



Synthesis, Crystal Structure, Hirshfeld Surface, Energy Framework and Molecular Docking of 2-(((6-Methoxy Pyridin-3-Yl) Imino) Methyl)Phenol

By M. Singh, S. Anthal, P. Akhileshwari, M. A. Sridhar, H. M. Vinusha, S. Bindya, M. Begum, R. K. Chandrasekaran, M. Saminathan & R. Kant

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Synthesis, Crystal Structure, Hirshfeld Surface, Energy Framework and Molecular Docking of 2-(((6-Methoxy Pyridin-3-Yl)Imino)Methyl)Phenol

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Abstract- The structure of 2-(((6-methoxypyridin-3-yl)imino)methyl)phenol (MPIMP) ($C_{13}H_{12}N_2O_2$) has been determined by X-ray diffraction methods. It crystallizes in the tetragonal crystal system with space group $P4_2/n$ and unit cell dimensions $a = 14.2958(3)$ Å, $b = 14.2958(3)$ Å, $c = 11.0179(3)$ Å, $V = 2251.73(12)$ Å³, $Z = 8$. The structure has been refined by full-matrix least square procedure to a final R -value of 0.0518 ($wR_2 = 0.1312$) for 1709 observed reflections. The molecules linked via two intermolecular (C-H...N and C-H...O) hydrogen bonds. The crystal structure was further stabilized by a strong intramolecular N-H...O hydrogen bond. The Hirshfeld surface analysis reveals the interaction contacts of the molecule and the strength of molecular packing in the crystal. The energy framework has been performed through different intermolecular interaction energies for structural stability. The molecular docking of MPIMP was performed against tuberculosis enzyme Decaprenylphosphoryl-b-Dribose 20-epimerase (DprE1, PDB code: 4KW5) to reconnoiter the binding interactions at the active sites.

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I. INTRODUCTION

The pyridine Schiff bases form one of the most significant classes of chemical compounds in organic chemistry and are generally characterized by the presence of pyridine and a phenolic ring connected by an azomethine group. The nitrogen present in the pyridine is usually responsible for the antifungal effects, while the phenolic ring participates in the bioactivity [1]. Schiff bases, being synthetically available and structurally varied, are obtained by replacing the carbonyl group ($>C=O$) present in the

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aldehyde or ketone with imine or azomethine group ($>C=N$) by reacting the carbonyl group with primary amines (-NH₂) [2].

As the Schiff bases with an imine group in its structure exhibit a wide range of potential for biological activities like antifungal, antibacterial, anticancer, antidepressant, anticonvulsant, anti-inflammatory, antiviral, antitumor, antioxidant, etc. [3-15], we present synthesis and single crystal X-ray (SCXR) structure and related spectroscopic characterization of a new pyridine Schiff base MPIMP. The SCXR molecular structure has been obtained by using direct methods, and Hirshfeld surfaces (HS) and Energy framework calculations have been performed by using Crystal Explorer (17.5) [16]. Since pyridine derivatives are capable of inhibiting Decaprenylphosphoryl-b-Dribose 20-epimerase (DprE1) [17, 18], *in silico* molecular docking of MPIMP has also been reported.

II. EXPERIMENTAL SECTION

All the chemicals of analytical grade required for our present research work were purchased from Sigma Aldrich and utilized without any purification.

The Schiff base, 2-(((6-methoxypyridin-3-yl)imino)methyl)phenol has been synthesized by condensation reaction between 6-methoxypyridin-3-amine with 2-hydroxybenzaldehyde. 6-methoxypyridin-3-amine (1.24g, 0.01mol) has been mixed with 2-hydroxybenzaldehyde (1.22g, 0.01mol) and 10 ml of methanol as solvent. The mixture was refluxed with 2-3 drops of glacial acetic acid at 60-70°C for 6hrs. The reaction progress was monitored by using TLC. The reaction mixture kept for overnight evaporation after the completion of the reaction. The reddish-brown colored crystals have been recrystallized using methanol. The Reaction scheme of the compound is presented in Fig. 1.

The intensity data of MPIMP were collected using Super Nova, Single source at offset/far, HyPix3000 diffractometer equipped with graphite monochromatic MoK_α radiation ($\lambda = 0.71073$ Å). Using Olex2 [19], the structure was solved using SHELXT [20] software routine with Intrinsic Phasing. Five cycles of full-matrix least-squares refinement have been performed, and the final



R-factor (0.0518) yielded the best possible atomic peaks. All non-hydrogen atoms of the molecule were located from the E-map and refined in anisotropic approximation using SHELXL [21]. The Crystallographic data are summarized in Table I. Hydrogen atoms bonded to C atoms were geometrically fixed and allowed to ride on the corresponding non-H atoms [$C-H = 0.93\text{-}0.96 \text{ \AA}$, and $U_{iso}(H) = 1.5 U_{eq}$ of the attached C atoms for methyl groups and $1.2 U_{eq}(C)$ for other H atoms]. Residual electron density ranges from -0.27 to 0.15 e.\AA^{-3} . The atomic scattering factors were taken from the International Tables for X-ray Crystallography (1992, Vol C, Tables 4.2.6.8 and 6.1.1.4). Olex2 [19], PLATON [22], PARST [23], and Mercury [24] defines the molecular geometry. The crystallographic information file (CIF) of the compound has been deposited at Cambridge Crystallographic Data centre (CCDC number 2091399). This CIF file can be accessed free of cost from Cambridge Crystallographic Data Center via <https://www.ccdc.cam.ac.uk/structures>.

The Hirshfeld surface (HS) mapped plots (d_{norm} , 2-D fingerprint, electrostatic potential, shape-index, and curvedness) and energy framework has been generated using Crystal-Explorer (version 17.5) with B3LYP function and 6-31G (d, p) basic set [16]. The atomic coordinates have been imported from the final validated CIF to Crystal Explorer (17.5). HS has been drawn for the asymmetric unit using high resolution of three-dimensional pictures of intermolecular close contacts in a crystal.

