverse effect of fungi infection on eggplant fruits as observed in the University of Calabar Farm, University of Calabar, Calabar, Cross River State, Nigeria and environs it became necessary to isolate and identify the fungal pathogens associated with the eggplant fruits in the field, determine the effect of the pathogens on the nutritional content of the eggplant fruits through proximate analysis as well as evaluate the phytochemical and antifungal effect of *Tetrapleura-tetraptera* fruit extracts on the isolated fungi *in vitro*.

II. MATERIALS AND METHODS

a) Collection of Samples

Healthy and infected eggplant fruits were obtained from the University of Calabar Farm, University of Calabar, Calabar Cross Rivers State, Nigeria. Proximate (nutrient) analysis of infected and non-infected eggplant fruits was carried out in the Department of Biochemistry, University of Calabar, Calabar, Nigeria.

b) Source of fungal pathogens and morphological identification

The fungal pathogens used in this research work were isolated from diseased eggplant fruits collected from the University of Calabar Farm, Calabar, Cross River State, Nigeria. Cut sections of the diseased assay fruits were surface sterilized with 70% sodium hypochlorite (bleach) solution for 1min and rinsed quickly in 3 changes of sterile distilled water, blotted dry on Whatman's No. 1filter paper and placed on Potato

Dextrose Agar (PDA) in Petri dishes. Four (4) sections were inoculated per Petri dish. The plates were incubated at $28 \pm 1^\circ\text{C}$ until fungal growth was noticed. After 5 days, the different isolates were sub-cultured on freshly prepared PDA to obtain their pure culture. Isolated fungi were microscopically (Olympus optical, Phillipines) identified as far as possible using the identification guides of the International Mycological Institute, Kew and of Barnett and Hunter (1998), Alexopolous and Mins (1989).

c) Pathogenicity Test

Pathogenicity tests were carried out using the techniques of Okigbo and Nmeko,(2005). Healthy eggplant fruits were washed in distilled water and surface sterilized with 1% Sodium hypochlorite solution. A5mm diameter cork borer was used to cut discs from the fruits (three discs per fruit) and cultures of the isolated discs were introduced into holes and replaced with the discs. They were kept for 24-48hours. The inoculated fruits established symptoms on the second day and tissue segments from the infected fruits were cultured.

d) Effect of Fungal Infection on Proximate Composition of Eggplant fruit

To ascertain the effects of fungal infection on nutritional composition of egg plants, matured eggplant fruits showing signs of fungal infection were obtained and the moisture content of the eggplant fruits were determined. The plant fruit were then oven dried at 60°C for 24 hrs and grounded into fine powder using mortar and pestle. The powdered samples were stored in plastic container for laboratory analysis. Most of the methods adopted in this research work were those recommended by Association of Official Analytical Chemist (AOAC 2002).

e) Proximate Analysis of Moisture Content

A clean 100 ml beaker was dried in an oven to constant weight (a). A known amount of the 5 g sample was introduced in the beaker and weighed (b). The samples were then fried in a ventilated electrically heated atmosphere oven at 75°C for about 24hrs and cooled in a desiccators until constant weight was obtained (c). The Percentage Moisture content was calculated from the formula:

$$\% \text{ Moisture content} = \frac{\text{Weight loss of sample} \times 100}{\text{Weigh of the original sample 1}}$$

The experiment was carried out in triplicates.

f) Ash content

5 kg sample were accurately weighed into the crucible. This was ignited at 55°C for about 24 hrs in desiccators and weighted. This step was repeated until a constant weight was obtained. The percentage ash content was calculated from:

$$\% \text{ Ash content} = \frac{\text{Weight of ash} \times 100}{\text{Weight of sample 1}}$$

Determination was made in triplicate.

g) *Crude fat or ether extract*

5 g samples were accurately weighed into a thimble. About 120 ml petroleum ether was poured into a previously dried and weighted round bottom flask. The Soxhlet extractor into which the thimble with content had been introduced was then filled into the round bottom flask and the condenser and extraction apparatus set up with a clamp and stand. Gentle heat had been applied then the heater evaporated and as it condensed, it dropped into the thimble where it extracted ether soluble constituents into the round bottomed flask. The extraction then continued for about 8 hours. The thimble was then removed and air-dried (later far free extract was used for fibre determination). The petroleum ether in the flask was distilled off and collected in the Soxhlet extractor tube. The flask was then fried in an air circulating desiccators for 8 hours. The round bottom flask and the lipid extract were then weighted. The flask and content was again dried and weighed till a constant weight was obtained. The amount of lipid extracted was obtained from the difference between the weight of the flask before and after extraction.

$$\% \text{ Fat} = \frac{\text{Weight loss of sample (extracted fat)} \times 100}{\text{Weight of sample 1}}$$

h) *Crude fibre*

5 g far free material was weighted and quantitatively transferred into 400 ml beaker, which had been previously marked at 200 ml level. 50 ml of 1.25% sulphuric acid were added and the mixture was made up to 200 ml mark with distilled water. The contents of the beaker were heated to boiling point for 30 minutes.

i) *Crude Protein (Micro Kjeldahl Method)*

40% Sodium hydroxide (NaOH) pellets (40 g pellets carbonates free were dissolved in 100 ml distilled water). Concentrated sulphuric acid (H₂SO₄), Selenium Kjeldahl Catalyst (each tablet containing 1g Sodium sulphate, and 0.05 g Copper sulphate (CuSO₄) was dissolved in 0.1% hydrochloric acid (HCL). Methyl red-ethylene blue indicator was prepared by mixing the equal volume of 0.2% twice recrystallized methyl red and 0.0% methylene blue made up in absolute ethanol. This sample was then stored in a dark brown bottle in a refrigerator.

j) *Digestion (Micro Kjeldahl)*

1 g sample was weighed out into a 50 ml Kjeldahl digestion flask. 20 ml of antidumping chips were added. The mixture was incinerated to gentle boiling on a digestion rack and then heated strongly until the digest became clear. The digest was removed, cooled and quantitatively transferred to a 100 ml volumetric flask and made up to mark. An Erlenmeyer flask

containing 10 ml of boric acid indicator solution was placed at the tip of the condenser extended below the surface of the solution. 10 ml of the sample digest was introduced into quick fit micro Kjeldahl flask and steam heated. 10 ml of 40% Sodium hydroxide (NaOH) solution was added to the digest and the digested steam distilled into the Erlenmeyer's flask until the contents become more than double of its original volume as the ammonia (NH₃) changed to green. A blank determination was carried out in a similar manner as described above except 1 g digestion sample was replaced by 1 ml of distilled water.

k) *Titration*

The content of the Erlenmeyer flask was titrated with 0.1% hydrochloric acid to a pink end point.

Calculation

$$1 \text{ ml of HCl (Test)} - \text{ml of HCl (Blank)} \times \text{NX} \times 100 \% \\ \text{Protein} = 1000 \times 10 \times 1$$

N = Normality of the acid

10 = Ml of digest use

I = Gram of sample used

III. PREPARATION OF PLANT MATERIALS

Dried fruits used in the study were separately washed thoroughly using distilled water and surface sterilized with 70% ethanol and sun-dried for 3 days. The dried plant fruits were blended separately using a sterile electric blender to obtain 200grams of fine powder of each fruit. Aqueous extracts of fruits were obtained by adding the dried powder (blended) of plant material to distilled water at room temperature 28±1°C. Three levels of concentrations were obtained by dissolving 5g, 10g and 15g of each sample with 100ml of distilled water. This was vigorously stirred and allowed for 24 hours. The solution was then filtered through four-folds of sterile cheese cloth for all the plant materials. The filtrates obtained were used as aqueous extracts of the test plants and stored in reagent bottles for further use. Ethanolic extracts of plant materials were obtained by adding the powdered sample at different concentrations, 5g, 10g and 15g to 100mls of ethanol. This was stirred vigorously and allowed for 24 hours at room temperature 28±1°C. The solution was then filtered through four-folds of sterile cheese cloth for all the plant materials. The filtrates obtained were used as ethanolic extracts of the test plants and stored in reagent bottles for further use.

a) *Susceptibility test*

The extracts percentage concentrations were prepared at 5g/100ml, 10g/100ml and 15g/100ml with ethanol and water as solvent.

b) *In vitro antifungal assay*

5ml of each concentration of both the aqueous and ethanol extracts was first poured into different Petri-

dishes using sterile syringe. The sterile potato dextrose agar (PDA) was also poured into the plates containing the solvent extracts after which the plates containing the solvent extracts were gently swirled to ensure mixing. The media was allowed to solidify and with a sterilized cork borer (5mm in diameter), a disc of the matured culture was punched out from advancing margin of a four- day old pure culture and inoculated at the center of plates and incubated at room temperature ($28 \pm 10^\circ\text{C}$) for 7 days. The experiment was replicated thrice. Area of inhibition was measured daily for 7days using a meter rule and recorded.

c) Phytochemical Screening

Phychemical screening of the aqueous and ethanolic fruit extract of *Tetrapleuratetraptera* was carried out using the method of (Harborn, 1973). Phytochemical screening was carried out in the Department of Biochemistry, University of Calabar, Calabar, Cross River State, Nigeria.

d) Statistical analysis

Data obtained in this study were analyzed using Student T-test and a one way Analysis of Variance (ANOVA) at 5% probability level ($p < 0.05$).

IV. RESULTS

a) Isolated fungal pathogens

The fungal pathogens isolated and identified as the causative agents of pre-harvest eggplant fruit rot from this study were: *Collectotrichummelongenae* and *Phomopsismelongenae*.

b) Pathogenicity Test

Symptoms observed on the fruits inoculated with *Collectotrichummelongenae* and *Phomopsis-melongenae* were similar to those observed on the

rotted eggplant fruit obtained from the field. Symptoms such as soft rots and lesions were observed on the fruits.

c) Effect of fungi infection on biochemical composition of the Eggplant

The results of proximate analysis in mg/100 g of *P. melongenae* infected eggplant fruits showed an increase in the moisture content of the fungal infected fruits of eggplant as compared to the healthy ones (control), whereas there was a decrease in the carbohydrate, fat, fibre and ash contents of the fungal infected fruits relative to the healthy ones (control). Moisture content increased from 42.90 ± 0.12 in the non-infected eggplant to 58.03 ± 0.20 in the infected eggplant fruit with *Phomopsismelongenae* and protein content increased from 1.40 ± 0.18 in the Post-infected eggplant to 5.60 ± 0.10 in the non-infected fruits while the following parameters were found to decrease in the infected than in the non-infected fruits viz carbohydrate content 1.02 ± 0.29 , fat 2.98 ± 0.25 , crude fibre 3.09 ± 0.06 , protein 1.40 ± 0.10 and ash content 0.25 ± 0.16 as presented in (Table 1). The results of proximate analysis in mg/100 g of *C. melongenae* infected eggplant fruits showed that there was an increase in the moisture content from (42.90 ± 0.12 to 60.12 ± 0.40) when infected with *Collectotrichummelongenae* and from (1.60 ± 0.11) in the Cs-infected garden egg to (5.60 ± 0.10) in the non-infected eggplant when infected with *Collectotrichummelongenae* while the following parameters were found to decrease when infected with *Collectotrichummelongenae* than in the non-infected fruits viz carbohydrate content (1.09 ± 0.21), fat (3.0 ± 0.31), crude fibre (3.30 ± 0.7), and ash content (0.30 ± 0.21) as presented in (Table 2).

Table 1 : Proximate composition of *Phomopsismelongenae* infected and non-infected eggplant mg/100g (dry matter)

Sample	Moisture	C/F	Fat	Protein	Ash	CHO
Post-infected	58.03 ± 0.20^a	3.09 ± 0.06^b	2.98 ± 0.27^b	1.40 ± 0.18^a	0.25 ± 0.16^b	1.02 ± 0.29^b
Non-infected	42.90 ± 0.12^b	5.12 ± 0.09^b	5.37 ± 0.17^b	5.60 ± 0.10^b	0.50 ± 0.15^b	3.64 ± 0.19^b
T-value	3.54					

Note: C/F = Crude fibre, CHO = Carbohydrate, Ps = *Phomopsismelongenae*

Table 2 : Proximate composition of *Collectotrichummelongenae* infected and non-infected eggplant mg/100g (dry matter).

Sample	Moisture	C/F	Fat	Protein	Ash	CHO
Cs-infected	60.12 ± 0.40^a	3.30 ± 0.7^a	3.0 ± 0.31^a	1.60 ± 1.11^a	0.30 ± 0.21^a	1.09 ± 1.21^b
Non-infected	42.90 ± 0.12^b	5.12 ± 0.09^b	5.37 ± 0.17^b	5.60 ± 0.10^b	0.50 ± 0.15^b	3.64 ± 0.19^b
T-value	3.53					

Note: C/F = Crude fibre, CHO = Carbohydrate, Cs = *Collectotrichummelongenae*.

d) *Phytochemical screening*

Phytochemical screening of the aqueous and ethanolic extract of *Tetrapleuratetraptera* showed that cardiac or cyanogenic glycosides, flavonoid, saponin and tannin were present in the aqueous extract while saponin, flavonoid, alkaloids, steroids, triterpens and tannin were present in the ethanolic extracts as presented in (Table 3).

Table 3 : Phytochemical screening of ethanolic and aqueous extract of *Tetrapleuratetraptera*

Phytochemical constituents	Ethanol extract	Water extract
Flavonoid	+	+
saponin	+	+
Tannin	+	+
Alkaloid	+	+
Triterpens	+	-
Cyanogenic glycosides	-	+
Steroids	+	-

Note: (+) = Present, (-) = Absent

e) *In vitro effect of ethanolic extract of Tetrapleura-tetraptera on Colletotrichummelongenae and Phomopsismelongenae at the different concentrations.*

The *in vitro* effect of the ethanolic plant extract at different concentrations on the radial growth of the fungal pathogens is presented in (Tables 4 and 5). Results from the study showed that extract of *Tetrapleuratetraptera* had a significant effect on the isolated fungal pathogens at all levels of concentration (5g/100ml, 10g/100ml and 15g/100ml) tested as compared with the aqueous extracts. Results (Table 4 and 5) showed that *Tetrapleuratetraptera* extract at 5g/100ml concentration completely inhibited the radial growth of *Colletotrichummelongenae* and *Phomopsismelongenae* in the first to fourth day of incubation and at 10g/100ml concentration on the first to fifth day of incubation respectively while at 15g/100ml concentration, the radial growth of *Colletotrichummelongenae* and *Phomopsismelongenae* was completely inhibited throughout the incubation period as compared with the aqueous extracts.

Table 4 : *In vitro* effect of ethanolic *Tetrapleuratetraptera* extract on *Colletotrichummelongenae*.

Concentrations	Days of incubation and radial growth (cm)						
	1	2	3	4	5	6	7
5g/100ml	0.0	0.0	0.0	0.0	0.1	0.2	0.3
10g/100ml	0.0	0.0	0.0	0.0	0.0	0.1	0.2
15g/100ml	0.0	0.0	0.0	0.0	0.0	0.0	0.0
LSD	0.7*						

Note: Values are means of three replicates

Table 5 : *In vitro* effect of ethanolic *Tetrapleuratetraptera* extract on *Phomopsismelongenae*.

