Epigenetics Theoretical Limits of Synthetic Genomes: the cases of Artificials *Caulobacter* (*C. eth-2.0*), *Mycoplasma Mycoides* (*JCVI-Syn 1.0, JCVI-Syn 3.0 and JCVI_3A*), *E-coli* and *YEAST chr XII*

By Jean-claude Perez

Abstract- In (Venetz et al., 2019), authors rebuilt the essential genome of *Caulobacter crescentus* through the process of chemical synthesis rewriting and studied the genetic information content at the level of its essential genes. Then, they reduced the native *Caulobacter crescentus* native *Caulobacter* NA1000 genome sequence real genome (4042929 bp) to the 785,701-bp reduced synthetic genome. Here we demonstrate the existence of a palindromic-like mirror structure that exists in real genomes and disappears totally in the synthetic genome. This biomathematic meta-organization is based on characteristic proportions of Fibonacci numbers between DNA single strand nucleotides proportions TC/AG on the one hand and TG/AC on the other hand. In both cases, we suggest that this meta-structure enhances the three-dimensional cohesion of the two DNA strands of the genome. We then generalize this study to the different synthetic genomes and synthetic cells published by the Craig Venter Institute on *Mycoplasma Mycoides* JCVI-syn1.0 (in 2010), JCVI-syn3.0 (in 2016) and JCVI-syn3A (in 2019). Finally, in the discussion section, we extend this study to synthetic genomes of *E-Coli* and *Yeast chromosome XII.*

Keywords: synthetic genomes, epigenetics, transposons, biomathematics, fibonacci numbers.

GJSFR-G Classification: DDC Code: 547.2 LCC Code: QD262

EPIGENETICSTHEORETICALLIMITSOFSYNTHETICGENOMESTHECASESOFARTIFICIALSCAULOBACTERCETH2OMYCOPLASMAMYCOIDESJCVISYN1OJCVISYN3OANDJCVI3AECOLIANDYEASTCHRXII

Strictly as per the compliance and regulations of:

Epigenetics Theoretical Limits of Synthetic Genomes: The Cases of Artificials *Caulobacter (C. eth-2.0), Mycoplasma Mycoides (JCVI-Syn 1.0, JCVI-Syn 3.0 and JCVI_3A), E-coli and YEAST chr XII*

Solved by standard Gammas, unvarying Deltas, uniform Epsilons. Millions of identical twins. The principle of mass production at last applied to biology
in « BRAVE NEW WORLD » Chapter 1, Aldous Huxley 1931
https://www.huxley.net/bnw/one.html

Jean-claude Perez

Abstract- In (Venetz et al., 2019), authors rebuilt the essential genome of *Caulobacter crescentus* through the process of chemical synthesis rewriting and studied the genetic information content at the level of its essential genes. Then, they reduced the native *Caulobacter crescentus* native *Caulobacter* NA1000 genome sequence real genome (4042929 bp) to the 785,701-bp reduced synthetic genome. Here we demonstrate the existence of a palindromic-like mirror structure that exists in real genomes and disappears totally in the synthetic genome. This biomathematic meta-organization is based on characteristic proportions of Fibonacci numbers between DNA single strand nucleotides proportions TC/AG on the one hand and TG/AC on the other hand. In both cases, we suggest that this meta-structure enhances the three-dimensional cohesion of the two DNA strands of the genome. We then generalize this study to the different synthetic genomes and synthetic cells published by the Craig Venter Institute on *Mycoplasma Mycoides* JCVI-syn1.0 (in 2010), JCVI-syn3.0 (in 2016) and JCVI-syn3A (in 2019). Finally, in the discussion section, we extend this study to synthetic genomes of *E-Coli* and *Yeast chromosome XII*.

Keywords: synthetic genomes, epigenetics, transposons, biomathematics, fibonacci numbers.