The molecular docking studies have been executed using AutoDock Vina software [25] which is a suite of automated docking tools (ADT). The target enzyme Decaprenylphosphoryl-*b*-Dribose 20-epimerase (4KW5) has been obtained from the protein data bank (www.rcsb.org/pdb). The coordinates of the grid center are fixed at $X = 6.47$, $Y = -8.61$, $Z = 39.07$, and grid sphere radius is 42. The conformational protein structure is modeled and visualized using Discovery Studio Visualizer [26].

III. RESULT AND DISCUSSION

The X-ray crystallographic analysis of MPIMP reveals its asymmetric unit consists of a benzene and pyridine ring. An ORTEP plot of the molecular structure is shown in Fig. 2 [24]. The bond distances, bond angles, and torsion angles between the non-H atoms are contained in Table II. All the bond distances and bond angles are within the normal range and are comparable with the related structures [27-28]. A C=N group seems to have a strong electron-withdrawing character in the azomethine group of the structure. Thus, the C1-O1 bond distance of $1.3490(17) \text{ \AA}$ is reliable with C-O single bond length; similarly, the C7-N1 distance of $1.2834(16)$ agrees well with the standard value [28]. The benzene and pyridine ring

systems [having a dihedral angle of $7.53 (5)^\circ$] are planar.

In the crystal structure, strong N-H...O intramolecular interactions are responsible for the stability of molecules within the unit cell. This intramolecular interaction leads to the formation of a virtual six-membered ring forming the S6 graph set motif [29]. In the crystal structure, there exist two intermolecular hydrogen bonds [C7-H7...N2 and C13-H13A...O1] that link the molecules into chains along the c-axis, as shown in Fig. 3 (Mercury) [24]. A summary of intra and inter-molecular hydrogen bonding are listed in Table III. The packing of molecules in the crystal viewed along the c-axis is visualized in Fig. 4, (PLATON) [22].

Hirshfeld surface (HS) analysis was carried out to explore the intermolecular interactions responsible for molecular packing in the crystal. Fig. 5(a) shows the 2D fingerprint plot of overall contribution from all the interactions to the total HS area, whereas Fig. 5 (a*) represents the HS over normalized contact distance d_{norm} ; where d_{norm} gives the distance between two atoms across the surface to the combined van der Waals radii of the atoms [30]. The d_{norm} value is either positive or negative, depending upon the longer or shorter intermolecular contacts than the van der Waals radii, respectively. The red-colored region represents shorter contacts with negative d_{norm} value, blue regions correspond to longer contacts with positive d_{norm} value, and the white region represent the distance of contacts is exactly equal to the van der Waals separation with a d_{norm} value of zero [31]. The three red spots shown on the HS represent the shortest hydrogen bond interactions due to strong C-H...O, C-H...N, and O-H...N contacts shorter than van der Waals radii as shown in Fig. 5(a*).

Fig. 5(b-f) represents the 2D fingerprint plots showing H-H, C-H/H-C, O-H/H-O, C-C, N-H/H-N contacts with the percentages contribution and (b*), (c*), (d*), (e*), (f*) represents the associated d_{norm} HS, respectively. These plots have been generated in the d_{norm} range -0.1466 \AA to 1.7068 \AA . The d_i and d_e on the plot are the distance from the HS to the nearest nuclei inside and outside the surface, respectively. For any given d_i and d_e pairs on the 2D plot, white color represents no occurrence, blue color shows some occurrence, and green indicates more frequent occurrence. These 2D fingerprint plots enable us to determine the percentage contribution of each type of contact to the total HS area. The major contribution to the total Hirshfeld area is H-H contacts with 50.9%, while the remaining significant contribution is from C-H/H-C (17.1%), O-H/H-O (12.8%), C-C (6.2%), and N-H/H-N (5.8%), as shown in Fig. 6. The d_i and d_e combination in the 2D fingerprint plot provides an outline of intermolecular contacts in the crystal, where one molecule acts as an acceptor ($d_e < d_i$) and the other as a donor ($d_e > d_i$). The O...H/H...O and

N...H/H...N molecular interactions represent by two distinct spikes in the bottom left and right region of almost equal length in 2D plots with $(d_i + d_e) \approx 2.50\text{\AA}$ and 2.45\AA , respectively, as shown in Fig. 5(d) and 5(f).

The Hirshfeld surface (HS) mapped over the calculated electrostatic potential for the title compound MPIMP as shown in Fig.7 (a). This mapping gives direct insight into the intermolecular interaction, which is responsible for molecular packing in the crystal [32]. The electrostatic potential map has been generated by using the B3LYP function with 6-31G (d) basis set [16]. The blue-colored area on the map is the electropositive region (i.e., the region around hydrogen bond donor) and the red-colored area is the electronegative region (i.e., the region around hydrogen bond acceptor). The electropositive blue region near C13-H13A atoms is complementary to the electronegative red region around the O1 atom, as shown in Fig. 7(a).

Fig. 7(b) shows the molecular HS mapped over the shape index for the compound. This mapping of shape index gives us the visual identification of the regions where the two molecular HS touch each other, and uses complementary pairs of red and blue colored schemes [30]. The concave red-colored region on the shape index represents the cluster of the surface around the acceptor atoms. In contrast, the blue-colored bumps regions represent the cluster of the surface around the donor atoms.

The curvedness map displays large regions of green (relatively flat) separated by dark blue boundaries (large positive curvatures), as shown in Fig.7(c). The plot shows no flat surface patches. Consequently, there is no indication of planner stacking between the molecules.

The energy frameworks give information on accurate inter-molecular interaction energies, which are responsible for the supramolecular construction of molecules in the crystal [33]. These calculations has been carried out using the Crystal Explorer (17.5) program based on 6-31G (d, p) basic set [16], where interaction energies have been calculated within a radius of 3.8\AA cluster around a single molecule of the title compound MPIMP. The scale factors used for the construction of energy framework for B3LYP/6-31G (d, p) electron densities are $k_{\text{ele}} = 1.057$, $k_{\text{pol}} = 0.740$, $k_{\text{disp}} = 0.871$, $k_{\text{rep}} = 0.618$ [34], respectively. Table IV contains the interaction energies, viz. electrostatic, polarization, dispersion, and repulsive energies. Fig. 8 represents the molecular pairs in which few molecules are separated by respective radial distance (as shown in Table IV) from the centroid molecule involved in the calculation of interaction energies along the c-axis.