Concentrations	Days of incubation and radial growth (cm)						
	1	2	3	4	5	6	7
5g/100ml	0.0	0.0	0.0	0.0	0.1	0.3	0.3
10g/100ml	0.0	0.0	0.0	0.0	0.0	0.1	0.2
15g/100ml	0.0	0.0	0.0	0.0	0.0	0.0	0.0
LSD	0.8*						

Note: Values are means of three replicates

f) *In vitro effect of aqueous extract of Tetrapleura-tetraptera on Colletotrichummelongenae and Phomopsismelongenae at the different concentrations*

The *in vitro* effect of the aqueous plant extracts at the different levels of concentration on the radial growth of the fungal isolates is presented in (Tables 6

and 7). Results from the study showed that aqueous extract of *Tetrapleuratetraptera* had little or no significant inhibitory effect on the isolated fungi (*Colletotrichummelongenae* and *Phomopsismelongenae*) at all levels of concentration (5g/100ml, 10g/100ml and 15g/100ml) tested when compared with the ethanolic extracts.

Table 6 : *Invitro* effect of aqueous *Tetrapleuratetraptera* extract on *Colletotrichummelongenae*

Concentrations	Days of incubation and radial growth (cm)						
	1	2	3	4	5	6	7
5g/100ml	2.4	3.4	4.5	4.5	4.5	4.5	4.5
10g/100ml	2.1	3.6	4.5	4.5	4.5	4.5	4.5
15g/100ml	2.3	3.2	4.0	4.5	4.5	4.5	4.5
LSD	0.96						

Note: Values are means of three replicates

Table 7 : Invitro effect of aqueous *Tetrapleuratetraptera* extract on *Phomopsismelongenae*

Concentrations	Days of incubation and radial growth (cm)						
	1	2	3	4	5	6	7
5g/100ml	2.7	3.0	4.5	4.5	4.5	4.5	4.5
10g/100ml	2.4	2.9	4.5	4.5	4.5	4.5	4.5
15g/100ml	1.7	2.9	4.2	4.3	4.4	4.5	4.5
LSD	1.07						

Note: Values are means of three replicates

V. DISCUSSION

In this study, two fungal pathogens were isolated as causative agents of fruit rot of eggplant (*Solanum melongena*), namely *Colletotricum melongenae* and *Phomopsis melongenae* in the infected fruits. Most of the fungal isolates have been found to be associated with the rots of most fruits and vegetables in Kano (Musa and Buashir, 2013), Ibadan (Adio *et al.*, 2013), Port Harcourt (Chuku and Emelike, 2013, Chuku and Barber, 2013), South Eastern States (Iwuagwu *et al.*, 2013) and in Delta state (Taiga and Eyegbagharen, 2013). Pathogenicity test carried out in this study showed that these fungi actually caused the fruit diseases earlier observed on the fruits in the field. The result of proximate analysis carried out showed that moisture content of infected eggplant fruits increase from 42.90 ± 0.12 – 58.03 ± 0.20 when infected with *Phomopsis melongenae* and from 42.90 ± 1.12 – 60.12 ± 0.40 when infected with *Collectrichum melongenae*. The result of proximate analysis (mg/100 g) of non-infected and infected eggplant revealed that moisture content increased in the infected garden egg fruits, to 5.60 ± 0.10 in the infected. This result is in agreement with the findings of Falaye and Fagbohun (2012) who reported an increase in moisture content from 5.09 in the non-infected to 6.13 in the infected and carbohydrate 5.01 to 5.53 of groundnut (*Arachis hypogea*) infected with *Phomopsis melongenae*. Similarly, Nweke and Ibiam (2012) reported an increase in the moisture and protein content of *Anoniamuricata* fruits infected by *Colletotrichum gloeosporoides* and *Rhizopus stolonifer*. Omokolo *et al.*, (1996) have also found that moisture content of non-infected pods (91.0) have decreased to (13.2) in infected cocoa beans. Onifade and Jeff-Agboola (2003) reported that moisture decreased from (36.49 to 10.4) g/100 g in infected samples. The decrease in fat (12.93 ± 0.26), crude fibre (4.67 ± 0.09), ash content (2.30 ± 0.12) and carbohydrate (1.02 to 0.29) in the infected sample is in agreement with the findings of Shehu and Aliero (2010) have also reported that the infected onion leaf showed a significant decrease in the quantity of the crude protein, fat, fibre and ash content. Opayemi (2012) reported that ash content of non-infected pods (10.7) and beans (8.0) were depleted when infected with *Phytophthora Palmivora* to (9.3) and (7.8) in cocoa pods and beans, respectively. It could therefore be deduced that the

relative increase of moisture in the infected eggplant fruits may be caused by the digestion, degradation and dissolution of the fruit tissue into a mush (water rot) by the pathogens. These degradation activities by pathogens might have also resulted in the relative reduction in the protein, fat, fibre, and ash contents of the infected fruits. The protein, fat, fibre and ash might have been broken down by the fungi into smaller molecules that they absorbed (Nweke and Ibiam 2012). Bonner (1997) reported that complex molecules such as polysaccharide and protein are required by fungi to build the hyphal wall (chitin, glucan and cellulose) and for respiration to obtain energy. Ward and Diener (1991) obtained similar results on groundnut seeds. A decrease in vitamin content of fruits could be due to the increase in moisture content, causing the vitamin to dissolve in it, since it is a water-soluble vitamin. The result of this study shows that the fungi caused deterioration of garden egg fruits and altered the nutritional value of the fruits. In a study conducted by Ameret *et al.*, (2007), the nutritional contents of garden egg fruits and seeds were greatly affected by the presence of the fungal pathogens *Aspergillus flavus* and *Phomopsis melongenae*. This suggests that these pathogens might have denied man of these essential nutrients upon consumption through their degradation activities, thereby causing some great damaging effects to human health. Van Duyn and Pivanka (2000) stated that the deficiency of fibre in our diet leads to diverticular diseases and intestinal cancer. However Fanny *et al.*, (2000) who also reported a decrease in fat, ash and protein content of maize infected by fungi, stated that the nutrient depletion in entire test plant sample might have been as a result of the internal defense system of the host tissue.

Phytochemical screening of the test plant (*Tetrapleuratetraptera*) extract used in this study was carried to determine its exact phytochemical contents. Results of the phytochemical screening carried out showed that cardiac or cyanogenic glycosides, flavonoid, saponin and tannin were present in the aqueous extract while saponin, flavonoid, alkaloids, steroids, triterpens and tannin were present in the ethanolic extracts as presented in (Table 3). Phytochemicals, as compounds which occur naturally in plants, form part of plants defense mechanisms against diseases (Eleazu *et al.*, 2012). They are classified into primary and secondary, based on their activity in plant

metabolism. The primary ones comprise of sugars, amino acids, proteins and chlorophyll (Krishnaiah *et al.*, 2007), while secondary ones include the phenolic compounds such as tannins, flavonoids, alkaloids, saponins, anthraquinones, phlobatannins, proanthocyanidins, etc. (Eleazu *et al.*, 2013). These phenolic compounds have been reported to possess considerable antimicrobial properties, which is attributed to their redox properties (Molan and Faraj, 2010, Zongoet *et al.*, 2011). Thus the antimicrobial properties of plants have been attributed to the presence of these secondary metabolites (Prakash and Hosetti, 2010).

In this study, the antifungal activity of *Tetrapleuratetraptera* fruit extract was tested *in vitro* on fungi isolated from infected fruits of *S. melongenae*. Results showed that the ethanolic and aqueous extracts of the fruit of *T. tetraptera* investigated, exhibited various antifungal activities against the species of fungi isolated. The antifungal activity of the ethanolic and aqueous extracts of *T. tetraptera* on the isolated fungal pathogens is presented in (Tables 4- 7) respectively. The results showed that, the ethanolic extract had a significant ($P < 0.05$) effect on the radial growth of the fungal pathogens than the aqueous extracts and the rate of antifungal activity differed from one concentration to the other. The differences in the fungitoxic potentials between these plant extracts may be attributed to the susceptibility of each of the fungal pathogens to the different plant extracts. This agrees with the results of some workers like Amadioha, (2000) and Okigbo and Nmeka, (2005). Ilondu *et al.*, (2001) reported that some plants contain phenolic substances and essential oils, which are inhibitory to micro-organisms. The presence of these compounds in these extracts has been reported to be responsible for their antifungal properties (Ahmed and Stoll, 1996). It is noteworthy that of all the tested plant extracts (aqueous and ethanolic), ethanolic extracts of *T. tetraptera* had a more significant effect than the aqueous extracts and the level of inhibition increased with a corresponding increase in the concentration of the extracts. Complete inhibition was observed with ethanolic extract of *T. tetraptera* at the highest concentration of 15g/100ml. The inhibitory potency of the plant extracts may be attributed to the phytochemical compounds like tannins, alkaloids, flavonoids and saponins in them as reported by Chiejina and Ukeh (2013). This is also in agreement with the works of Amadioha and Obi, (1999) and Umana *et al.*, 2014, Umana *et al.*, 2015) who reported that the high potency of plant extracts containing the same bio-active compounds could be used for the control of fungal pathogens of plants.

VI. CONCLUSION

The fungal pathogens isolated and identified from this study as the causative agents of fruit rot of

eggplant were *Collectotrichummelongenae* and *Phomopsis melongenae*. The results of proximate analysis of fungal infected and non-infected eggplant fruits showed that there was an increase in the moisture content of the fungal infected fruits of eggplant relative to the healthy ones (control), while there was a decrease in the carbohydrate, fat, fibre, protein and ash contents of the fungal infected fruits relative to the healthy ones (control). It is possible that the isolated fungal pathogens might be resident on the leaves and roots of the plant from where they were dispersed into the fruits to initiate infection spore during rainfall. Results of the *in vitro* antifungal assay carried out showed that the ethanolic extracts of *Tetrapleuratetraptera* were effective against the fruit rot fungal pathogens of eggplant at the different concentrations tested. To this effect, use of resistant seed and timely spraying of egg plant crops with extracts of *Tetrapleuratetraptera* prepared at higher concentrations during flowering and fruiting will reduce the damaging activities of the fungal pathogens and contamination with mycotoxins and other related fungal metabolites that might be hazardous to human health.

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Estimation of Liver Glycogen in Normal Control, Diabetic Control and *Tinospora Cordifolia* Extract Treated Albino Rats

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Abstract- Investigations have been carried out to examine the presence of liver glycogen in normal control, diabetic control and extract treated albino rats. Estimation of liver glycogen was carried out by taking liver samples from normal control, diabetic control and extract treated albino rats. The rats weighing 150-190gm were administered intraperitoneally with 180mg/kg body weight dose of alloxan monohydrate for the induction of diabetes, with alcoholic leaf extract of *Tinospora cordifolia* with a oral dose of 20 ml/kg body weight from day 2 to 30 half an hour prior to feeding twice a day. It produces significant decrease in liver glycogen level.

Keywords: liver glycogen, *tinospora cordifolia*, alloxan, diabetes mellitus.

GJSFR-C Classification : FOR Code: 270599



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Kinkar Shobha Bhanudas ^α & Patil Kishor Gopal ^σ

Abstract- Investigations have been carried out to examine the presence of liver glycogen in normal control, diabetic control and extract treated albino rats. Estimation of liver glycogen was carried out by taking liver samples from normal control, diabetic control and extract treated albino rats. The rats weighing 150-190gm were administered intraperitoneally with 180mg/kg body weight dose of alloxan monohydrate for the induction of diabetes, with alcoholic leaf extract of *Tinospora cordifolia* with a oral dose of 20 ml/kg body weight from day 2 to 30 half an hour prior to feeding twice a day. It produces significant decrease in liver glycogen level.

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1. INTRODUCTION

In the past 200 years, dramatic advances in our understanding of the regulation of normal glucose metabolism have been made. Beginning in the mid-19th century, Claude Bernard showed that blood glucose levels are regulated not just by the absorption of dietary carbohydrate but also by the liver, which plays a central role in producing glucose from non-glucose precursors [1]. Other investigators built on this discovery to identify the enzymes responsible for the synthesis and breakdown of glycogen [2], the role of anterior pituitary hormones in glucose metabolism and the onset of diabetes [3], the role of reversible protein phosphorylation by a protein kinase [4] and the discovery of cyclic AMP and its role in hormonal action, particularly that of epinephrine and glucagon, both of which elevate the blood glucose concentration and contribute to diabetic hyperglycemia [5].

Number of researches have been accomplished experimental diabetes induce in various mammalian species [6, 7, 8, 9, 10].

It is investigated that the whole plant extract of *Tinospora cordifolia* significantly decreases the blood glucose towards the normal blood [11]. Although several therapies are in use for the treatment, there are certain limitations due to high cost and side effects such as development of hyperglycemia, weight gain,

gastrointestinal disturbances, liver toxicity etc.[12]. Based on recent advances and involvement of oxidative stress in complicating diabetes mellitus, efforts are on to find suitable antidiabetic and antioxidant therapy. Present investigations were carried out on albino rat *Rattus norvegicus* due to its metabolic relatedness with human.

Medicinal plants are being looked upon one again for the treatment of diabetes. Many conventional drugs have been derived from prototypic molecules in medicinal plants. Metformin exemplifies an efficacious oral glucose –lowering agent. Its development was based on the use of *Galiga officinalis* to treat diabetes. *Galiga officinalis* is rich in guanidine, the hypoglycemic component. Because guanidine is too toxic for clinical use, the alkyl biguanides synthalin A and Synthalin B were introduced as oral antidiabetic agents in Europe in the 1920s but these were discontinued after insulin became more widely available. However, experience with guanidine and biguanides prompted the development of metformin. Upto now, over 400 traditional plant treatments for diabetes have been reported, although only a small number of these have received scientific and medical evaluation to assess their efficacy. The hypoglycemic effect of some herbal extracts has been confirmed in human and animal models of type 2 diabetes. The World Health Organization (WHO) Expert Committee on diabetes has recommended that medicinal herbs be further investigated. Based on this recommendation and need for conducting clinical research in herbal drugs, developing simple bioassays for biological standardization, pharmacological and toxicological evaluation, current study is performed to evaluate the antidiabetic potential of *Tinospora cordifolia*.

Tinospora cordifolia (Guduchi) is an evergreen perennial climber. This deciduous and dioecious plant belong to the family Menispermaceae which consists of about 70 genera and 450 species that are found in tropical lowland regions. They are generally climbing or twining, rarely shrubs. This family is rich source of alkaloid and terpenes [13]. In Hindi the plant is commonly called Giloe [14] which is Hindu mythological term that refers to the heavenly elixir that has saved celestial beings from old age and kept them eternally young. In Ayurveda, it is designated as Rasayana drug

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recommended to enhance general body resistance, promote longevity and as anti-stress and adaptogen [15,16].

II. MATERIAL AND METHODS

The plant material *Tinospora cordifolia* was collected from hygienic places from in and around the Nagpur city. The plant material was washed with water in order to make it free of dirt and other impurities and was shade dried. Shade dried whole Plant material was grind with mortar and pastel into the fine powder and alcoholic and aqueous extract of *Tinospora cordifolia* was prepared according to the standard procedure.

Healthy albino rats (9 months old) of both the sexes, weighing 150-190gm were used for the experiment. Animals were free to access drinking water and food. Animals were cared for and used in accordance with the Institutional Animal Ethics Committee (IAEC), P.G.T. Department of Zoology, RTM Nagpur University, Nagpur (Registration No.-478/01/a/CPCSEA).