I. Introduction

The story which led to the development of the first synthetic genome JCVI-syn1.0 has its origins as far back as 1995, when Venter and his team published the sequence of *Mycoplasma genitalium* (Fraser, 1995) and (Sleator, 2010). In 2010, a 1079-kb genome based on the genome of *Mycoplasma mycoides* (JCV-syn1.0) was chemically synthesized and supported cell growth

Author: Retired interdisciplinary researcher (IBM Artificial Intelligence European Research Center, Montpellier France), Bordeaux metropole France, Luc MONTAGNIER Foundation Scientific Council, Quai Gustave-Ador 62 1207 Geneve Switzerland. e-mail: jeanclaudeperez2@gmail.com

when transplanted into cytoplasm. (Gibson, 2010). In 2016, Hutchinson et al design, build, and test cycle to reduce this *Mycoplasma mycoides* genome to 531 kb (473 genes). JCV-syn3.0 retains genes involved in key processes such as transcription and translation, but also contains 149 genes of unknown function. Since 2012 the Synthetic Yeast Genome Project (Sc2.0 http://synthetic yeast.org/sc2-0/) results from a worldwide partnership, « Sc2.0 International Consortium team », members spanning 4 continents to provide remote mentorship and solve challenges associated with synthetic individual chromosome design features and assembly (Jee Loon Foo 2018). Read the analysis in §Discussion. In January 2019, Breuer et al. published a synthetic cell resulting from the synthetic genome JCVI-syn3A, a robust minimal cell with a 543 kbp genome and 493 genes, providing a versatile platform to study the basics of life. Simultaneously, in 2019, Venetz et al. reduced the native Caulobacter crescentus NA1000 genome sequence real genome (4042929 bp) to the 785,701-bp reduced synthetic genome Caulobacter ethensis-2.0 (C. eth-2.0). Finally, also in 2019 (Fredens, 2019), researchers published a synthetic genome of E COLI changing systematically genetic code equivalent codons. They replaced every occurrence of the serine codon TCG with AGC, every TCA (also serine) with AGT, and every TAG (stop) with TAA. Read the analysis in § Discussion.

In a completely different field, 30 years ago, we had just published the first 2 French books on Artificial Intelligence (AI) neural networks (Perez, 1988; Perez, 1989; Perez, 1990a). It is the exploration of our network FRACTAL CHAOS (Perez 1990c), (Pellionisz et al, 2012), (Perez§Montagnier, 2021), which will reveal a hyper-sensitivity of this network to successive ratios of

Fibonacci numbers, for example 34/21 (Perez, 1990b). While the big project of sequencing of the human genome "HUGO" just begins, we have the intuition to look for ratios of Fibonacci numbers between the contiguous proportions of TCAG nucleotides of genes and small genomes available at that time (like HIV, mtDNA, viruses, bacteria, or small genes). We published a first article in 1991 (Perez, 1991; Marcer, 1992) demonstrating the evidence of such biomathematic structures (Perez, 1991). This discovery was completely published 22 years ago in the book "DNA decrypted" (Perez, 1997). This method, which the Nobel prize winner Luc Montagnier called "DNA supracode" (Fleaux, 1995), was used to search exaustively in DNA searched exhaustively in DNA sequences for remarkable proportions of Fibonacci numbers (https://en.wikipedia.org/wiki/Fibonacci_number) between nucleotides called "resonances": for example if a contiguous sequence of 377 bases TCAG is subdivided into 233 (C + A) and 144 (T + G), there is a resonance of CA / TG of length 377 (where 144, 233 and 377 are three Fibonacci numbers). In (Perez, 2017a), it is precisely such resonances CA / TG that characterize this optimality of the mtDNA genome of humans. It is still such resonances that are affected during mutations associated with cancers. In particular, we have analyzed this type of resonance in the 3 respective mtDNA genomes of humans, mice, and the famous naked mole rat as well as in more than a dozen other mammalian species.

In a comprehensive analysis of all (ALL) listed mutations of the human mitochondrial mtDNA genome associated with cancers : effectively, multiple mutations associated with the mitochondrial genome of tumor cells have been reported. An open question is whether these mutations are only the CONSEQUENCE of the cancer process or if, on the contrary, they would be a possible ORIGINAL CAUSE of the cancer genesis process. In a paper in preparation (Perez, 2019) we'll propose a generic and universal law (of a numerical nature) allowing us to detect and classify these mutations at the early stage of the genesis of the tumors. Finally, in (Perez 2019) we will present a generic law of prediction and classification of tumors by the simple analysis of the DNA SUPRACODE of the mitochondrial genomes associated with these tumors. In this upcoming article, we analyse all known somatic mutations listed all cancers combined. We then discover a global strategy of mutation of all these basic somatic mutations materialized by a numerical score which systematically increases in ALL the cases of elementary somatic mutations related to 91 referenced cases involved in 9 different cancers (prostate, pancreatic, colon, thyroid, bladder, breast, head § neck, meduloblastoma, ovarian) with a success rate of 100%. This predictive method should make it possible to categorize and classify the potential pathogenicity of tumors from the early stage.