The images of different interactions energies – coulomb interaction energy (red), dispersion energy (green), and total energy (blue) of the title compound MPIMP along a, b, and c-axes are illustrated in Fig. 9. The cylinders represent the energies between the

molecular pairs joining with the center of mass of the molecules, and the radius of the cylinder is proportional to the magnitude of the interaction energy. The framework of the cylinders represents the relative strengths of molecular packing in different directions. An overall scale factor is applied to expand or contract the size of the cylinders in the framework [33]. To avoid very crowded structures, weaker interactions have been omitted below a certain threshold energy, due to which there is an absence of the cylinders in a particular direction. The calculated energies for electrostatic, polarization, dispersion, and repulsion are -34.5 KJ/mol , -8.5 KJ/mol , -170.1 KJ/mol , and 94.9 KJ/mol , respectively. The total interaction energy is -132.5 KJ/mol . The dispersion energy dominates over the electrostatics Coulomb energy in the title compound MPIMP.

The molecular docking has done performed against DprE1(PDB code: 4KW5), and its resolve is to calculate the binding modalities of MPIMP. Since the hydrogen bond is an essential key factor in the structure and function of biological compounds, the ligand-receptor interactions have been inspected based on the hydrogen bonding. The molecular binding site of compound MPIMP and pyrazinamide interaction into DprE1 enzyme is present in Fig.10. The binding energy, distance and bonding type of compound MPIMP, and standard drug pyrazinamide with DprE1 are listed in Table V. In Compound MPIMP-4KW5 complex, the oxygen (O) atom of MPIMP has branched H-bond polar interaction with the atom NE2 and OE1 of residue HIS123 and GLN336 at a distance 2.506 and 2.204\AA , respectively. Moreover, the oxygen atom of MPIMP interacts with atom HZ1 of LYS418 at a separation of 2.530\AA . Three hydrophobic Alkyl- π interactions can be noticed between atoms of residue (CYS387, VAL365 & PRO116) and rings of compound MPIMP.

In Pyrazinamide -4KW5 complex, the oxygen (O) atom of pyrazinamide has interaction with the atom O of residue ALA53 at a distance of 2.027\AA , and the hydrogen (H) atom of pyrazinamide has interaction with the atom N of residue GLY55 at a distance of 2.262\AA . Moreover, the ring of pyrazinamide Hydrophobic interacts with atom N of GLY125 and atom CD of ALA128 at a separation of 3.800 and 4.847\AA , respectively. The outcome predicts that the compound MPIMP could strongly fix in the active site of DprE1, with binding energy of 7.4 kcal/mol , compared to 5.4 kcal/mol of pyrazinamide.

IV. CONCLUSIONS

2-(((6-methoxypyridin-3-yl)imino)methyl)phenol crystallizes in tetragonal crystal system with $P4_2/n$ space group. The crystallographic analysis shows the presence of different inter and intramolecular interactions, which aids the crystal packing. The

Hirshfeld surface analysis has been carried out to understand the intermolecular interaction contacts and the percentage contribution of each type of contact. The major contribution is from H-H contacts with 50.9%. The energy framework has also been carried out to calculate the different interaction energies viz., electrostatic, dispersion, polarization, and repulsion between the molecular pairs in the crystal, where dispersion energy was the dominant value among all the interaction energies. The molecular docking analysis has been performed. The molecular docking results recommend

that the compound MPIMP might exhibit strong inhibitory activity against DprE1 enzymes compare to the available pyrazinamide drug. It may result in the development of the new antituberculosis drug.

ACKNOWLEDGMENTS

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Table I: Crystal data and experimental details

Empirical formula	C ₁₃ H ₁₂ N ₂ O ₂
CCDC No.	2091399
Formula weight	228.25
Temperature (K)	293(2)
Crystal system	Tetragonal
Space group	P4 ₂ /n
a (Å)	14.2958 (3)
b (Å)	14.2958 (3)
c (Å)	11.0179 (3)
α (°)	90
β (°)	90
γ (°)	90
Volume (Å ³)	2251.73 (12)
Z	8
ρ calc (g/cm ³)	1.347
μ (mm ⁻¹)	0.093
F(000)	960.0
Crystal size (mm ³)	0.12 × 0.08 × 0.06
Radiation	MoKα (λ = 0.71073 Å)
2Θ range for data collection (°)	6.794 to 54.774
Index ranges	-17 ≤ h ≤ 17, -17 ≤ k ≤ 16, -13 ≤ l ≤ 14
Reflections collected	17000
Independent reflections	2430 [Rint = 0.0311, Rsigma = 0.0227]
Data / restraints / parameters	2430/0/156
Goodness-of-fit on F ²	1.059
Final R indexes [l ≥ 2σ (l)]	R1 = 0.0518, wR2 = 0.1312
Final R indexes [all data]	R1 = 0.0723, wR2 = 0.1456
Largest diff. peak /hole (e.Å ⁻³)	0.15/-0.27

Table II: Bond distances (Å), Bond angles (°) and Torsion angles (°)

Bond	Distance (Å)	Bond	Distance (Å)
O2-C10	1.3454(17)	C6-C7	1.4401(19)
O2-C13	1.4326(17)	C6-C1	1.4053(19)
O1-C1	1.3490(17)	C6-C5	1.3997(19)
N1-C8	1.4122(18)	C10-C11	1.3934(19)
N1-C7	1.2834(16)	C1-C2	1.380(2)
N2-C10	1.3130(16)	C5-C4	1.364(2)
N2-C9	1.3394(17)	C12-C11	1.3611(19)
C8-C9	1.3715(18)	C4-C3	1.384(2)
C8-C12	1.3967(18)	C3-C2	1.377(2)
Bond	Angles (°)	Bond	Angles (°)
C10-O2-C13	117.21(11)	N1-C7-C6	122.52(12)
C7-N1-C8	121.51(11)	O1-C1-C6	120.94(14)
C10-N2-C9	116.74(11)	O1-C1-C2	119.33(13)
C9-C8-N1	117.25(11)	C2-C1-C6	119.73(13)
C9-C8-C12	116.60(13)	N2-C9-C8	125.05(12)