Four batches of experiment was carried out for standardization of dose. Each batch includes three groups of rats (n=6). Group I included young rats of less than 6 month old, group II included the rat of age group 6-12 month old and group III was included rats of more than 12 month age group. Diabetes was induced in 16hrs fasted albino rats with single intraperitoneal dose of alloxan monohydrate. Alloxan injection was prepared in 0.9% normal saline. Rats with fasting blood glucose more than 220 mg/dl was considered for study. The batch I was injected with 120mg/kg bw, batch II with 140mg/kg, bw, batch III with 160mg/kg bw and batch IV with 180 mg/kg bw. During dose standardization study it was found that 180mg/kg intraperitoneal dose of alloxan monohydrate was suitable for diabetes induction with the 6-12 month old rats.

For experimental induction of diabetes alloxan monohydrate (A74/3 Sigma Aldrich was used).

For this study the animals were divided into three groups (n=6),

Group-I (NC): Kept as normal control, the animals of this group was free to access drinking water and food; they were neither injected by alloxan nor feed on plant extract.

Group- II (DC): These group of animals were injected with alloxan monohydrate (180 mg/kg bw) and kept as diabetic control. They were not feed on extract.

Group-III (DC+TCE): This group was injected with alloxan monohydrate (180mg/kg bw) and from day 2 to 30 half an hour prior to feeding, orally administrated with TCE (20ml/kg bw) twice a day.

a) Estimation of liver glycogen

Liver sample was dissected out from 16 hrs fasted rat and digested in hot 30% KOH. Liver glycogen was precipitated with alcohol and the precipitate was dissolved in 10% TCA. The sample was processed for centrifugation to sediment the proteins, after centrifugation supernatant was precipitated once again with alcohol. After suitable dilution of the sediment with water, estimation of liver glycogen was carried out with anthrone reagent [17].

III. RESULTS AND DISCUSSION

In the present study we observed the very significantly ($P < 0.005$) increased liver glycogen in diabetic control group compared to the normal. This abnormally increased glycogen content is reversed by the *Tinospora cordifolia* whole plant extract. Although almost all the liver glycogen of the normal rats disappeared after only 24 hours of fasting, rather large amount of liver glycogen was reserved in alloxan diabetic rats when they were fasted for 16 hours before testing. The amount of the remaining liver glycogen was affected by the blood glucose level. Several factors which suppress liver glycogen in normal control by 16 hrs fasting may be pointed out; first one of them is the decrease of the blood glucose which may be followed by marked suppression of glucokinase. Since, glucokinase is the key enzyme catalysing glucose phosphorylation in liver. Impairment of glucokinase activity suggests the impaired oxidation of glucose via glycolysis causes its accumulation resulting in hyperglycemia. Secondly, the suppression of glycogen synthetase together with the stimulation of phosphorylase may be related to the suppression of liver glycogen. The high liver glycogen level in diabetic rats may be due to either increase in gluconeogenesis or hyperglycemia due to 16 hr fasting before testing. In *Tinospora Cordifolia* whole plant extract treated group the significant ($P < 0.005$) reversion of the liver glycogen towards the normal may be due to it's activating effect on the glucokinase and glycogen synthetase.

Table 1 : Liver glycogen Estimation.

Groups	Liver glycogen (mg/g wet wt)
NC	5.2±0.87
DC	11.7±1.2 ^a
DC+TCE	6.3±0.45 ^a

NC- Normal control

DC- Diabetic control

TCE- *Tinospora cordifolia* extract

(Values are expressed as Mean ± SEM (n=6), paired t-test was performed to compared between groups. ^a $P < 0.005$ when DC compared with NC and DC+TCE compared with DC).

After absorption into a cell, glucose can be used immediately for release of energy to the cell, or it can be stored in the form of glycogen, which is a large

polymer of glucose. All cells of the body are capable of storing at least some glycogen, but certain cells can store large amounts, especially liver cells, which can be stored upto 5 to 8 per cent of their weight as glycogen, and muscle cells, which can be stored up to 1 to 3 per cent glycogen.

The importance of the liver in the regulation of carbohydrate metabolism is recognized by its ability to store carbohydrates in the form of glycogen (glycogenesis) and to release them in the form of glucose (glycogenolysis) when needed. These processes are regulated by 2 key enzymes: glycogen synthase and glycogen phosphorylase.

According to Dewalkar *et al.*, [18] the fasted liver glycogen concentration was found very significantly higher ($P < 0.001$) in diabetic control (DC) than normal control (NC). This may be due to the increase rate of glycogen synthase enzyme in alloxan treated group, which in turn account for accumulation of glycogen in diabetic rat liver. Fresh etiolated wheat grass and fruit squash of *Lagerstroemia speciosa* treated groups show statistically significant difference in glycogen concentration like NC. This reversal of glycogen concentration to normal indicates their preventive effect on alloxan induced increase rate of glycogen synthase. They found fresh etiolated wheat grass posses significant ($P < 0.05$) effect on lowering of glycogen concentration than fruit squash of *Lagerstroemia speciosa*.

According to Daisy and Rajathi, [19] the aqueous extracts of *Clitoria ternatea* leaves and flowers decrease in glycogen content of liver and skeletal muscle in diabetic rats is probably due to lack of insulin in the diabetic state. Prevention of glycogen depletion in the liver and muscles, following the administration of the extracts, could therefore have been achieved by stimulation of insulin release. Administration of *Clitoria ternatea* leaves and flowers to the diabetic animals increased the activity of glucokinase in liver. The extract-induced decrease in the concentration of blood glucose in alloxan-treated rats may be the result of improved glucose uptake. Similar observations have been made by Singh *et al.* and Shanmugasundaram *et al.* in respect of the extracts of *Catharanthus roseus*, and *Gymnema sylvestre* respectively [20, 21]. The activity of the gluconeogenic enzyme, glucose-6- phosphatase, is usually enhanced during diabetes. following extract administration, blood glucose level falls; while liver glycogen content rose. This may be due to the mobilization of blood glucose into the liver glycogen reserve [22, 23, 24].

IV. SUMMARY AND CONCLUSION

Beside blood glucose, quantitative analysis of liver glycogen content was carried out to interpret the relation between blood glucose and liver glycogen here we observed the significantly increased liver glycogen in

diabetic control group compared to the normal. This abnormally increased glycogen content is reversed by the *Tinospora cordifolia* whole plant extract. Although almost all the liver glycogen of the normal rats disappeared after only 24 hours of fasting, rather large amount of liver glycogen was reserved in alloxan diabetic rats when they were fasted for 16 hours before testing. The amount of the remaining liver glycogen was affected by the blood glucose level. Several factors which suppress liver glycogen in normal control by 16 hrs fasting may be pointed out; first one of them is the decrease of the blood glucose which may be followed by marked suppression of glucokinase. Since, glucokinase is the key enzyme catalysing glucose phosphorylation in liver. Impairment of glucokinase activity suggests the impaired oxidation of glucose via glycolysis causes its accumulation resulting in hyperglycemia. Secondly, the supression of glycogen synthase together with the stimulation of phosphorylase may be related to the suppression of liver glycogen. The high liver glycogen level in diabetic rats may be due to either increase in gluconeogenesis or hyperglycemia due to 16 hr fasting before testing. In *Tinospora cordifolia* whole plant extract treated group the reversion of the liver glycogen towards the normal may be due to its activating effect on the glucokinase and glycogen synthetase.

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Parkia Biglobosa Jacq (dawa-dawa): The Threatened Giant of the Guinea Savanna of Nigeria (The Cross River State Situation)

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Abstract- *Parkia biglobosa* (Jacq) is a perennial tree of considerable multipurpose importance. In Nigeria, over 60 million people depend on it for food, fuel, wood work and income. This priority tree with enormous economic and social values to the local people of Cross River State (North) is rapidly declining due to rapid human population growth, livestock population pressures, increasing land fragmentation, over exploitation for seed (food) and a high demand for wood fuel especially charcoal. In view of this, a survey was carried out in the Guinea Savanna of Cross River State (North) to determine the level of abundance (A) or rarity (R) of the specie. In each site, ten 20m × 20m randomly selected points were laid along four 1km line transect and accordingly assessed. Seventeen economic important trees were assessed and counted as present with particular emphasis on *P. biglobosa* (taking into consideration the age of the plant). Results showed that the tree was rare (R) and facing extinction with low densities of 2.1/ha for (Ogoja L.G.A), 1.6/ha (Bekwarra), 2/ha (Obudu), 1.7/ha (Obanliku) and 1.5/ha for (Yala).

Keywords: *parkia biglobosa, threatened specie, logistic model, guinea savanna, cross river state.*

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Parkia Biglobosa Jacq (dawa-dawa): The Threatened Giant of the Guinea Savanna of Nigeria (The Cross River State Situation)

Udo, S. E. ^α, Akwaji, P. I. ^σ, Markson, A. A. ^ρ, Umana, E. J. ^ω, Okey, E. N. [¥] & Asuquo, M. [§]

Abstract- *Parkia biglobosa* (Jacq) is a perennial tree of considerable multipurpose importance. In Nigeria, over 60 million people depend on it for food, fuel, wood work and income. This priority tree with enormous economic and social values to the local people of Cross River State (North) is rapidly declining due to rapid human population growth, livestock population pressures, increasing land fragmentation, over exploitation for seed (food) and a high demand for wood fuel especially charcoal. In view of this, a survey was carried out in the Guinea Savanna of Cross River State (North) to determine the level of abundance (A) or rarity (R) of the specie. In each site, ten 20m × 20m randomly selected points were laid along four 1km line transect and accordingly assessed. Seventeen economic important trees were assessed and counted as present with particular emphasis on *P. biglobosa* (taking into consideration the age of the plant). Results showed that the tree was rare (R) and facing extinction with low densities of 2.1/ha for (Ogoja L.G.A), 1.6/ha (Bekwarra), 2/ha (Obudu), 1.7/ha (Obanliku) and 1.5/ha for (Yala). On average, there were 48.34% old trees than juveniles and 20.82% more young trees than juveniles. The Logistic model of population dynamics (with modifications), was used to prove what will be the population size of the specie after ten years (10yrs) at equilibrium critical points of (P=0, P=N and P=M). The study showed that high demand for food, bush burning, farming activities and urbanization within these areas have brought about decimation and near extinction of this valuable tree specie. Domestication and plantation establishment of the specie to enhance its productivity and to curb the threat of extinction is strongly recommended.

Keywords: *parkia biglobosa*, threatened specie, logistic model, guinea savanna, cross river state.

I. INTRODUCTION

Parkia biglobosa Jacq is a perennial tree of the genus *Parkia* in the family Fabaceae. In Nigeria, its fruits are fermented into a condiment called "dawa-dawa" (Salim *et al.*, 2002).

Parkia biglobosa is an important economic tree legume of considerable multipurpose importance. The tree attracts bees and is a popular tree among bee

keepers. Whole pods are eaten by domestic stock including cattle. The young seedlings are nutritious and heavily browsed by livestock. The seed of the African locust bean when boiled and fermented is known as "dawa-dawa" in Hausa language in Nigeria, a black smelling tasty seasoning, rich in lipid 29%, protein 35%, carbohydrate 16%, good source of protein, fat, calcium for rural dweller. The bark is used as mouth wash, vapour inhalant for toothache, or for ear complaints. It is macerated in baths for leprosy and used for bronchitis, pneumonia, skin infections, sores, ulcers, and washes for fever, malaria, diarrhea, and sterility. Roots are used in a lotion for sore eyes. Pulp is supposedly a water purifier but possibly just sweetens and disguises taste of foul water. The sweet yellow pulp contains 60% sugar when ripe and the seeds contain vitamins as well as minerals (Sacande and Clethero, 2007). The fruit pods are used to produce an insecticide powder for treating crops. *Parkia* tree is used as timber for making pestles, mortars, bows, hoe handles and seats (Joshi and Joshi, 2009).

Parkia biglobosa (dawa-dawa) is indigenous specie that is economically and socially important for local people in Cross River State (North). The tree serves as source of wood, food, fodder, and medicine for local people. They also provide ecological services including soil fertility and microclimate amelioration. In addition to direct domestic use of the tree products, they are a source of cash for local people (Odebiyi *et al.*, 2004).

According to Odebiyi *et al.*, (2004) the population of this priority tree is rapidly declining due to rapid human population growth, livestock-population pressures, increasing land fragmentation, over exploitation for seed and a high demand for wood fuel especially charcoal. Also, fallowing of farmlands a practice on which the regeneration of this tree relied is no longer being followed. As a result, existing trees are aging.

From the forgoing, the main aim of this research, therefore, was to investigate the population size of *P. biglobosa* in the Guinea savanna of Cross River State (North) as well as build a modified Logistic Model of population dynamics of *Parkia biglobosa* in the next ten years in the face of continuous exploitation without replacement.

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II. MATERIALS AND METHODS

Cross River State has a total land mass of 20,156km²(7,782squ mi). Of the total land mass, 21,600 hectares is covered by savanna woodland and mostly found in Cross River (North). The state is located in South Eastern Nigeria Latitudes 5°45'N' and Longitude 8°30'E. It has a humid tropical climate of (1300-3000mm) rainfall and a mean annual temperature of 30°C.

The study was conducted in January to July, 2013. The study was conducted in the cultivated and fallow land uses in the Guinea Savanna of Cross River (North) making a total of 5 Local Government Areas (5 Locations) Ogoja, Yala, Bekwarra, Obudu, and Obanliku and 3 villages in each of the Local Government Areas (15 study sites) Ishindede, Ndok, Idum (Ogoja), Abuoichiche, Afrike, and Gakem (Bekwarra), Okpoma, Ugaga, and Ijegu (Yala), Ukpada, Alege, and Betukwel (Obudu), Sankwala, Busi, and Udeshi (Obanliku). The

summary of the agro climatic characteristics of the study sites are given in (Table 1). The study was conducted using line transects. In each site, four 1km line transects were laid, each separated by a distance of 200m. The transects ran through the cultivated and fallow land uses of the Guinea Savanna of Cross River (North). Along each transect, ten 20m × 20m sample plots were laid at randomly selected points with the help of States Forestry Department Officers. In all, a total area of 40,000m² was assessed along each transect which gives a gross total area of about 20 hectares. The numbers of stands of *P. biglobosa* and other tree species producing various economically valued products present on the marked plots were counted and recorded (taking into consideration the age of the *P. biglobosa* trees). Data collated was used to determine each species percentage frequency (%), Density, and Relative Abundance using the method of (Sharma, 2009). The (%) frequency for the species in the marked Area was calculated using the formula:

$$\text{Frequency (\%)} = \frac{\text{No. of sampling units in which a particular species occurred}}{\text{Total no. of sampling units studied}} \times 100$$

After determining the percentage frequency of each species, various species were distributed among Sharma's (2009) five frequency classes depending upon their frequency values as presented in (Table 2). A plant species was classified as abundant (A) or rare (R) based on its frequency class in all the assessed segments. A plant species found in 41% and above of all the assessed segments was considered abundant

(A) while a species present in less than 41% of all the assessed segments was regarded as rare (R).