Particularly, we find an interesting symmetric property of resonances with very short periods: for example, the resonances 3 (1 TC 2AG) and 3 (2TC 1AG) correspond to the symmetrical beginnings of the Fibonacci and Lucas sequences. Similarly, the resonances 5 (2 TC, 3AG) and (3TC 2AG) correspond to the symmetrical beginnings of the Fibonacci and FibLuc[1][2] sequences. By looking for these resonances in all the known tumor mutations of human mtDNA genomes applied to the genomes inherited by evolution of the RSRS mother sequence (EVE), it appears the functional role of such local resonances whose repercussion on the global scale of the genome becomes a indicator of early diagnosis of tumors.

It is this type of symmetry that we will generalize in this article by extending it to longer Fibonacci, and Lucas sequences.

Example 34 TCAG ==> 13 TC, 21 AG in one hand (regular)

and 34 TCAG ==> 21 TC, 13 AG in other hand (reverse).

II. Experimental Section

Part I: Genomes analysed

We will analyze 8 bacterial genomes, 3 real reference genomes, one transgenic genome, and four synthetic genomes.

==> *The 2 Caulobacter genomes:*

Name: NA1000 real

Reference: Caulobacter crescentus NA1000, complete genome

Publication: Venetz, 2019

Length: 4042929 bp

Access: native *Caulobacter* NA1000 genome sequence [National Center for Biotechnology Information (NCBI) accession no. NC_011916.1]

https://www.ncbi.nlm.nih.gov/nuccore/NC_011916.1

Name: Ethensis CETH 2.0

Reference: Synthetic Caulobacter sp. 'ethensis' strain CETH2.0 chromosome, complete genome

Publication; Venetz, 2019

Length: 785701 bp

Access:

https://www.ncbi.nlm.nih.gov/nuccore/CP035535

==> *The 6 Mycoplasma Mycoides genomes:*

---

[1] This sequence 3 2 5 7 12 19 31 50... results adding Fibonacci (1 2 3 5 8...) and Lucas (1 3 4 7 11...) sequences like : 2 (1+1), 5 (2+3), 7 (3+4), 12 (5+7)... Curiously, we discovered this sequence in (Perez, 2017b) resulting from stationary waves observed in DUF1220 repeat proteins in mammals brain coding DNA genomes.

[2] This sequence was also curiously used in new orleans jazz negro spiritual music (Parayon 2011).

Name: MycRef

Reference: Mycoplasma mycoides subsp. mycoides strain izsam_mm5713, complete genome

Publication: Orsini, 2015

Length: 1192498 bp

Access:
https://www.ncbi.nlm.nih.gov/nuccore/CP010267.1?report=genbank

Name: JCVI-syn1.0

Reference: Synthetic Mycoplasma mycoides JCVI-syn1.0 clone sMmYCp235-1, complete sequence
Publication: Gibson, 2010

Length: 1078809 bp

Access:
https://www.ncbi.nlm.nih.gov/nuccore/296455217

Name: Capritrans

Reference: Mycoplasma mycoides subsp. capri str. GM12 transgenic clone tetM-lacZ, complete genome

Publication: Direct Submission
JOURNAL Submitted (14-MAY-2009) The J. Craig Venter Institute, 9702 Medical
Center Drive, Rockville, MD 20850, USA

Length: 1089202 bp

Access:
https://www.ncbi.nlm.nih.gov/nuccore/CP001621.1

Name: Capri real

Reference: Mycoplasma mycoides subsp. mycoides SC str. PG1

Length: 1211703 bp

Access:
https://www.ncbi.nlm.nih.gov/nuccore/NC_005364.2

Name: JCVI-Syn3.0

Reference: Synthetic bacterium JCVI-Syn3.0, complete genome

Publication: Hutchinson, 2016

Length: 531490 bp

Access:
https://www.ncbi.nlm.nih.gov/nuccore/CP014940.1

Name: JCVI-Syn3A

Reference: Synthetic bacterium JCVI-Syn3A chromosome, complete genome

Publication: Breuer, 2019

Length: 543379 bp

Access:
https://www.ncbi.nlm.nih.gov/nuccore/CP016816.2

Part II: Computing DNA Supra Code Resonances:
Let us consider the 2 digital sequences :

Fibonacci: 1 1 2 3 5 8 13 21 34 55 89

Lucas: 2 1 3 4 7 11 18 29 47 76

For any contiguous sequence of nucleotides, one will search for "resonance" or exact proportions of the TG / CA types then mainly TC / AG.