C12-C8-N1	126.14(11)	C4-C5-C6	121.65(13)
C1-C6-C7	121.97(12)	C11-C12-C8	119.55(12)
C5-C6-C7	119.82(12)	C12-C11-C10	118.66(12)
C5-C6-C1	118.21(13)	C5-C4-C3	119.34(14)
O2-C10-C11	116.88(11)	C2-C3-C4	120.51(15)
N2-C10-O2	119.74(12)	C3-C2-C1	120.56(15)
N2-C10-C11	123.38(14)		
Bond	Torsion Angles (°)	Bond	Torsion Angles (°)
C13-O2-C10-N2	-2.66 (19)	C7-C6-C1-O1	0.86 (21)
C13-O2-C10-C11	176.99 (13)	C7-C6-C1-C2	-178.90 (14)
C7-N1-C8-C9	172.36 (13)	C5-C6-C1-O1	-179.45 (13)
C7-N1-C8-C12	-8.87 (20)	C5-C6-C1-C2	0.79 (21)
C8-N1-C7-C6	178.37 (12)	C7-C6-C5-C4	178.79 (14)
C9-N2-C10-O2	179.26 (12)	C1-C6-C5-C4	-0.91 (21)
C9-N2-C10-C11	-0.36 (20)	O2-C10-C11-C12	-178.76 (13)
C10-N2-C9-C8	-0.57 (21)	N2-C10-C11-C12	0.88 (22)
N1-C8-C9-N2	179.81 (13)	O1 -C1-C2-C3	-179.97 (14)
C12-C8-C9-N2	0.92 (21)	C6- C1-C2-C3	-0.21 (23)
N1-C8-C12-C11	-179.12 (13)	C6-C5-C4-C3	0.43 (23)
C9-C8-C12-C11	-0.34 (20)	C8-C12-C11-C10	-0.48 (21)
C1-C6-C7-N1	1.94 (21)	C5-C4-C3-C2	0.19 (24)
C5-C6-C7-N1	-177.75 (13)	C4-C3-C2-C1	-0.29 (25)

Table III: Hydrogen bonding geometry (e.s.d.'s in parentheses)

D-H...A	D-H(Å)	H...A(Å)	D...A(Å)	D-H...A(°)
O1-H1...N1	0.82	1.88	2.6137(15)	148
C7-H7...N2 ⁽ⁱ⁾	0.93	2.57	3.4273(17)	153
C13- H13A...O1 ⁽ⁱⁱ⁾	0.96	2.58	3.384(2)	141

Symmetry codes: (i) 1/2 - x, 3/2 - y, z (ii) 1 - x, -1/2 + y, 1/2 + z

Table IV: Different interaction energies of the molecular pairs in KJ/mol.

N	Symmetry operation	Radial Distance	Electron Density	E_ele	E_pol	E_dis	E_rep	E_tot
1	-x+1/2, -y+1/2, z	4.46	B3LYP/6-31G(d,p)	-7.7	-2.0	-54.8	28.5	-39.8
1	-x, -y, -z	5.77	B3LYP/6-31G(d,p)	-5.7	-1.5	-48.3	19.7	-37.0
2	-y+1/2, x, -z+1/2	9.32	B3LYP/6-31G(d,p)	-4.5	-0.6	-10.4	6.1	-10.5
1	-x, -y, -z	10.34	B3LYP/6-31G(d,p)	-0.6	-0.3	-17.8	8.1	-11.3
2	y+1/2, -x, z+1/2	6.35	B3LYP/6-31G(d,p)	-14.2	-3.1	-24.1	24.3	-23.3
2	-y, x+1/2, z+1/2	6.35	B3LYP/6-31G(d,p)	-1.8	-1.0	-14.7	7.7	-10.6

Table V: Binding energy, hydrogen bond and hydrophobic contacts of 2-Methoxy 4-(2-hydroxybenzylidene) amino pyridine (MPIMP) and pyrazinamide with 4WK5.

Inhibitor	Binding Energy (Kcal m ⁻¹)	Interactions	Distance Å	Bonding	Bonding Types	Binding Site of Protein	Binding Site of Ligand
MPIMP	7.4	HIS123[NE2...H-O] GLN336[OE1...H-O] LYS418[HZ1...O] VAL365[VA...π] PRO116[PR... π] CYS387[CY... π]	2.506 2.204 2.530 4.833 5.445 5.019	Hydrogen Hydrogen Hydrogen Hydrophobic Hydrophobic Hydrophobic	H-bond H-bond H-bond Alkyl- π Alkyl- π Alkyl- π	NE2 OE1 HZ1 VA PR CY	O O O Methoxy ring Pyridine ring Methoxy Ring
pyrazinamide	5.4	ALA53144[O...H-O] GLY55[N-H...O] GLY125[N...π] ALA128[CD...π]	2.027 2.262 3.800 4.847	Hydrogen Hydrogen Hydrophobic Hydrophobic	H-bond H-bond Amide-π Alkyl-π	O N N CD	O O Ring Ring

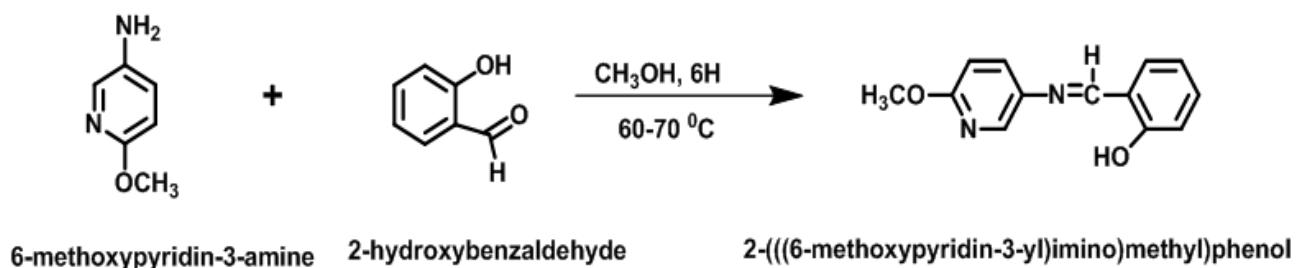


Fig. 1: Reaction scheme of 2-(((6-methoxypyridin-3-yl)imino)methyl)phenol.