However, since frequency does not give correct idea of the distribution of any species unless it is correlated with other characters such as density and abundance. The percentage density of the population was also determined using the formula:

$$\text{Density} = \frac{\text{Total no. of individuals of the species in all the sampling units}}{\text{Total no. of sampling units studied}}$$

While the relative abundance of the species was calculated thus:

$$\text{Abundance} = \frac{\text{Total no. of individuals of the species in all the sampling units}'}{\text{Total no. of sampling units in which the species has occurred}}$$

To predict what will be the population size of *P. biglobosa* after ten years. The Logistic model of population dynamics also called the (Verhulst-pearl) model was used. Modeling was carried out in this study using the methods of (Volterra, 1926, Yodzis, 1978, Law and Blackford, 1992, Cohen, 1995, Mckelvey, 1996, Alexei, 1996, Vandameer, 2010, Arditi and Ginzberg, 2012, Knaust, 2013). The model states that the rate of population increase may be limited i.e., it may depend on population density:

$$r = \frac{r_0 (1 - N)}{K}$$

At low densities ($N < K$), the population growth rate is maximal and equals to r_0 . Parameter r_0 can be interpreted as population growth rate in the absence of intra-specific competition. Population growth rate declines with population numbers N , and reaches 0 when $N=K$. Parameter K is the upper limit of population

growth and it is called carrying capacity. It is usually interpreted as the amount of resources expressed in the number of organisms that can be supported by these resources. If population numbers exceed K , then population growth rate becomes negative and population numbers decline. The dynamics of the population is described by the differential equation:

$$\frac{dN}{dt} = rN \left(1 - \frac{N}{K}\right)$$

Where $N(t)$ represents number of individuals at time t , r the intrinsic growth rate and K is the carrying capacity, or the maximum number of individuals that the environment can support. Logistic model has two equilibriums $N=0$ and $N=K$. The first equilibrium is unstable because any small deviation from this equilibrium will lead to population growth. The second equilibrium is stable because after small disturbances the population returns to this equilibrium state. Logistic

model combines two ecological processes: reproduction and competition. Both processes depend on population numbers or density (Hogeweg and Hesper, 1978, Alexei, 1996, Alder, 1997, May, 2004, Campbell and Reece, 2008).

a) What will be the population size of *P. biglobosa* after n years (or generations)?

Using the differential equation of the Logistic model which is:

$$\frac{dP}{dt} = kP \left(1 - \frac{P}{M}\right)$$

In order to solve this equation, a non-linear equation which is separable was used. The constant solutions were $N=0$ and $N=K$. The non-constant solutions was obtained by separating the variables

$$\frac{dP}{P \left(1 - \frac{P}{M}\right)} = k dt$$

And integration

$$\int \frac{dP}{P \left(1 - \frac{P}{M}\right)} = \int k dt .$$

The partial fraction technique gives

$$\int \frac{dP}{P \left(1 - \frac{P}{M}\right)} = \int \left(\frac{1}{P} + \frac{1/M}{1 - P/M} \right) dP$$

This gives

$$\ln |P| - \ln \left| 1 - \frac{P}{M} \right| = kt + c .$$

Easy algebraic manipulation gives

$$\frac{P}{1 - P/M} = C e^{kt} ,$$

Where C is a constant solving for P , we get

$$P = \frac{MC e^{kt}}{M + C e^{kt}} .$$

If we consider the initial condition $P(0) = P_0$ (assuming that P_0 is not equal to both 0 or M), we get

$$C = \frac{P_0 M}{M - P_0}$$

Which when once substituted into the expression for $P(t)$ and simplified we find

$$P(t) = \frac{MP_0}{P_0 + (M - P_0)e^{-kt}} .$$

It is easy to see that

$$\lim_{t \rightarrow +\infty} P(t) = M .$$

However, this is still not acceptable because this model does not tell us when a population is facing extinction since it never implies that. It can be concluded that extinction never occurs under the logistic model. Even starting with a small population it will always tend to the carrying capacity M . Hence, it became necessary to modify the model using the method of (Mckelvey, 1996) which increased the models realism and usefulness. This was the addition of the ecological idea of a minimum viable population. The idea here is that a population of any species has a minimum level at which the population can thrive. If the population drops below this minimum level, various environmental and genetic factors lead to the elimination of the population. The relevant factors might include inability of trees to reproduce, loss of genetic diversity and increased vulnerability to short and long term environmental changes and disease events. The logistic model was modified to account for the existence of a minimum viable population by using the difference equation:

$$\frac{dP}{dt} = kP \left(1 - \frac{P}{N}\right) \left(\frac{P}{M} - 1\right)$$

Where N is the carrying capacity and indicates when the population is too big and the sparsity parameter M indicates when the population is too small (minimum viable population).

Since *P. biglobosa* is a perennial tree that is confined to a particular habitat (mostly savanna):

- If the population is large, their rate of growth decreases or even becomes negative as a result of intra-specific competition.
- If the population is too small (as with the case in this study), old trees run the risk of not being able to reproduce and as such the rate of growth is also negative. Based on these observations, the modified logistic model was used to find the equilibrium (or critical) points of the *P. biglobosa* population for the next ten years.

b) Application of the Modified Logistic Model of Population Dynamics

$$\frac{dp}{dt} = kp \left(1 - \frac{p}{N}\right) \left(\frac{p}{M} - 1\right).$$

$$\Leftrightarrow \frac{DP}{KP \left(1 - \frac{P}{N}\right) \left(\frac{P}{M} - 1\right)}$$

Integrated both side to get

$$\int \frac{dp}{kp \left(1 - \frac{p}{N}\right) \left(\frac{p}{M} - 1\right)} = \int dt + C_0 \quad *$$

Decompose $\frac{1}{kp(1-\frac{p}{N})(\frac{P}{M}-1)}$. into partial fractions

$$\frac{1}{kp(1-\frac{p}{N})(\frac{P}{M}-1)} = \frac{A}{KP} + \frac{B}{(1-\frac{P}{N})} + \frac{C}{(\frac{P}{M}-1)}$$

$$= \frac{[A(1-\frac{P}{N}) + BKP](\frac{P}{M}-1) + CKP(1-\frac{P}{N})}{KP(1-\frac{P}{N})(\frac{P}{M}-1)}$$

$$\frac{1}{KP(1-\frac{P}{N})(\frac{P}{M}-1)} = \frac{(\frac{A}{M} - BK + \frac{A}{N} + CK)P + (-\frac{A}{NM} + \frac{BK}{M} - \frac{CK}{N})P^2 - A}{KP(1-\frac{P}{N})(\frac{P}{M}-1)}$$

A, B, C, are the unknown means

$$\begin{cases} A = -1 \\ -\frac{A}{M} - BK + \frac{A}{N} + CK = 0 \\ -\frac{A}{NM} + \frac{BK}{M} - \frac{CK}{N} = 0 \end{cases} \Rightarrow \begin{cases} A = -1 \\ (-K)B + KC = \frac{1}{M} + \frac{1}{N} \\ (\frac{K}{M})B + (-\frac{K}{N})C = -\frac{1}{NM} \end{cases}$$

$$D = \begin{vmatrix} -K & K \\ \frac{K}{M} & -\frac{K}{N} \end{vmatrix} = \frac{K^2}{N} - \frac{K^2}{M} \text{ Then}$$

$$= \left| \frac{1}{M} + \frac{1}{N} K \right|$$

$$B = \frac{-\frac{1}{NM} - \frac{-K}{N}}{D} = \frac{\frac{K}{N}(\frac{1}{M} + \frac{1}{N}) + \frac{K}{NM}}{D}$$

$$= \frac{\frac{-K}{NM} - \frac{K}{N^2} + \frac{K}{NM}}{D}$$

$$= \frac{-K}{DN^2}$$

$$B = \frac{-K}{KN(\frac{N}{M}-1)} = \frac{K}{K^2(-N+\frac{N^2}{M})} = \frac{1}{KN(\frac{N}{M}-1)}$$

$$B = \frac{I}{KN(\frac{N}{M} - 1)}$$

$$C = \frac{\frac{-K}{\frac{K}{M} - \frac{I}{NM}} \frac{I}{M} + \frac{I}{M}}{\frac{NM - \frac{K}{M}(\frac{I}{M} + \frac{I}{N})}{D}} = \frac{K}{D}$$

$$= \frac{\frac{K}{NM} - \frac{K}{M^2} - \frac{K}{NM}}{D} = \frac{-K}{DM^2}$$

$$= \frac{K}{M^2(\frac{I}{N} - \frac{I}{M})K^2} = \frac{I}{K(M - \frac{M^2}{N})}$$

$$C = \frac{I}{KM(\frac{I}{N} - \frac{M}{N})} \text{ With A, B, C, obtained}$$

The left hand side of (*) is

$$\begin{aligned} & \int \frac{dp}{kp(1 - \frac{p}{N})(\frac{P}{M} - 1)} \\ &= \frac{1}{k} \left[-\int \frac{dp}{p} + \frac{1}{N(\frac{N}{M} - 1)} \int \frac{dp}{(1 - \frac{p}{N})} + \frac{1}{M(1 - \frac{M}{N})} \int \frac{dp}{(\frac{p}{m} - 1)} \right] \\ &= \frac{1}{k} \left[Lnp + \frac{N \ln}{N(\frac{N}{M} - 1)} (1 - \frac{P}{N}) + \frac{M \ln}{m(1 - \frac{m}{N})} (\frac{P}{M} - 1) \right] \\ &= Lnp - \frac{1}{k(1 - \frac{p}{N})} - \frac{1}{(\frac{N}{M} - 1)(\frac{P}{M} - 1)} - \frac{1}{(1 - \frac{M}{N})} \end{aligned}$$

The right hand side of (*) is

$$\int dt + C_o = t + C_o$$

Hence the differential equation has the general solution

$$\ln p - \frac{1}{k(1-\frac{p}{N})} - \frac{1}{(1-\frac{N}{M})(\frac{P}{M}-1)} - \frac{1}{(1-\frac{M}{N})} = t - C_o$$

Take the exponential of both sides to finally get:

$$\frac{(1-\frac{p}{N})(1-\frac{N}{M})(\frac{P}{M}-1)\frac{1}{(1-\frac{M}{N})}}{P\frac{1}{N}} = C, e^t \text{ where } C, = e^{C_o}$$

Since the model is described by the differential equation:

$$\frac{dP}{dt} = kP \left(1 - \frac{P}{N}\right) \left(\frac{P}{M} - 1\right)$$

Where

P is the population

t is time

N is the carrying capacity which indicates when the population is too big (max).

In this problem N = 2.1

M the sparsity parameter indicates when the population is too small (min).

In this problem M = 1.5

Tentative solution

Assuming the survey took place in 2006 and found that the total population was $\alpha = 2.1 + 1.8 + 2.0 + 1.7 + 1.5 = 9.1$

So that at t = 0; P = 9.1

Assuming also that at the same time N = 2.1 and M = 1.5

Let t be the extinction period which is 10 years. This means that at time t, P should be very close to zero (0).

Mathematically, this is written as:

$$\begin{aligned} \int_{\alpha}^0 \frac{dp}{kp(1-\frac{p}{N})(\frac{P}{M}-1)} &= \int_0^T dt \\ \Leftrightarrow \int_a^0 \frac{dp}{kp(1-\frac{1}{N})(\frac{P}{M}-1)} &= \int_0^{10} dt \\ \Leftrightarrow \frac{(1-\frac{p}{N})(1-\frac{N}{M})(\frac{P}{M}-1)(\frac{1}{1-\frac{M}{N}}) \int_a^0}{P\frac{1}{K}} &= 10 \end{aligned}$$

With N = 2.1; M = 1.5, we have:

$$\left[\frac{(1-\frac{0}{2.1}) \frac{1}{(1-\frac{2.1}{1.5})(\frac{0}{1.5}-1)(1-\frac{1.5}{2.1})}}{0\frac{1}{k}} \right] - \left[\frac{(1-\frac{9.1}{2.1}) \frac{1}{(1-\frac{2.1}{1.5})(\frac{9.1}{1.5}-1)(1-\frac{1.5}{2.1})}}{(9.1)\frac{1}{k}} \right] = 10$$

Note: The 0 in the denominator means the population is close to zero and not exactly equal to zero.

III. CONCLUSION

Since there is a balance between the left and right hand side of the above equation, then it is proved that the minimum viable population *M* of *P. biglobosa* decreased and will continue to decrease over time (*t*)

unless viable intervention measures are taken. Based on these findings, It can be concluded that populations less than *M* (1.5) will decrease to oblivion and will continue to in the face of continuous exploitation without replacement.

Table 1 : Characteristics of the Agro-ecological zone surveyed for *P. biglobosa* in Cross River (North) woodland savanna

Ecozone	StudyLocations	Latitude(N)	Longitude(E)	Elevation(ft)	Annual rainfall (mm)	Rainfall pattern
CROSS RIVER STATE Guinea Savanna	Ishindede	6.50°	8.61°	403	1250-1300	Unimodal
	Ndok	6.59°	8.79°	452	1250-1300	Unimodal
	Idum	6.53°	8.90°	597	1250-1300	Unimodal
	Abuochiche	6.69°	8.97°	485	1250-1300	Unimodal
	Afrike	6.61°	8.87°	436	1250-1300	Unimodal
	Gakem	6.77°	8.99°	498	1250-1300	Unimodal
	Okpoma	6.59°	8.63°	383	1250-1300	Unimodal
	Ugaga	6.67°	8.73°	413	1250-1300	Unimodal
	Ijegu	6.74°	8.67°	400	1250-1300	Unimodal
	Ukpada	6.71°	8.90°	439	1250-1300	Unimodal
	Alege	6.52°	8.43°	393	1250-1300	Unimodal
	Betukwel	6.61°	9.12°	646	1250-1300	Unimodal
	Sankwala	6.55°	9.22°	1266	1250-1300	Unimodal
	Busi	6.55°	9.27°	2057	1250-1300	Unimodal
	Udeshi	6.55°	9.12°	1571	1250-1300	Unimodal

Table 2 : Sharma's (2009) five frequency classes

Frequency (%)	Frequency class
0-20	A(R)
21-40	B(R)
41-60	C(A)
61-80	D(A)
81-100	E(A)

Note: (A) Abundant, (R) Rare

IV. RESULTS AND DISCUSSION

a) Species Distribution and Density

Cross River state (North) Guinea savanna was assessed for the Abundance (A) or Rarity (R) of *Parkia biglobosa*. In all, seventeen tree species producing various economically important products were encountered. Two of the 17 tree species were observed to be rare (R), while 15 species were abundant (A) as presented in (Table 3). The rarity of *Adansonia digitata* is quite understood, since it is found further up north of the country in abundance (A) mostly in the Sudan savanna. However, *P. biglobosa* which is very common in the Guinea savanna belt of Nigeria was rare (R) in all the assessed areas with low densities of 2.1/ha for Ogoja L.G.A, 1.6/ha (Bekwarra), 2/ha (Obudu), 1.7/ha (Obanliku), and 1.5/ha for Yala respectively (Tables 4, 5, 6, 7 & 8), While *Vitellaria paradoxia* and *Elaeis guinensis*

had the highest densities of 3.2/ha for *V. paradoxia*, 10/ha for *E. guinensis* (Ogoja L.G.A), 4/ha and 4.8/ha (Bekwarra), 3.8/ha and 5.9/ha (Obudu), 3.6/ha and 5.2/ha (Obanliku) and 5.3/ha (*V. paradoxia*), 5.7/ha (*E. guinensis*) for Yala L.G.A respectively. On average, there were more old trees (of *P. biglobosa*) than juveniles, and more young trees than juveniles.

The population of *P. biglobosa*, a priority tree with enormous economic and social values to the people of Cross River State (North) was observed to be rare and is rapidly declining. This is due to rapid human population growth, livestock population pressures, increasing land fragmentation, over exploitation for seed and a high demand for wood fuel especially charcoal. In this study, the overall densities for *P. biglobosa* in all the surveyed areas were quite low and facing threat of extinction. The population densities were 2.1stands/ha (Ogoja), 1.6stands/ha (Bekwarra), 2stands/ha (Obudu), 1.7stands/ha (Obanliku) and 1.5stands/ha (Yala) in the cultivated and fallow land uses. However, these findings disagrees with that of Odebiyi *et al* (2004) who reported higher densities 8.2stands/ha, 6.6stands/ha and 5.2 stands/ha respectively in the cultivated and fallow land uses of the Northern Guinea Savanna of Kwara State, Nigeria. In this study we observed that, on average, there were more old trees than juveniles and more young trees than juveniles. Saplings of *P. biglobosa*

were absent in all the surveyed areas, these findings agrees with that of (odebiyi *et al* (2004), Joshi and Joshi (2009). The scarcity of the saplings may be as a result of grazing by livestock, uncontrolled incessant bush burning and over exploitation for seeds which may have hampered their regeneration and aggravated the threat of extinction that the specie is subjected to. More

worrisome is the fact that the specie is neither domesticated at present nor planted in organized forestry plantations, and large trees are being over exploited for timber, fuel wood, and production of charcoal and removal of stands to give way for construction works (urbanization).

Table 3 : List of different tree species assessed in the Guinea savanna of Cross River State (North) and their Ecological Status.

S/N	Name of the species	Frequency (%)	Frequency class	Ecological Status
1.	<i>Parkia biglobosa</i>	30	B	R
2.	<i>Elaeis guinensis</i>	60	C	A
3.	<i>Hyphaena thebaica</i>	50	C	A
4.	<i>Vitellaria paradoxa</i>	60	C	A
5.	<i>Isoberlina doka</i>	70	D	A
6.	<i>Prosopis africana</i>	60	C	A
7.	<i>Adansonia digitata</i>	20	A	R
8.	<i>Ceiba pentandra</i>	60	C	A
9.	<i>Anarcadium occidentale</i>	50	C	A
10.	<i>Gmelina arborea</i>	60	C	A
11.	<i>Magnifera indica</i>	70	D	A
12.	<i>Azadirachta indica</i>	50	C	A
13.	<i>Raffia ruffia</i>	60	C	A
14.	<i>Acacia nilotica</i>	70	D	A
15.	<i>Moringa oleifera</i>	50	C	A
16.	<i>Milicia excelsa</i>	70	D	A
17.	<i>Cocos nucifera</i>	80	D	A

Note: A (abundant), R (rare).

Table 4 : List of different tree species and other data recorded by Transect method in Ogoja L.G.A.

S/N	Name of the species	Segments number 1 2 3 4 5 6 7 8 9 10	Total no. Of individuals Of species	Total no. of segments the species occurred	Total no. of segments studied	Frequency (%)	Frequency class	Density	Abundance
1.	<i>Parkia biglobosa</i>	-- 3 - 5 -- 7 --	15	3	10	30	B	1.5	5
2.	<i>Elaeis guinensis</i>	4 3 - 12 - 9 - 12 - 20	60	6	10	60	C	6	10
3.	<i>Hyphaena thebaica</i>	5 3- 20 35 - 20 ---	83	5	10	50	C	8.3	16.6
4.	<i>Vitellaria paradoxa</i>	13 - - 5 3- - 2 4 5	32	6	10	60	C	3.2	5.3
5.	<i>Isoberlina doka</i>	3 21 4 - 6 8 -- 2 9	53	7	10	70	D	5.3	7.6
6.	<i>Prosopis africana</i>	-- 10 5 20 - 3 - 6 1	45	6	10	60	C	4.5	7.5
7.	<i>Adansonia digitata</i>	5 ---- 1 --- 2	8	2	10	20	A	0.8	4
8.	<i>Ceiba pentandra</i>	- 9 6 9 2 - - 13 - 3	42	6	10	60	C	4.2	7
9.	<i>Anarcadium occidentale</i>	2 10 3 7 - - 15 - -	37	5	10	50	C	3.7	7.4
10.	<i>Gmelina arborea</i>	4 - - 19 - 2 8 - 6 20	59	6	10	60	C	5.9	9.8
11.	<i>Magnifera indica</i>	10 - 3 - 7 - 15 - 5 9 8	57	7	10	70	D	5.7	8.1
12.	<i>Azadirachta indica</i>	-- 6 13 - 2 - 10 - 2	33	5	10	50	C	3.3	6.6
13.	<i>Raffia ruffia</i>	- 3 8 2 - 5 20 - 1 -	39	6	10	60	C	3.9	6.5
14.	<i>Acacia nilotica</i>	9 2 - - 5 8 1 12 - 3	40	7	10	70	D	4	5.7
15.	<i>Moringa oleifera</i>	- - - 10 4 7 2 8 - -	31	5	10	50	C	3.1	6.2
16.	<i>Milicia excels</i>	2 8 5 19 1 - - - 6 9	50	7	10	70	D	5	7.1
17.	<i>Cocos nucifera</i>	3 12 6 - 10 - 9 2 4 7	43	8	10	80	D	4.3	5.3

Table 5 : List of different tree species and other data recorded by Transect method in Bekwarra L.G.A.

S/N	Name of the species	Segments number 1 2 3 4 5 6 7 8 9 10	Total no. Of individuals Of species	Total no. of segments the species occurred	Total no. of segments studied	Frequency (%)	Frequency class	Density	Abundance
1.	<i>Parkia biglobosa</i>	8 - - 3 - - 6 - -	17	3	10	30	B	1.7	5.6
2.	<i>Elaeis guinensis</i>	9 10 6 8 5 3 - - 7	48	7	10	70	D	4.8	6.8
3.	<i>Hyphaena thebaica</i>	11 4 7 9 - - 20 5 - -	56	6	10	60	C	5.6	9.3
4.	<i>Vitellaria paradoxa</i>	6 8 5 - 9 - 9 - 2 1	40	6	10	60	C	4	6.6
5.	<i>Isobertina doka</i>	- 4 - 9 12 - 6 3 - 7	41	6	10	60	C	4.1	6.8
6.	<i>Prosopis africana</i>	2 9 7 5 8 - - 1 3 -	35	7	10	70	D	3.5	5
7.	<i>Adansonia digitata</i>	- - - - 2 - - 1	3	2	10	20	A	0.3	1.5
8.	<i>Ceiba pentandra</i>	1 4 - - 7 8 10 2 - -	32	6	10	60	C	3.2	5.3
9.	<i>Anarcadium occidentale</i>	8 12 6 9 3 2 4 - - 6	50	8	10	80	D	5	6.2
10.	<i>Gmelina arborea</i>	- 1 4 20 - 9 2 - -	36	5	10	50	C	3.6	7.2
11.	<i>Magnifera indica</i>	12 4 8 - - 10 2 6 - -	42	5	10	50	C	4.2	8.4
12.	<i>Azadirachta indica</i>	- - 10 6 9 11 - - 7	43	5	10	50	C	4.3	8.6
13.	<i>Raffia ruffia</i>	- - 14 2 18 3 - -	37	4	10	40	B	3.7	9.2
14.	<i>Acacia nilotica</i>	9 - - 8 6 4 9 - -	36	5	10	50	C	3.6	7.2
15.	<i>Moringa oleifera</i>	- - 10 9 4 7 - 5 - 3	38	6	10	60	C	3.8	6.3
16.	<i>Milicia excels</i>	2 8 6 9 - - 5 - -	30	5	10	50	C	3	6
17.	<i>Cocos nucifera</i>	6 10 9 - - 3 8 4 -	40	6	10	60	C	4	6.6

Table 6 : List of different tree species and other data recorded by Transect method in Obudu L.G.A.

S/N	Name of the species	Segments number 1 2 3 4 5 6 7 8 9 10	Total no. Of individuals Of species	Total no. of segments the species occurred	Total no. of segments studied	Frequency (%)	Frequency class	Density	Abundance
1.	<i>Parkia biglobosa</i>	- - - 7 - 5 - - - 8	20	3	10	30	B	2	5
2.	<i>Elaeis guinensis</i>	18 5 9 8 - 2 4 7 - 6	59	8	10	80	D	5.9	7.3
3.	<i>Hyphaena thebaica</i>	20 8 6 9 - - 10 - 6 -	59	6	10	60	C	5.9	9.8
4.	<i>Vitellaria paradoxa</i>	15 4 3 7 9 - - - -	38	5	10	50	C	3.8	7.6
5.	<i>Isobertina doka</i>	- - - 9 12 - 7 - 5 7	36	5	10	50	C	3.6	7.2
6.	<i>Prosopis africana</i>	2 9 6 4 - - - 7 - 4	32	6	10	60	C	3.2	5.3
7.	<i>Adansonia digitata</i>	- - - - - 1 - - 1	2	2	10	20	A	0.2	1
8.	<i>Ceiba pentandra</i>	7 9 - 4 - - 8 3 - -	31	5	10	50	C	3.1	6.2
9.	<i>Anarcadium occidentale</i>	16 3 8 4 - 2 - 6 9 5	53	8	10	80	D	5.3	6.6
10.	<i>Gmelina arborea</i>	- 10 7 4 - 3 9 - - 2	35	6	10	60	C	3.5	5.8
11.	<i>Magnifera indica</i>	8 6 3 5 7 - - 3 2 -	34	7	10	70	D	3.4	4.8
12.	<i>Azadirachta indica</i>	2 9 3 6 3 - - 1 -	24	6	10	60	C	2.4	4
13.	<i>Raffia ruffia</i>	- - - 9 3 1 8 4 - -	25	5	10	50	C	2.5	5
14.	<i>Acacia nilotica</i>	2 8 - 4 - - - 9 1 -	24	5	10	50	C	2.4	4.8
15.	<i>Moringa oleifera</i>	- - - 5 9 12 3 2 6	37	6	10	60	C	3.7	6.1
16.	<i>Milicia excels</i>	2 9 3 - - 1 - - 7	22	5	10	50	C	2.2	4.4
17.	<i>Cocos nucifera</i>	3 7 4 - 3 - 6 9 - -	32	6	10	60	C	3.2	5.3

Table 7 : List of different tree species and other data recorded by Transect method in Obanliku L.G.A.

S/N	Name of the species	Segments number 1 2 3 4 5 6 7 8 9 10	Total no. Of individuals Of species	Total no. of segments the species occurred	Total no. of segments studied	Frequency (%)	Frequency class	Density	Abundance
1.	<i>Parkia biglobosa</i>	2 - 8 - - 6 - - -	16	3	10	30	B	1.6	5.3
2.	<i>Elaeis guinensis</i>	8 15 9 5 2 6 3 2 - 2	52	9	10	90	E	5.2	5.7
3.	<i>Hyphaena thebaica</i>	6 3 18 26 3 - - - 2	58	6	10	60	C	5.8	9.6
4.	<i>Vitellaria paradoxa</i>	4 7 2 1 9 - - 5 - 8	36	7	10	70	D	3.6	5.1
5.	<i>Isobertina doka</i>	2 6 3 1 6 2 - - - 1	21	7	10	70	D	2.1	3

6.	<i>Prosopis africana</i>	5 2 1 - 4 - - 9 7 -	28	6	10	60	C	2.8	4.6
7.	<i>Adansonia digitata</i>	- - - - 2 - - - 2 -	4	2	10	20	A	4	2
8.	<i>Ceiba pentandra</i>	4 9 6 - - 4 8 - - 2	33	6	10	60	C	3.3	5.5
9.	<i>Anarcadium occidentale</i>	8 2 7 - - - - 9 12 -	38	5	10	50	C	3.8	7.6
10.	<i>Gmelina arborea</i>	- 9 3 7 8 - - - 2 -	29	5	10	50	C	2.9	5.8
11.	<i>Magnifera indica</i>	2 7 9 - 10 3 - - 7 4	42	7	10	70	D	4.2	6
12.	<i>Azadirachta indica</i>	- - - 12 3 1 - 4 8 -	28	5	10	50	C	2.8	5.6
13.	<i>Raffia ruffia</i>	2 8 4 7 - 5 - 3 - 10	39	7	10	70	D	3.9	5.5
14.	<i>Acacia nilotica</i>	2 - - - 6 - - 9 6 4	27	5	10	50	C	2.7	5.4
15.	<i>Moringa oleifera</i>	- - 10 4 3 2 1 - - 2	22	6	10	60	C	2.2	3.6
16.	<i>Milicia excels</i>	3 6 8 - - - 9 - - -	26	4	10	40	B	0.4	6.5
17.	<i>Cocos nucifera</i>	10 4 2 6 - 8 - - 5 8	43	7	10	70	D	4.3	6.1

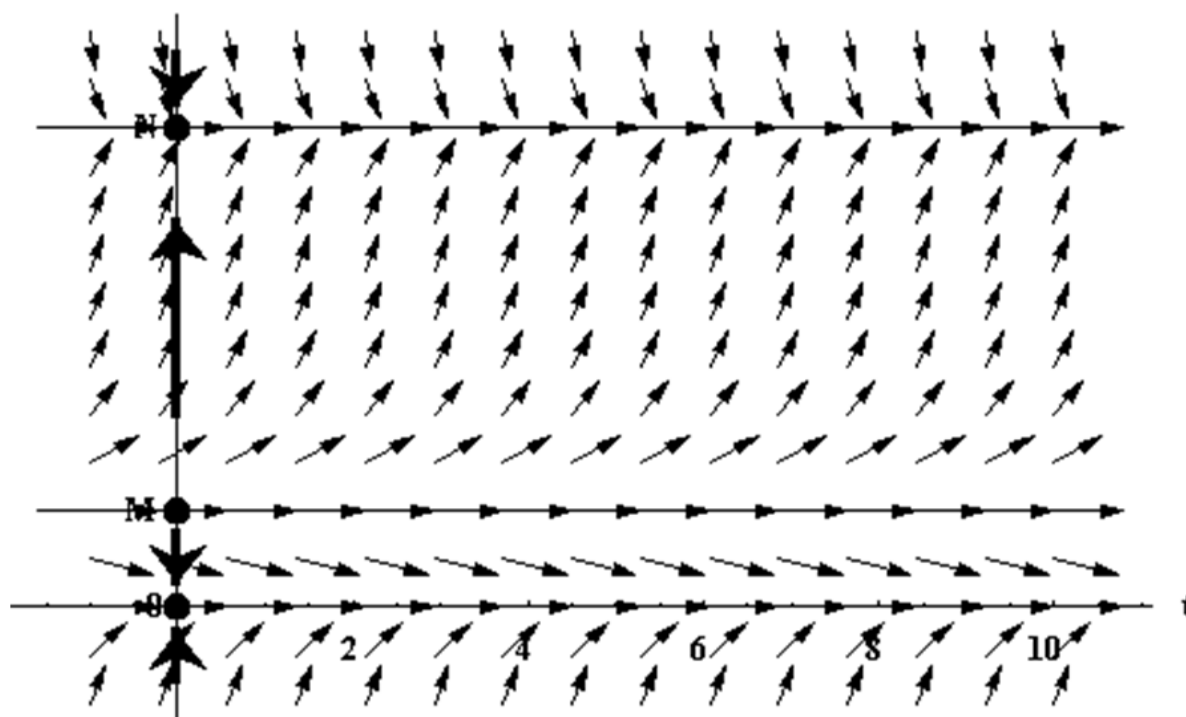
Table 8 : List of different tree species and other data recorded by Transect method in Yala L.G.A.