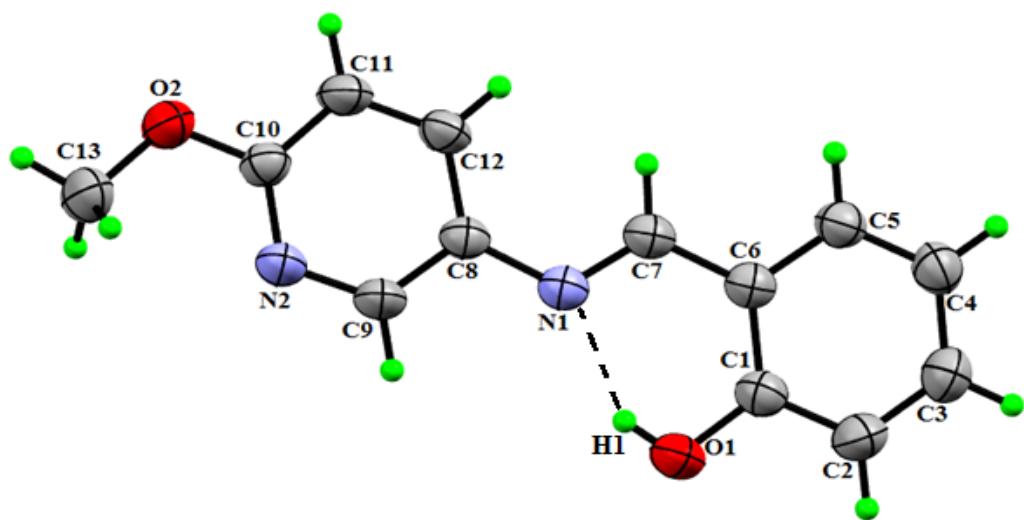


Fig. 2: ORTEP view of molecule with displacement ellipsoids drawn at 40% probability level (H atoms are shown as small arbitrary radius). The dotted line shows the intramolecular hydrogen bond.

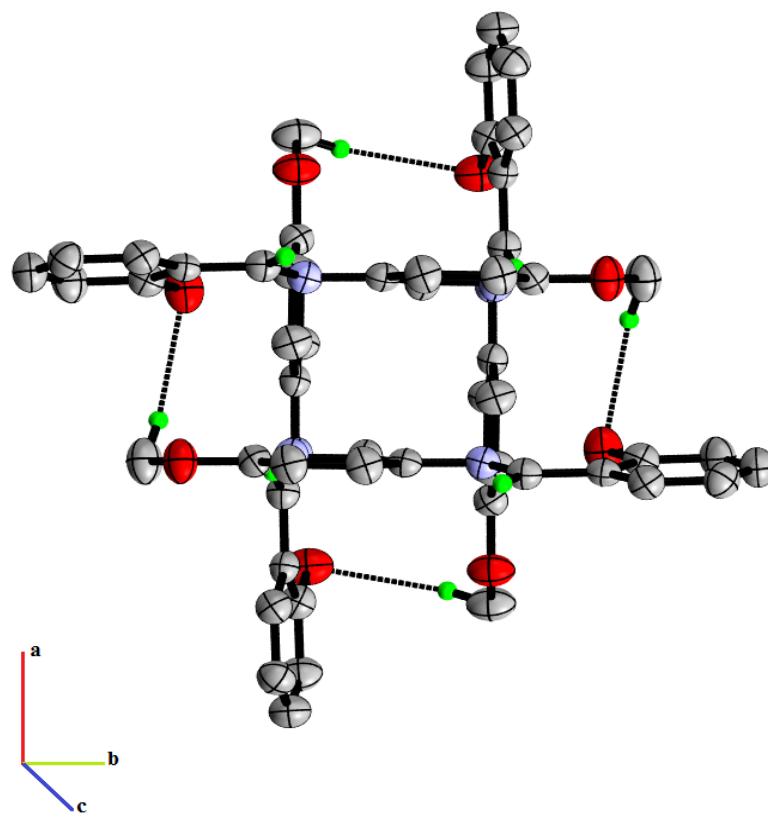


Fig. 3: Packing of crystal structure view along c-axis (dashed lines represents H-bonded interactions)

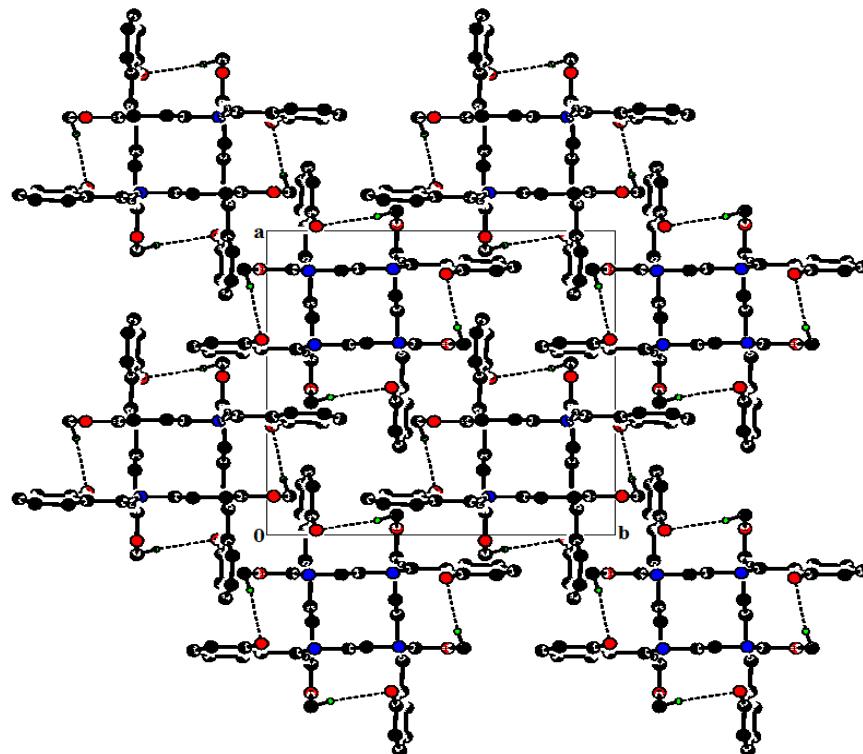
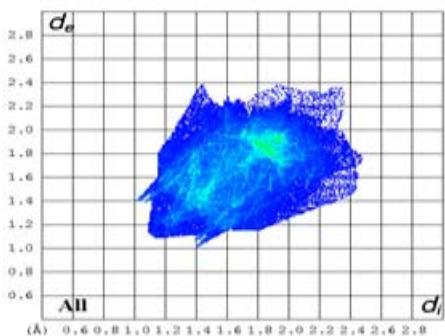
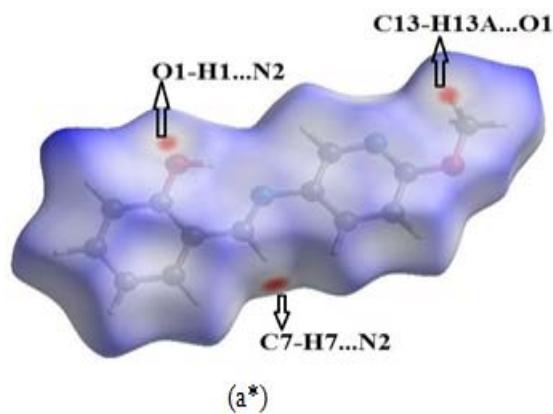


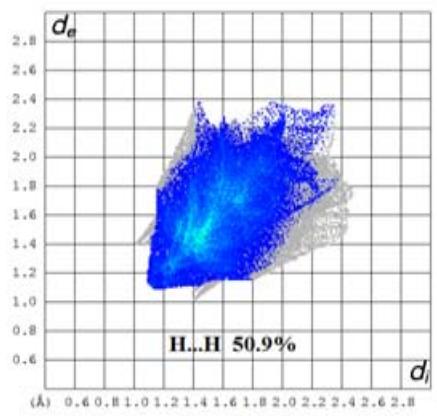
Fig. 4: Packing of crystal structure view along c-axis (dashed lines represents H-bonded interactions).



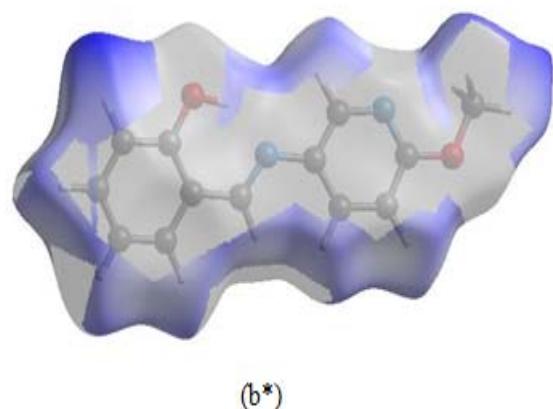
(a)



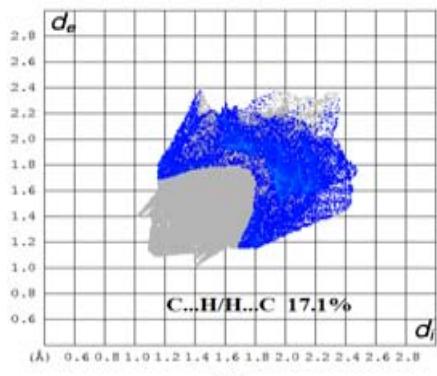
(a*)



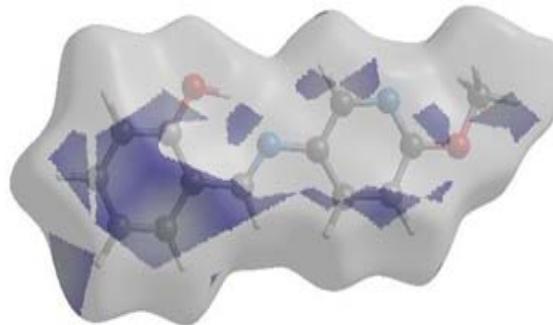
(b)



(b*)



(c)



(c*)

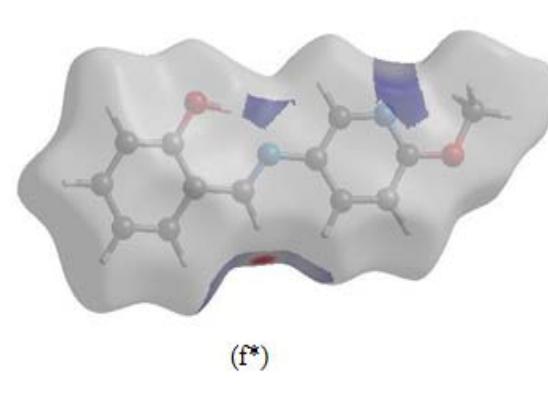
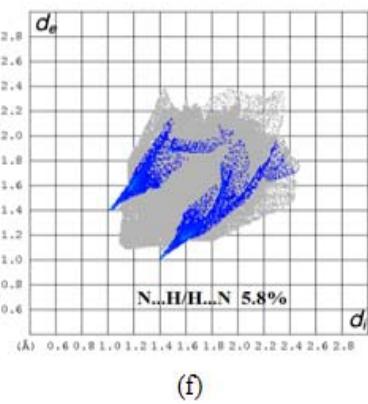
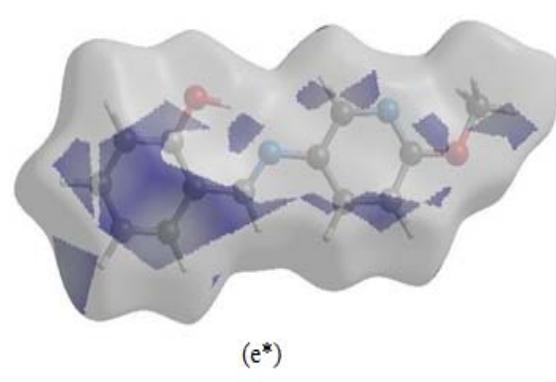
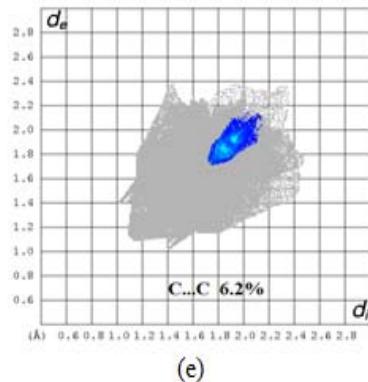
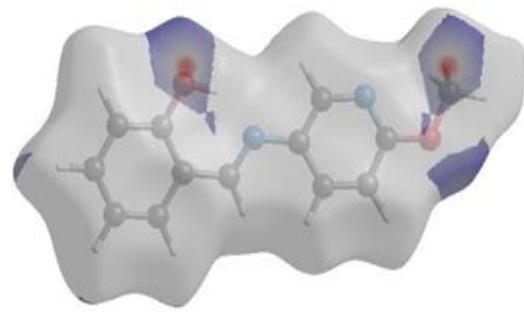
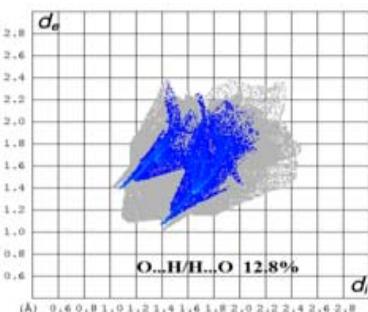