S/N	Name of the species	Segments number 1 2 3 4 5 6 7 8 9 10	Total no. Of individuals of species	Total no. of segments the species occurred	Total no. of segments studied	Frequency (%)	Frequency class	Density	Abundance
1.	<i>Parkia biglobosa</i>	8 2 - - - - 9 - - 2	21	4	10	40	B	2.1	5.2
2.	<i>Elaeis guineensis</i>	9 4 8 9 5 2 7 4 5 8	57	10	10	100	E	5.7	5.7
3.	<i>Hyphaena thebaica</i>	9 6 22 - 4 8 7 9 4 -	69	9	10	90	E	6.9	7.6
4.	<i>Vitellaria paradoxa</i>	16 8 3 6 2 9 5 - 4 -	53	8	10	80	D	5.3	6.6
5.	<i>Isobertina doka</i>	- - - - 6 10 8 9 - 5	38	5	10	50	C	3.8	7.6
6.	<i>Prosopis africana</i>	2 - - 8 5 7 - - - 5	27	5	10	50	C	2.7	5.4
7.	<i>Adansonia digitata</i>	- 3 2 4 - - - - -	9	3	10	30	B	0.9	3
8.	<i>Ceiba pentandra</i>	6 3 9 - - - - 8 - 7	32	5	10	50	C	3.2	6.4
9.	<i>Anarcadium occidentale</i>	14 3 5 3 7 9 2 1 3 -	47	9	10	90	E	4.7	5.2
10.	<i>Gmelina arborea</i>	- - - 6 9 3 5 2 - -	25	5	10	50	C	2.5	5
11.	<i>Magnifera indica</i>	7 9 6 - - 3 5 2 - 6	38	7	10	70	D	3.8	5.4
12.	<i>Azadirachta indica</i>	- 6 9 2 - - 8 5 - -	30	5	10	50	C	3	6
13.	<i>Raffia ruffia</i>	4 9 7 - 6 - - 3 - 10	39	6	10	60	C	3.9	6.5
14.	<i>Acacia nilotica</i>	- 5 3 - - - 9 9 - -	26	4	10	40	B	2.6	6.5
15.	<i>Moringa oleifera</i>	7 5 3 2 9 - - - - 2	28	6	10	60	C	2.8	4.6
16.	<i>Milicia excelsa</i>	- - - 4 7 3 2 1 2 3 6	28	7	10	70	D	2.8	4
17.	<i>Cocos nucifera</i>	7 3 6 9 3 2 1 4 9 -	44	9	10	100	E	4.4	4.4

b) Threat of extinction

Since *P. biglobosa* is a perennial tree that is confined to a particular habitat (mostly savanna):

If the population is large, their rate of growth decreases or even becomes negative as a result of intra-specific competition. If the population is too small (as with the case in this study), old trees run the risk of not being able to reproduce and as such the rate of growth is also negative. Application of the Modified Logistic Model of population dynamics of *Parkia biglobosa* in this study, showed that at $P=M$ (1.5) (population decreased) and will continue to decrease over time (t) unless viable intervention measures are taken. Based on these findings, It can be concluded that populations less than M (minimum viable population) will decrease to oblivion (face extinction) as shown in (figure 1).



31

Global Journal of Science Frontier Research (C) Volume XVI Issue II Version I

The study has proven that the population of *P. biglobosa* in the Guinea Savanna of Cross River State (North) is rapidly declining at an alarming rate. In the face of threat of extinction due to continued exploitation without replacement, domestication and plantation establishment of this tree to enhance its productivity and to curb the threat of extinction is strongly recommended.

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Overview of the Biology, Epidemiology and Control Methods Against Hard Ticks: A Review

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Overview of the Biology, Epidemiology and Control Methods Against Hard Ticks: A Review

Getahun Asebe ^α, Yacob Hailu ^ο, Ak Basu ^ρ & Dereje Shibru ^ω

Abstract- Ticks are among the obligated ectoparasites that feed blood of vertebrates; particularly mammals; birds; and they are arachnids in the subclass Acari; closely related to mites; surviving for up to several years. This review paper deals about the biology; epidemiology and control methods of hard ticks. Morphologically they are classified into two families known as Ixodidae and Argasidae. All hard ticks feed only on blood of their hosts. They are active during warm periods and easily find their hosts by grappling using their front legs and attach on the suitable site. In their life cycle they have three active stages called larvae; nymphs and adults after eggs is released which is dormant. Three types of Ixodidae are present based on their life cycles called; one; two and three host ticks. Hard ticks life cycles are influenced by factors such as host; climatic; vegetation cover. They widely distributed both in temperate and tropical areas of the world. Hard ticks cause cytotoxicity; cell reaction; and toxic effects on the host besides anaemia. The commonest tick control methods include chemical; ecological; biological; genetic and immunizations systems.

Keywords: control, epidemiology, ixodidae, lifecycle, morphology.

I. INTRODUCTION

Ticks are obligated blood feeding ectoparasite of vertebrates, particularly mammals, birds, and they are arachnids in the subclass Acari, closely related to mites, surviving for up to several years. During this time they feed periodically taking large blood meals, often interspersed with long intervals each meal (Wall and Shearer, 1997). There are at least 884 tick species in two major families, namely the Ixodidae comprises approximately 80% and Argasidae 20%. There are two well-defined families of ticks, the Ixodidae or hard ticks and the Argasidae or soft ticks, and the two groups differ from each other markedly in appearance, habits and development (Pegram *et al.*, 1987).

It has been estimated that 80% of the world's 1,281 million cattle are infested with ticks and while in Africa about 186 million head of cattle are at risk of ticks and Tick-borne diseases (TBDs) (<http://www.fao.org/docrep/U9550T/u9550T04.htm>). Besides the direct impact, ticks also have a greatest impact due to their large number and variety of microbial disease that they

transmit among domestic animals. Mostly the tick-borne infections of humans, farm and companion animals are essentially associated with wildlife animal reservoirs (Baneth, 2014).

Ticks and the diseases they transmit are widely distributed throughout the world (Nicholson *et al.*, 2010; Dantas-Torres, 2007; Piesman and Eisen, 2008), particularly in tropical and subtropical countries (<http://www.mayomedicallaboratories.com/articles/features/tick-borne/>).

Ticks impact on animal production is similar in nature and importance; they are responsible for severe losses caused either by the direct effect of the tick through: tick worry, blood loss, damage to hides and udders and the injection of toxins or through mortality or debility caused by the disease transmitted (FAO, 1984, www.nda.agric.za/publications, Rajput *et al.*, 2006). DeCastro (1997) estimated that the annual global costs associated with ticks and TBDs in cattle amounted to between US \$ 13.9 and 18.7 billion, globally. In Africa, tick-borne diseases are considered the most important animal disease problem (Young *et al.*, 1988). Even though the above estimates are too old reports, yet they reflect the impacts of the problem seriously affecting livestock worldwide. This review paper tried to give an overview of the the Biology, Epidemiology and Control Methods against Hard Ticks.

II. CLASSIFICATION AND MORPHOLOGY

a) Classification of Ticks

Ticks are grouped under the phylum Arthropoda; subphylum Chelicerata; Class Arachnida; Order Acarina (Urquhart *et al.*, 1996). The present tick species number worldwide estimated to be 889 (702 Ixodid, 186 Argasid and 1 Nuttalliella tick (sub-) species), including their synonyms, (limited) distribution data and common names) which is a searchable taxonomic catalogue of all known ticks (Acari: Ixodida) of the world and is facilitated by the International Consortium on Ticks and Tick-borne Diseases (ICTTD) (Nijhof *et al.*, 2016). Family names Ixodidae or "hard ticks" and Argasidae or "soft ticks" indicate the so called by the virtue of their hard dorsal shield and due to their flexible leathery cuticle respectively (Robert *et al.*, 1976; http://parasitipedia.net/index.php?option=com_content&view=article&id=2530&Itemid=2804; Latif and Walker, 2004). Recently a new hard tick species, Ixodes ariadnae has been discovered, adding to the two

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known ixodid tick species (*I. vespertilionis* and *I. simplex*) of bats in Europe (Hornok *et al.*, 2015). They are dioecious having separate sex (Furman and Loomis, 1984).

b) Morphology of Ixodidae

Ixodid ticks are relatively large, ranging between 2 and 20 mm in length. The body of the unfed tick is flattened dorsoventrally and is being divided into only two sections; the anterior gnathosoma (or capitulum) and posterior idiosoma, which bears the legs. The terminal gnathosoma is always visible when an ixodid tick is viewed from the above. The gnathosoma carries a pair of four segmented palps, which are simple sensory organs, which helps the tick to locate its host. The fourth segment of each palp is reduced and may articulate from the ventral side of the third, forming a pincer-like structure. Between the palps lies a pair of heavily sclerotized, two-segmented appendages called chelicerae, housed in cheliceral sheaths. At the end of each chelicera is a rigid, somewhat triangular, plate bearing a number of sclerotized tooth-like digits. The chelicerae are capable of moving back and forth and the tooth-like digits are used to cut and pierce the skin of the host animal during feeding (Wall and Shearer, 1997). The enlarged, fused coxae of the palps are known as the basis capituli. The basis capituli varies in shape in the different genera, being rectangular, hexagonal or triangular. The lower wall of the basis capituli is extended anteriorly and ventrally to form an unpaired median hypostome, like an underlip, which lies below the chelicerae. The hypostome does not move but in larvae, nymphs and adult females are armed with rows of backwardly directed ventral teeth. The hypostome is thrust into the hole cut by the chelicerae and the teeth are used to attach the tick securely to its host. As the hypostome is inserted the palps are spread flat on to the surface of the host's skin (Wall and Shearer, 1997; Robert *et al.*, 1976).

c) Biological Characteristics of Hard Ticks

i. Feeding

Ticks feed only on the blood of their hosts. According to Radostits *et al.*, (1994) all are blood suckers and may cause death from anaemia. All ticks at each stage of the life cycle are parasitic feeding solely on blood, liquefied tissues, and tend to feed at varying depths according to the species. Their bite is relatively painless, but invasions by large number are debilitating (Gray, 1995; Robert *et al.*, 1976; Kemp *et al.*, 1982).

The salivary gland of ticks produces a secretion, which prevents blood from coagulation. Blood is pumped in by a muscular pharynx and forced backward into the oesophagus and midgut. Diverticula attach to the mid-gut become greatly distended to store a large quantity of blood. The outer body covering (cuticle) is leather-like and capable of great distension during

feeding. Un-engorged female ticks are somewhat flattened and tapered anteriorly, but become engorged as much as twenty fold following feeding; male hard ticks take only small blood meals and increase in size only slightly (Bay and Harris, 1988; Robert *et al.*, 1976).

ii. Mechanism of finding their host

Most ticks are active during warm periods of the year and go into hibernation or become dormant during the cold periods; they hide within fissures in the ground, under rocks or in cracks and in buildings (Galloway, 1974; Robert *et al.*, 1976). Ticks find their hosts in several ways. Some ticks live in an open environment and crawl onto vegetation to wait for their hosts to pass by. This is a type of ambush and the behaviour for waiting on vegetation is called questing (Kettle, 1984; Walker *et al.*, 2003). Thus in genera such *Rhipicephalus*, *Haemaphysalis* and *Ixodes* the larvae, nymphs and adults will quest on vegetation. The ticks grab on to the hosts using their front legs and thus crawl over the skin to find a suitable place to attach and feed (Yismashewa, 2005).

Much of the behaviour in ticks is regulated by pheromones; this compound greatly enhances inter-species communication, where vision, auditory and tactile means of communication is not believed to play a significant role. Pheromones modulate a wide range of behaviour, from mating (sex pheromones) to warning other members of the species of impending communication via pheromones affecting assembly, mating and host finding has been demonstrated in ticks (Harwood and James, 1979). Basically tick hormones classified based on their activity profiles tick pheromones are characterized as: (1) assembly pheromones, (2) aggregation/attachment pheromones, (3) pre-attachment or prefeeding pheromones, (4) attractant sex pheromones, (5) contact sex pheromones, (6) fecundity-reducing pheromone and (7) various ungrouped pheromones (Gothe, 1987).

Adult ticks of the genera *Amblyomma* and *Hyalomma* are active hunters; they run across the grounds to seek hosts that are nearby. The general behaviour of seeking hosts in an open environment is described as exophilic. Ticks such as *Argasids* and many *Ixodes* species spend their life cycle in their hosts' nest and attach to their hosts there. This is endophilic or nidicolous behaviour. A few species of ticks, such as the dog tick *Rhipicephalus* attach to animals there. This is called domestic behaviour (Walker *et al.*, 2003).

iii. Reproduction

There are three active stages in the life cycle of hard ticks, larvae, nymphs, and adults; each instar takes a blood meal only once and long periods are spent on vegetation between meals. In the hard ticks, mating takes place on the host, except in species of the genus *Ixodes* where it may also occur when the ticks are still on the vegetation. Male ticks remain on the host and

will attempt to mate with many females whilst they are feeding. They transfer a sac of sperm to the female. The females mate only once before they are ready to fully engorge with blood. When they finally engorged, they detach from the host and have enough sperm to fertilize all their eggs (Walker *et al.*, 2003). Oviposition usually begins a few days after the female dropped from the host and continues for several days (FAO, 1984). The female dies afterward. Males remain to the host for long period and may mate with other females (Yismashewa, 2005).

In a single batch a female hard tick may lay eggs (2,000 to 20,000). The place where all ticks, laid eggs in the physical environment, never on the host (Hoogstral, 1956; Walker *et al.*, 2003). The temperature and humidity affects largely the hatching of the eggs and the activity of the larvae. Hot conditions are favourable for hatching (Hoogstraal, 1956 and Gebreab, 1983).

iv. Life cycle of Ixodidae

In *Ixodidae*, three types of life cycles can be distinguished based on similarities or differences in tropisms shown by ticks at different instars. These are the monotropic cycle or one-host, the ditropic cycle or two-host and the telotropic cycle or three-host ticks. According to the species, the development is either completed on one, two or three hosts. (Brian, 1997; Walker *et al.*, 2003)

In **one-host ticks**, the larvae that emerge from the eggs three to four weeks after deposition at the earliest attaches themselves to host animals where they complete their entire development. On the host they develop to nymph then to adult and then copulate. Afterwards, they drop off and deposit their eggs on the ground. The entire development cycle takes mostly 19-21 days as a rule, with minimum of fifteen and maximum of 40 days, each stage taking one week (Seifert, 1996). In these ticks a stricter adaptation eliminates the need to drop to the ground for metamorphosis. All the instars occur on a single vertebrate, attacked by the larva. The larval and nymphal metamorphoses take place on the host, at the point of attachment of the larva and nymph. There is only one parasitic phase (Douglas, 1969).

The **two-host ticks** attach it as a larva to a host, feeds on blood and develops into the nymph stage. After a maximum of 14 days, it drops off on to the ground where it reaches the imago stage in 20-30 days time. Male as well as female ticks then look for another host, feed on blood and copulate. After 6-11 additional days, the female drops to the ground and deposits its eggs. The entire cycle from the time the larva emerges until the engorged female deposits the eggs mainly depends on the time the adult spent on the ground to find a new host. According to the species the nymph may survive on the ground for several weeks (Seifert, 1996). In these ticks the three stages develop on two

different individuals that may or may not belong to the same species. In the first phase, the engorged larva molts on the host and the nymph reattaches close by. At the end of the blood meal, the nymph detaches and metamorphoses on the ground. Engorgement of adults occurs during the second parasitic phase. There are only two searches for a host, which eliminates the risks linked with the need for nymphal attachment (Hoplaet *al.*, 1994).

The **three-host ticks** looks for a new host during each stage of development in order to feed. The larva emerges from the egg on the ground, looks for a host, feeds on it for three to seven days, drops off and molts after three to four weeks on the ground. The nymph attacks a second host for three to seven days feed on it and drops and develops into an adult on the ground after two to eight weeks. After that, the adult tick looks for a third host to feed on and for copulation that takes one to three weeks. Finally it drops off and completes the cycle with oviposition on the soil. Because of the different time spent in each stage on the ground, the entire development cycle may last up to one year (Seifert, 1996). These ticks require three hosts for development, irrespective of the host species. There are three parasitic phases, separated by two phases on the ground, when metamorphosis occurs (Hopla *et al.*, 1994).

d) Life cycle influencing factors

i. Host factors

a. Character of the hosts

The adults of many ticks occur on the grass cover and have access to a wide range of host's ungulates or carnivores, wild or domestic. They are not specific but selective towards a group of vertebrates based on their size and origin and mobility. The indigenous wild animals of a given region are important factors regarding the origin and maintenance of population of domestic mammal ticks. The different instars of a given species may or may not have definite microclimatic requirements. These determine their location in the most suitable microhabitat and the choice of hosts according to availability at different levels of the plant cover (Morel, 1980). Specific ticks are associated with hosts of a definite resting site or habitat. The area is small or with a distinct environment nest, burrows, caves, rock piles, thickets, dense specialization in relation to that of the host, rather than phylogenetic specificity of the tick (Samir, 1980; Morel, 1989).

b. Predilection sites on the hosts

On initial infestation, ticks were picked up during the day when cattle were grazing, and attached temporarily near the hooves. Subsequently, when cattle rested, especially at night, the ticks detached and then reattached more firmly at the usual predilection sites. Many ticks disappeared during relocation, and other

ticks transferred to different hosts. The tick's location on the host is linked to the possibility of penetration by the hypostome. On ungulates, species with a short hypostome. On usually attach to the head within the ear, nape of the neck, margin of the anus, and under the tail. Long-hypostome. On species attach to the lower part of the body where the skin is thicker, such as the dewlap, groin, udder, testes, perineum, and margin of the anus. Small ticks, all instars of Genus *Rhipicaphalus*(*Boophilus*), larvae and nymphs of *Amblyomma* have no marked preference, and can be found all over the body (Strachurski, 2000).

ii. Climatic factors

Ticks are found on all continents and are bound to certain climates as far as the requirements for temperature, humidity; sun-radiation and shade of each species are concerned. Furthermore, each species requires specific environmental conditions for its habitat. The respective species only have a chance for survival when these prerequisites are fulfilled (Seifert, 1996). Ticks in a tropical zone the more rapid development pattern are determined during the beginning rainy season. The whole cycle takes one year. Whereas, ticks in an equatorial zone climatic uniformity and the absence of an unfavorable season allow development throughout the year. There is no annual cycle determined by a diapause's, generation follow one another in a pattern depending on the species (Walker and Fletcher, 1982).

a. Temperature: a dynamic factor

Each species has its particular threshold temperature below that diapauses occurs in all instars. Egg and larvae development, and egg production in engorged females are inhibited, while immature and unfed adults become quiescent. The average weekly or monthly temperature is useful for predicting the activity threshold and optimum temperature. The tick development and activity periods can be determined from monthly isotherm charts (Morel, 1989).

b. Relative humidity: a static factor

Relative humidity is considered at microclimate level. Humid rather than wet conditions are essential for the development and survival of eggs, and the survival of unfed hatched larvae. Each species is adapted to a particular relative humidity range in a biotope and it varies with the instars and its size. Larvae and nymphs have high humidity requirements, whereas the adults can protect themselves better against evaporation because of their larger size and thicker tegument. The requirement ranges from 100 % to very low relative humidity.

Larvae and nymphs adept their humidity requirements by developing in holes in the ground, cracks in rocks, litter, and the base of the vegetation layer and other shelters places. They may alternatively immediately seek a host and not leave it before the

engorged female stage. The surface of medium-height (30-150 cm) vegetation and especially bare ground (sand, pebbles, rocks) are less protected. Ticks rarely occur in these sites except under special circumstances in all seasons if adults have thick teguments, or only in the rainy season, or if the site is shaded by a tree to prevent evaporation. Larvae or nymphs found in open spaces are usually active in the rainy or cool seasons (Morel, 1989).

iii. Climate related factors: seasonal activity

Climatology includes the particular temperature and humidity conditions prevailing in a country. These parameters are the result of the simultaneous action of several factors, such as latitude, altitude, and their effects sunlight, temperature, rainfall, wind patterns. On a regional scale, these data should be studied for a better understanding of vectors and the diseases they transmit, especially for the application of control measures (Pegram *et al.*, 1982). In a region with uniform conditions, a comparison of data on species distribution with isotherm and isohyets maps enables the identification of natural distribution zones in relation to latitude. In mountainous regions, the determining factor is altitude. Various climatic factors condition the presence or absence of a tick species. According to its micro or mesoclimatic requirements, the species will be found in certain similar bioclimatic zones, and not in others. Moreover, seasonal variations within a bioclimatic zone will favor or hinder the development or activity of a tick species during certain periods. In tropical climates, the dominant factor is rainfall. The start and end of the rainy season influence the different phases of the life cycle. Parasitism is reduced during the dry months and increases sharply within days following the first major winter rainfall. The population remains stable for a few weeks, and then slowly diminishes. At the end of the rainy season, there is a marked decrease, with progressive fall to almost zero in the dry season. Tick distribution therefore corresponds to the isohyets. In these cases, the cold and dry seasons impose pattern on tick development that can be observed in parasitism in large mammals. The cycle of seasons determines the alternation of appearance, reduction, and disappearance of ticks. These variations in tick populations represent the frequency or seasonal dynamics of a species, or its phenology (seasonal pattern of appearance) (Fourie *et al.*, 1996).

iv. Vegetation

The plant cover as a whole is not an inert intermediate factor between climatic phenomena and the fauna, since it is not in depressant of these factors. It is the result of the adaptation of a particular flora to the temperature, rainfall, and wind patterns prevailing in an area with particular geological and pedological characteristics. In turn, vegetation is also related to temperature and rainfall. Its distribution and feature in

given latitude and altitude represent equilibrium. It is a response to external conditions that creates variety of microclimates at different levels and physical support to the fauna (Glen and Pete, 1969). Vegetation is not only the result of various elements that make up the environment, but it also determines, by its composition, the various microclimates at different levels. It is the best ecological integrator, which influences the biological phenomena seen at a given point. A comparison of the distribution of tick species with the features of the vegetation in a natural zone is very useful. It is of practical use for determining the distribution of a given species, with all the consequences concerning the epidemiology of diseases caused or transmitted by ticks and the possibilities of controlling them (Morel, 1989).

III. EPIDEMIOLOGY AND DISTRIBUTION OF HARD TICKS

a) Epidemiology

The distribution of ticks in a temperate climate with frequent and non-seasonal rainfall is closely linked with availability of a micro-environment with a high relative humidity such as occurs in the mat which forms under the surface of rough grazing. In contrast, in tropical grazing areas the grass cover on pasture is discontinuous and often interspersed with bare or eroded patches. Where suitable grass cover does exist it has been generally accepted. Since temperatures are suitable for development throughout a large part of governed by rainfall, and with the exception of *Hyalomma* spp., a mean annual rainfall of more than 60cm is required for survival. However, recent studies in East Africa have shown that the factors underlying the maintenance of the necessary microclimate with a high relative humidity are rather more complex and depend on the transpiration of plant leaves. As long as this continuous adequate humidity is maintained in the microclimate despite the dryness of the ambient temperature. However, when the rate of evaporation increases beyond a certain level, the stomata on the leaves close, transpiration ceases and low humidity created in the microclimate rapidly becomes lethal to the ticks. In the field of course, the stability of the microclimate is dependent on factors such as the quantity of herbage or plant debris and the grass species. The various genera of ticks have different threshold of temperature and humidity within which they are active and feed and their distribution is governed these threshold. Generally, ticks are most active during warm season provided there is sufficient rainfall, but in some species the larval and nymphal stages are also active in milder weather and this affects the duration and timing of control programmes (Urquhart *et al.*, 1996).

In recent years, it is common to see increasing of the geographic coverage as well as the incidence and prevalence of TBDs affecting domestic animals and

humans has increasing. many important zoonotic TBDs, such as anaplasmosis, babesiosis, ehrlichiosis, and Lyme borreliosis are increasingly gaining more attention from physicians and veterinarians worldwide. With the help and timely instruments developed techniques such as molecular biology, it can be easy to isolate new species, strains, or genetic variants of microorganisms present in ticks worldwide (Pacheco *et al.*, 2011; Duh, *et al.*, 2010) (Table 1). Some of the list of potential tick-borne pathogens continues to increase (Shapiro *et al.*, 2010; Pritt *et al.*, 2011; Silva *et al.*, 2011; Subramanian *et al.*, 2012).

Agents, such as *Rickettsia slovaca*, *Rickettsia parkeri*, and *Rickettsia massiliae*, were identified in ticks, decades before these were associated with human diseases (Paddock, 2009). Many other tick-borne pathogens, including flaviviruses (e.g., Omsk hemorrhagic fever virus, Powassan encephalitis virus, and Kyasanur forest disease virus) have been implicated in human disease in new geographical regions (Dobler, 2010; Piesman, and Eisen, 2009) (Table 1).

Table 1 : Distribution of Ticks and TBDs

Diseases	Pathogens	Tick species/vectors	Geographic distribution	Reported in
Mediterranean spotted feve	Rickettsia conorii	Rhipicephalussanguineus,R. turanicus	Africa, Asia, Europe	(Jongejan and Uilenberg, 2004; Piesman and Eisen; 2008; Matsumoto et al. , 2005)
Lyme borreliosis	Borrelia burgdorferi sensu lato	Ixodes hexagonus,I. pacificus, I. persulcatus, I. ricinus, I. scapularis	Asia, Europe, North America	(Jongejan and Uilenberg, 2004; Piesman and Eisen; 2008)
Human monocytic ehrlichiosis	Ehrlichia chaffeensis	Amblyomma americanum	North America	(Jongejan and Uilenberg, 2004; Piesman and Eisen; 2008)
African tick bite fever	Rickettsia africae	Amblyomma hebraeum, A. variegatum	Africa, West Indies	(Jongejan and Uilenberg, 2004; Piesman and Eisen; 2008)
Human granulocytic anaplasmosis	Anaplasma phagocytophilum	Haemaphysalis concinna,H. punctata, Ixodes ricinus, I. pacificus,I. scapularis,Rhipicephalus bursa	Europe, North America	(Jongejan and Uilenberg, 2004; Piesman and Eisen; 2008)
Q fever	Coxiella burnetii	Many species of differentgenera	Africa, Asia, Australia, Europe, North America	(Jongejan and Uilenberg, 2004; Piesman and Eisen; 2008)
Relapsing fever	Borrelia spp.	Ornithodoros spp.	Africa, Asia, Europe, North America	(Jongejan and Uilenberg, 2004; Piesman and Eisen; 2008; Cutler, 2010)
Rocky Mountain spotted fever	Rickettsia rickettsii	Amblyommaamericanum,A. aureolatum,A. cajennense,Dermacentor andersoni,D. variabilis,R. sanguineus	North, South and Central America	(Jongejan and Uilenberg, 2004; Piesman and Eisen; 2008; Labruna, 2009; Breitschwerdt et al.,2011)
Tularemia	Francisella tularensis	Many species of differentgenera	Asia, Europe, North America	(Jongejan and Uilenberg, 2004; Piesman and Eisen; 2008; Milutinovic et al., 2008; Petersen et al., 2009; Gyuranecz et al., 2011)
Babesiosis	Babesia divergens, B. microti	Ixodes ricinus, I. scapularis	Europe, North America	(Jongejan and Uilenberg, 2004; Piesman and Eisen; 2008)
Colorado tick fever	Coltivirus	Dermacentor andersoni	Western North America	(Jongejan and Uilenberg, 2004; Piesman and Eisen; 2008)
Crimean–Congo hemorrhagic fever	Naivirus	Amblyomma variegatum,H. punctata, Hyalomma anatolicum,H. marginatum,H. truncatum, R. bursa	Africa, Asia, Europe	(Jongejan and Uilenberg, 2004; Piesman and Eisen; 2008)
Kyasanur forest disease [1,5]	Flavivirus	Haemaphysalis spinigera,H. turturis	Indian subcontinent	
Louping ill	Flavivirus	Ixodes ricinus	Western Europe	(Labuda. and Nuttall,2004)
Omsk hemorrhagic fever	Flavivirus	Dermacentor marginatus,D. reticulatus,I. persulcatus	Asia	(Piesman and Eisen; 2008)
Tick-borne encephalitis	Flavivirus	Ixodes persulcatus,I. ricinus, H. concinna,H. punctata	Asia, Europe	(Piesman and Eisen; 2008; Labuda. and Nuttall, 2004)
Powassan encephalitis	Flavivirus	Dermacentor andersoni,Haemaphysalislongicornis, I. cookei,I. scapularis	Asia, North America	Dobler, 2010; Labuda. and Nuttall, 2004)

IV. PATHOGENIC EFFECTS OF TICKS

a) Cytolysis effects

The primary attachment lesion causes cytolysis following production of the hyaline sheath. There is itching accompanied by a tissue and humoral reaction

of the host, with hyperemia, eosinophil infiltration and a local edematous reaction. The damaged tissues are pulled by the weight of the feeding tick and this produces a sensation of pain (Morrisan, 1989).

The salivary glands of ticks perform numerous vital functions. They secrete cement that anchors the

mouthparts to the skin soon after the tick attaches to the host. The salivary glands are the major organs of osmoregulation and possess a water vapor uptake mechanism that enables ticks to fast for many months. During feeding the salivary glands of the female secrete excess fluid from the blood meal back in to the host's circulation, thus concentrating the nutrients components of the meal and regulating hemolymph volume and ionic composition. These factors lead to the reaction of the host. The primary challenge or infestation: when ticks attach to a host (primary infestation), they secrete cement and other antigenic materials. In the guinea pig, 40-60 % of the leucocytes infiltrating the lesion are neutrophil, up to the third day of feeding. A day or two later when most of the larvae have engorged the proportions of basophils and eosinophils have increased significantly. Early in tick attachment, the capillaries near the mouthparts become dilated and edema results. This is probably due to vasoactive substances, in tick saliva such as prostaglandins, and to a few deregulated mast cells, which release histamine. Hemorrhage is an obvious characteristic of the feeding lesion. Finally, the langerhans cells of the epidermis trap antigenic material from tick saliva and present it to lymphocytes in the skin and lymph nodes for processing.