Fig. 5: Fingerprint plots of the compound from all the intermolecular contacts, (a*) Hirshfeld surface mapped over d_{norm} , (b), (c), (d), (e) and (f) are the 2D fingerprint plots showing H-H, C-H/H-C, O-H/H-O, C-C, N-H/H-N contacts with the percentages contribution and (b*), (c*), (d*), (e*) and (f*) represent the associated d_{norm} Hirshfeld surfaces respectively

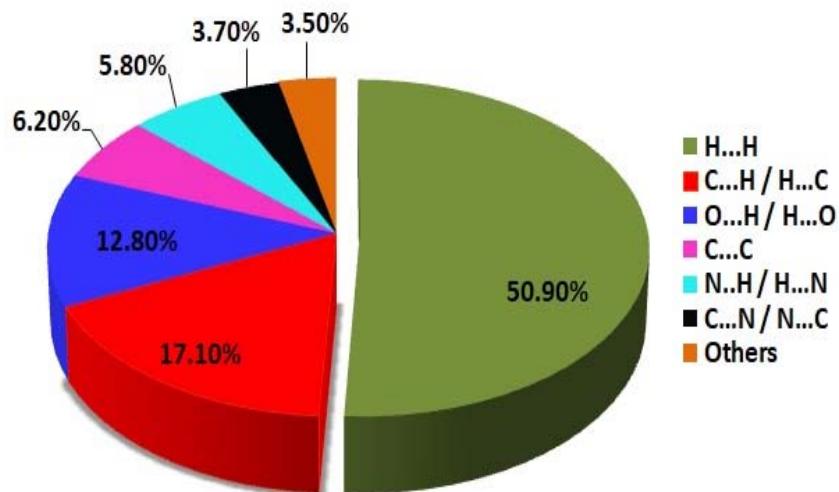


Fig. 6: Relative contributions of various intermolecular interactions to the Hirshfeld surface area.

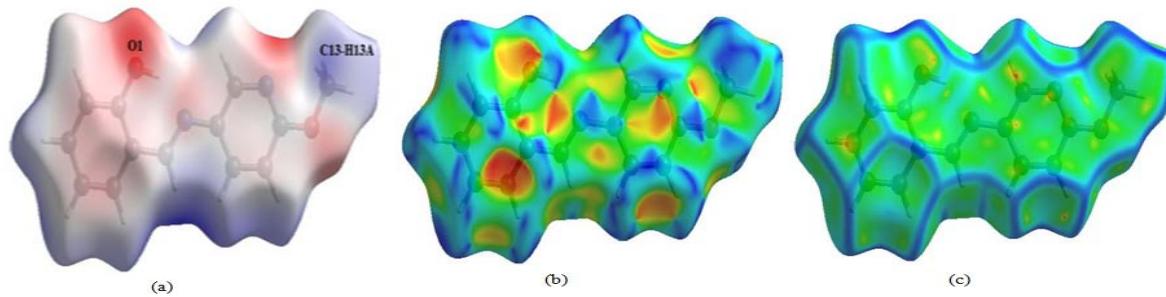


Fig. 7.(a): Electrostatic potential map (b) Shape index mapped on Hirshfeld surface (c) Curvedness plot mapped on Hirshfeld surface

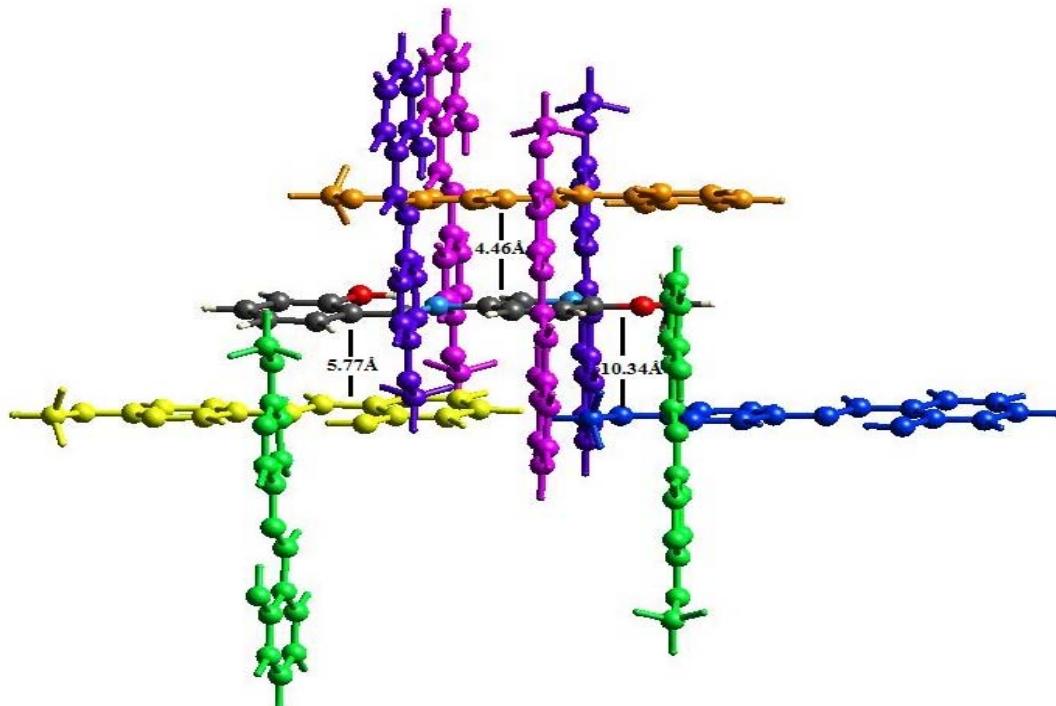


Fig. 8: Molecular pairs separated by respective radial distance from the centriod molecule involved in the calculation of interaction energies along c-axis

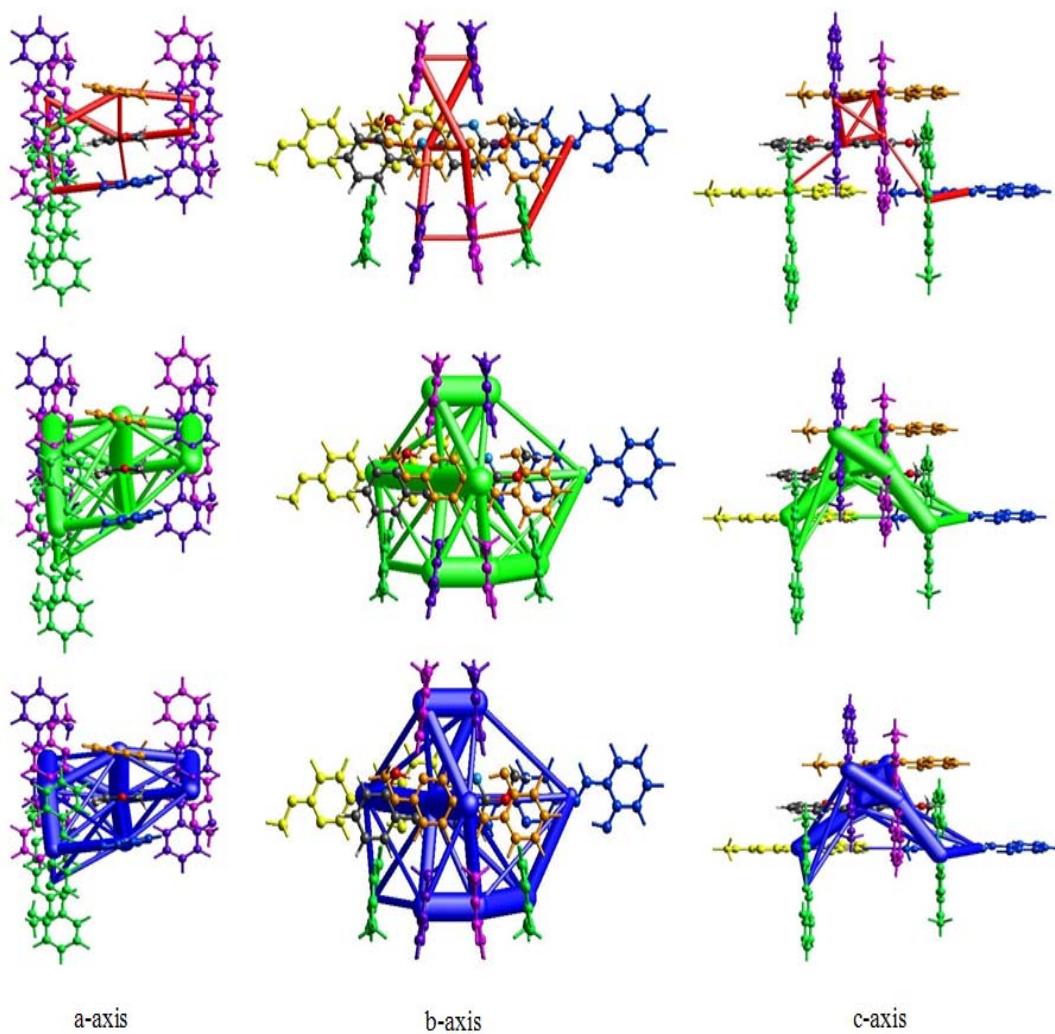


Fig. 9: The graphical representation of electrostatic interactions viz. coulomb interaction energy (red), dispersion energy (green), total interaction energy (blue) of the compound along different axes

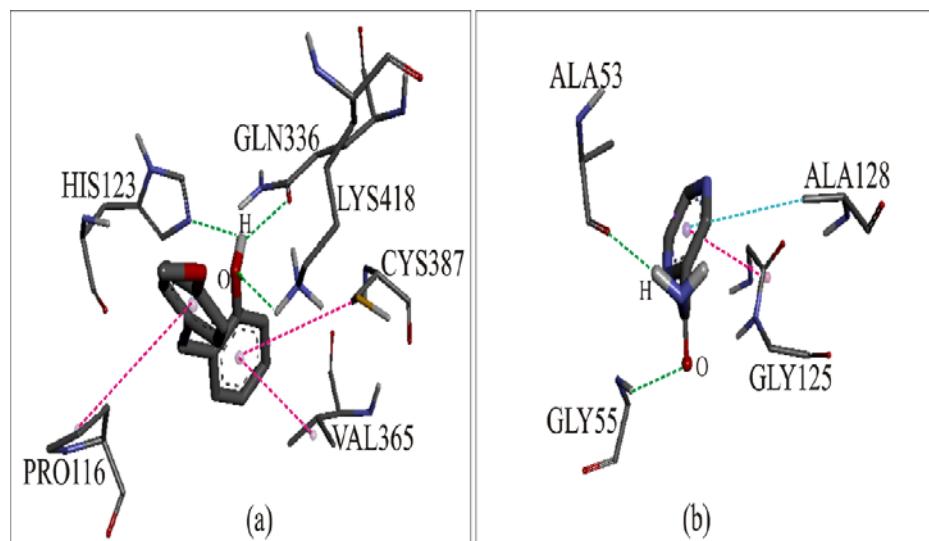


Fig. 10: Interaction of (a) MPIMP and (b) pyrazinamide to 4KW5 binding site

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