- (a) Tick attaches to host. Secretes antigens tissue damage Antigens taken up release of mediators Langerhans cell in skin. Antigens presented to lymphocytes inflammation T-lymphocyte activation mild degranulation of mast cells and basophils by slavery B-lymphocyte activation (assumed) gland esterases IgG IgE Complement erythema and edema activation (primary immune response) (inflammatory response) Host reaction to tick challenge by primary infestation (Morrisson, 1989).
- (b) Reactions to a primary infestation. Other than a mild inflammatory response leading to erythema and edema, there are few obvious clinical reactions. The specific immune response involves the activation of T- and B- lymphocytes.

Secondary challenge or infestations: During secondary infestation, the granulocytes in the peripheral blood rise to much higher levels than during the primary infestation and basophils rapidly invade the feeding lesion, degranulate and liberate vasoactive mediators. The later cause edema and might contribute to the formation of blister-like epidermal vesicles underneath the attached ticks. Histamine 5-HT inhibit feeding and salivation, the histamine may induce detachment from the host. Premature detachment and much reduced engorged weights may result and those ticks that do not detach feed very slowly, if at all, and most die *in situ*. As a result of the laceration of blood vessels by the probing actions of tick mouthparts, the host's circulatory homeostatic mechanisms begin to act and in response to these, ticks release salivary secretions to maintain

blood flow to the feeding lesion. However the salivary composition of a particular species may be a partial determinate of effective host range. Thus, ticks that possess antihistamine salivary activity probably feed successfully on basophile-rich hosts (cattle, guinea-pigs, rabbits) but may be rejected more effectively by mice, which rely on bradykinin and anaphylatoxins as vasoactive mediators (Bianchi *et al.*, 2003).

After detachment of the tick the necrotic lesion remains indurate, itching, and hot, and can discharge for several months. Bacterial complications may set in with abscess formation. The hypostome may break and remain in the lesion when a tick is extracted. In receptive cattle, the tick remains in place, and lesion is formed with eosinophil infiltration around the necrotic patch near the chelicerae. The allergic reaction is sign of host resistance (Tatchell, 1969).

b) Cell reaction

The lysis cavity opens into internal tip of the sheath, and is formed of amorphous necrotic tissue, with cutaneous and blood cell debris. An edematous zone surrounds the lysis area when the cell structures gradually disappeared collagen remains intact and extravasations and vascular ruptures occur, with hemorrhagic patches. In receptive hosts, the main phenomenon is neutrophil infiltration around the attachment point and capillaries. The resulting inflammation produces vasodilatation, vascular ruptures, and hemorrhages near lysis cavity, while the surrounding dermis becomes fibrous due to the multiplication of fibroblasts. The other cells degenerate at the same time. The dermis and epidermis become edematous. Vesicles and necrotic zones appear at the end of the attachment process, in the presence of eosinophils and basophile. In the receptive host, the main phenomena are vasodilatation and vessels rupture (Allen, 1994). In resistant hosts, tissue reactions are more violent and occur earlier. They are dominated at first by considerable edema of the epidermis and dermis, accompanied by eosinophil and basophile infiltrations from the second day of attachment. Cell decomposition and tissue necrosis are rapid and extensive, but there are not many capillary ruptures. The vesicles appear very early and develop in to pustules. Although the tick saliva has a limited lysis effect, it causes inflammation through degranulation and high antigenic activity. The ingested blood is concentrated by excretion of water and mineral salts starting from the beginning of the meal. Highly engorged species therefore ingest about three times the volume of blood at the end of the meal (Uspenskiy, 1982).

c) Toxic effects (tick toxicosis)

Apart from the mechanical cytolysis effect and blood loss, ticks have a specific pathogenic effect due to the presence of toxins in the saliva. The toxins affect not only the attachment site but also the entire organs of

the host. The toxins act on certain tissues. Neurotropic toxins induce tick paralysis, while dermatotropic toxins cause sweating sickness (Tatchell, 1969; Allen, 1979).

V. CONTROL MEASURES FOR TICKS

a) *Steps of a controlling campaign*

The aim of a tick control campaign is not to control all ticks simultaneously, but a definite species because of its particular role. The strategy should therefore be based on the biological characteristics of the target species. Moreover, there is no perfect control method. The efficacy of these methods depends on rational and methodical use (FAO, 1977; Jongejan and Ulenberg, 1994).

Factors to be defined when a campaign is planned against ticks: -

i. *Campaign objectives*

The objectives must be defined on the basis of the biology of the species and epidemiology of the disease caused or transmitted by the particular tick species. Temporary or regular deparasitizing of infested animals is a short- or medium-term measure. It provides relief for the host, but does not affect ticks in pastures. Reduction of tick populations in a pasture has long-term prophylactic effects as it decreases parasitism and frequency of pathogen inoculation to a tolerable level. Premunition is established or maintained by regular treatments on fixed dates or occasional treatments when the parasitism rate exceeds a certain level. Eradication of ticks has a long-term prophylactic effect and controls transmission of a particularly dangerous pathogen (Kaaya, 2003).

ii. *Target location*

A site is selected for the campaign from the different successive microhabitats of a tick in its free-living phases, and during the parasite phases on the host. The selection is based on ecological data. The control operations are carried out in the field and on the host. In the field, the objective is to attack ticks in their microhabitat during the free-living phases by chemical or ecological measures. On the host, the aim is either to rid it directly of its parasites, or indirectly by means of hosts that collect ticks and serve as bait. This enables long-term deparasitizing of pastures, which is the final goal (Okello et al., 2003).

iii. *Campaign schedule*

Information on dynamics is useful for determining the pattern of host treatments and suspension of treatments during seasons of inactivity, on the basis of the life cycle and duration of the parasite phase during seasons of tick activity. In methods requiring removal of livestock, data on survival possibilities of the various instars without feeding will determine how long cattle should be excluded from the pastures. The period required for completion of the cycle should be considered, in the case of species with

certain instars that develop on hosts other than ungulates or on wild vertebrates for a part of the tick population (Jongejan and Ulenberg, 1994).

b) *Types of tick control*

Tick biology data are fundamental factors in chemical control and represent method in biological control. The two control methods differ only in the use of acaricides. Procedures directly affecting the microhabitat and host availability such as using hyperparasites and predators, and immunological control form a part of an integrated biological control program. The practical importance of these methods varies. Some are effective on their own, but it is important to combine them. The use of acaricides is inconceivable without data on the natural environment of ticks and their hosts (Kemp, 1994).

i. *Chemical control*

Arsenic was the first compound to be widely used for tick control but due to problems of toxicity, lack of residual effect and resistance it was largely replaced by the organochlorines in the late 1940s. Increased environmental contamination and consumer resistance to unacceptable levels of organochlorines in meat together with onset of tick resistance to this group of insecticides led to their replacement in the 1960s by several organophosphorus compounds, the carbamates, butocarbonyl, more recently the formamidine, amitraz, and some synthetic pyrethroids. Ivermectin or closantel given by the parental route have also been shown to be useful aid in the control against the one-host tick *Rhipicephalus (Boophilus)* (Urquhart et al., 1996) (Table 2).

Table 2 : Some of the chemicals used for tick control

Group name	Chemical name
Organophosphates	Bromocyclene
	Coumaphos
	Crotoxyphost dichloruos
	Diazinon
	Dichloruos
	Malathion
	Phoxin
	Propoxur
Carbamates	Stirofos
	Trichlofon (Metrifonot)
Chlorinated Hydrocarbons	Carbaryl
	Lindone
	Methoxychlor
Pyrethrins and Pyrethroids	Flumethrin
	Fenualerate
	Permethrin
	Pyrethrin
Avermectin	Ivermectin

Source: Kaufmann (1996).

Avermectins

Avermectins are products of the fungus *Actinomyces* species that are divided into two major components A and B depending on the structures. Avermectins compound is active against intestinal parasites, lungworms, warble flies, lice, midges and different genera of ticks. The active substance inhibits the transmission of stimuli between the interneurons of the ventral nerves and the motor neurons of the parasites. The compound is deposited for a short time in the liver, fat and only in small amounts in muscle and kidney. About 50% is excreted unchanged with the feces. Therefore, a withdrawal time of 38 days is required for milk and beef. Because of this, it also does not make sense to utilize the compound in tropical animal production (Seifert, 1996).

ii. Ecological control

Information on the ecology of different instars is used for habitat and host linked treatments. Tick control in the habitat and vegetation requires modification of the plant cover by removal of vegetation that shelters ticks. Vegetation is periodically removed by burning, but spontaneous or induced fires have little direct effect on ticks since they occur in the season when adults are not active. Annual dry-season fires are widespread in semiarid regions; the value of these fires is very controversial, as they influence not only the availability of an important source of grazing during long harsh dry seasons, but may also diminish the abundance of ticks and vermin such as rats. The influence of burns on tick abundance varies markedly with the time of year, intensity of burn, and the tick species present (Wilkinson, 1979; Baars, 1999).

Replacement of natural vegetation, cropping, and soil cultivation are integrated methods that enable pasture improvement and tick eradication. Using

acaricides, careful plastering of walls and ceilings, and installing concrete floors sanitize localized habitats such as stables, sheds, poultry houses, and kennels and the soil of cattle enclosures, markets, areas around wells, etc.

Wild ungulates and carnivores, as well as other possible hosts of the different instars should be eliminated. Since wild ungulates are alternative hosts and can maintain tick populations as effectively as cattle, their removal is the most effective measure. The objective of briefly or periodically withdrawing domestic hosts is to cause the ticks to disappear through inanition since the only available hosts are cattle. Pasture rotation can be used exclusively, or together with the use of acaricides for this purpose. The rotation pattern and pasture area should be determined according to the target species, type to cycle, and hosts of the successive instars, as well as the period of stay and number of cattle. Other important factors are the period required for inanition by each instar, periods of tick activity depending on the seasons and type of climate (McCosker, 1979).

iii. Biological control

Entomopathogens are group of organisms that attack ticks and insects. Entomopathogens can be macro- or microorganisms that affect arthropods. In biological tick control the activities of the hyperparasites *chalcid* flies *Hunterellus* are probably important in nature, but they are difficult to evaluate. It is still more difficult to manipulate or reproduce them for practical use. Predators are most effective, especially ants and birds (*Buphagus* sp. *Oroxpeckers*, *Crotophagus*, various magpies, village fowl). Depending on the conditions, these predators can consume a large number of ticks (Samish and Alekseev, 2001).

iv. Genetic control

In resistant animals, the blood level of histamine rises considerably within 48 hours of tick attachment. In moderately resistant cattle, there is only a slight increase in the histamine level when larvae attach; it is higher for nymphs, and even higher for females. In receptive animals, there is no change in the histamine level. Antigens extracted from eggs or larvae give the same results in resistant animals. The reaction intensity is not correlated, however, to the degree of resistance (Willadsen, 1999). Resistance to ticks in animals occurs early but is effective mainly in adults. This resistance can be broken by the occurrence of physiological or pathological disorders. The diseases induced or transmitted by ticks can also reduce resistance and are accompanied by an increase in the animal's quantitative tolerance threshold for ticks, which may lead to severe parasitism, with clinical signs (Willadsen and Jongejan, 1999).

The ability is a hereditary character. The ability to resist ticks is acquired, but it develops according to genetic factors. If it is possible to breed cattle for tick resistance, as is done for certain other characteristics, it will provide a solution allowing the costly and dangerous use of acaricides to be abandoned, with the related risk of the emergence of non-sensitive tick strains. The question remains whether breeding for tick resistance can be compatible with breeding for a particular production characteristic (meat, milk) (Brown and Askenas, 1982).

c) Immunization (anti-tick vaccine)

Research in to controlling cattle tick *Rhipicaphalusmicroplus* (formerly known as *Boophilusmicroplus*) has focused on developing vaccines; cattle vaccinated with antigens from the midgut of female ticks were protected from challenge with *Rh. microplus* (Opdebeeck et al., 1988; Willadsen et al., 1988; https://en.wikipedia.org/wiki/Rhipicephalus_microplus).

Ingestion of the blood meal by ticks feeding on vaccinated cattle leads to uptake of antibodies and other components of the host's humoral immune system, resulting in damage to the gut. As a result, the number of ticks engorging, their average weight and their ability to lay eggs may all be adversely affected. Australian tick control project develop a vaccine against *B. microplus*. It was commenced in 1981 with the observation of the cattle, vaccinated with crude or partially purified material from semi engorged adult female ticks, could be effectively protected against tick infestation. The major antigen responsible for this effect, called Bm86, was isolated 4 years later. The antigen is located on the surface of the digest cells, which line the tick's gut. The feeding tick takes in antibody to Bm86. Binding of antibody to the protein on the tick gut leads eventually to lysis of the tick's gut cells. The number of

ticks engorging on vaccinated cattle is reduced tick weight and even stronger reduction in the ability of the female ticks to lay eggs. In fact, there is considerable mortality of engorged ticks in the first days following engorgement. The antigen has been sequenced and expressed in *Escherichia coli* or *Aspergillus nidulans* (Willadsen et al., 1989; Tellam et al., 2002).

This recombinant tick vaccine was registered for commercial sale in Australia in 1994, called "Tick Guard®". With this vaccine over 18000 cattle had been vaccinated in northern Australia. In a variety of tests of efficacy and safety it was shown that the vaccine was completely safe for use, effective in a variety of different geographical locations and effective against a wide range of ticks that were resistant to the different classes of chemical acaricide. It was found that there were some differences in the vaccine susceptibility of different isolates of tick for reasons, which are not understood. It was further shown that a range of different cattle breeds responded to the vaccine in equivalent ways, as measured by antibody responses following vaccination (Willadsen, 1997).

VI. CONCLUSIONS AND RECOMMENDATIONS

In Ixodid ticks, there are only two moults following larval hatching: larvae to nymph and nymph to adult, both of which usually occur on the ground. However, in the 2-host cycle, larvae moult to nymphs on the host. Adult Ixodid females usually mate and complete engorgement on the host and then detach and drop to the ground prior to ovipositing. Depending on temperature and relative humidity, oviposition usually begins a few days later and continues for several days. The female dies soon after wards although males remain attached to the host for longer periods and may mate with other females. The number of females attached to the host is thus the most significant indicator of the seasonal activity and population dynamics of these ticks.

Designing an economical, integrated tick control strategy for a particular production system in a specific area are chemical control, ecological control biological control, genetic control, immunization.

Therefore based on the above conclusion the following recommendations are forwarded:

- Strategic tick control based on the biology, ecology aspect should be conducted
- Other safe control methods such as immunization control with other must be implemented
- Identification of tick species must be carried out
- Detailed epidemiology of ticks should be studied
- Movement of animals should be restricted

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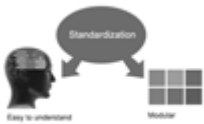
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INDEX

A

Acaricides · 8, 9, X, XI
Albino · 14, 16
Alexopolous · 7, 12
Anoniamuricata · 11
Aspergillusflavus · 11

B

Babesiosis · 6
Biguanides · 14
Bulbous · 1

C

Chelicerae · 2, 7

D

Debris · 5, 7
Diapause's · 4
Dioecious · 14, 2

E

Edematous · 7
Environs · 6
Eroded · 5

H

Hyaline · 7

I

Ichnotaxa · 1, 4
Ixodidae · 3, 2, 3

O

Osedacoides · 1, 2, 3, 4, 5
Osmoregulation · 7

P

Perennial · 6, 14, 17, 18, 25
Plesiosaur · 4

R

Rhipicephalus · 2, 6

S

Sclerotium · 6
Siboglinidae · 2, 5
Solanaceae · 6

T

Telotropic · 3

V

Vitellaria · 22, 23, 24, 25



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