For example, if 34 TCAG bases are subdivided exactly into 13 TC bases and 21 AG bases, we will consider having discovered a TC / AG resonance of length 34. We will do the same for the search for Lucas resonances. The whole genome is explored by taking each of the positions as successive exploration points. On the other hand, the genome being circular, the analysis from the last pivots at the end of the sequence is looped back to the positions of the start nucleotides.

We will thus search for 2 symmetrical types of resonances:

Main resonances (or forward): Exp. 34 TCAG ==> 13 TC, 21 AG.

Inverse Resonances (or backward): Exp. 34 TCAG ==> 21 TC, 13 AG.

For each length of Fibonacci (or Lucas) 3 5 8 13 21 34 55 89, we memorize the respective accumulations of the forward resonances on the one hand, and backward on the other hand.

It appears then that these 2 values are very close in the case of REAL genomes, whereas they are very different in the case of SYNTHETIC genomes.

We will therefore consider very significant: The forward / backward ratios. Forward-backward spreads. Since the lengths of real and synthetic genomes are generally very different, we will weight the forward-backward differences by the respective lengths of the real or synthetic genomes.

III. Results

We analyse here, in one hand, Caulobacter crescentus NA1000 genome and synthetic genome Caulobacter ethensis-2.0 (C. eth-2.0), and, in other hand, Mycoplasma Mycoides JCVI-syn1.0, JCVI-syn3.0 and JCVI-syn3A.

Part I: Caulobacter crescentus NA1000 genome and synthetic genome Caulobacter ethensis-2.0 (C. eth-2.0).

The actual NA1000 genome being about 5 times longer than the synthetic genome C. eth-2.0, one might think that the comparison of these 2 genomes is skewed. However, in all the above results, we had already incorporated this difference by weighting the results by the length of the respective genomes.

TC/AG analysis:

Nota: All tables in this article are identical: each box contains 4 numerical values: 1/The number "L" of Fibonacci or Lucas constituting the length of the sub-sequence analyzed. 2/The cumulated volume of the corresponding resonances (n x L) in regular exploration (forward). 3/The cumulative volume of the corresponding resonances (L x n) in reverse (backward) exploration. 4/The ratio of the 2 values below regular/reverse.

Table 1: TC/AG Fibonacci and Lucas analysis for real NA1000 genome and synthetic Caulobacter 2.0.

TC/AG Real genome NA1000				TC/AG Synthetic genome CAULOBACTER 2.0			
Fibonacci		Lucas		Fibonacci		Lucas	
3	1580254 1582201 0.9987694357	3	1582201 1580254 1.00123208	3	290047 293417 0.9885146396	3	293417 290047 1.011618807
5	1346353 1346512 0.9998819171	4	993497 994619 0.9988719299	5	239292 242875 0.9852475553	4	194318 198624 0.9783208474
8	924521 926003 0.9983995732	7	1186993 1186953 1.0000337	8	168546 172325 0.9780705063	7	209444 211890 0.9884562745
13	650861 652564 0.9973902943	11	661049 662784 0.9973822542	13	120565 124586 0.9677251055	11	123932 128203 0.966685647
21	377329 378580 0.9966955465	18	489878 492487 0.9947023982	21	74278 77737 0.9555038141	18	92499 96693 0.9566256089
34	188645 190539 0.9900597778	29	232050 233697 0.9929524127	34	40816 44246 0.9224788681	29	48895 52692 0.9279397252
55	71250 72070 0.9886221729	47	104152 104668 0.9950701265	55	18131 20974 0.8644512253	47	24743 27988 0.8840574532
89	19694 19535 1.008139237	76	30804 31161 0.9885433715	89	6300 8172 0.7709251101	76	9026 11226 0.8040263674
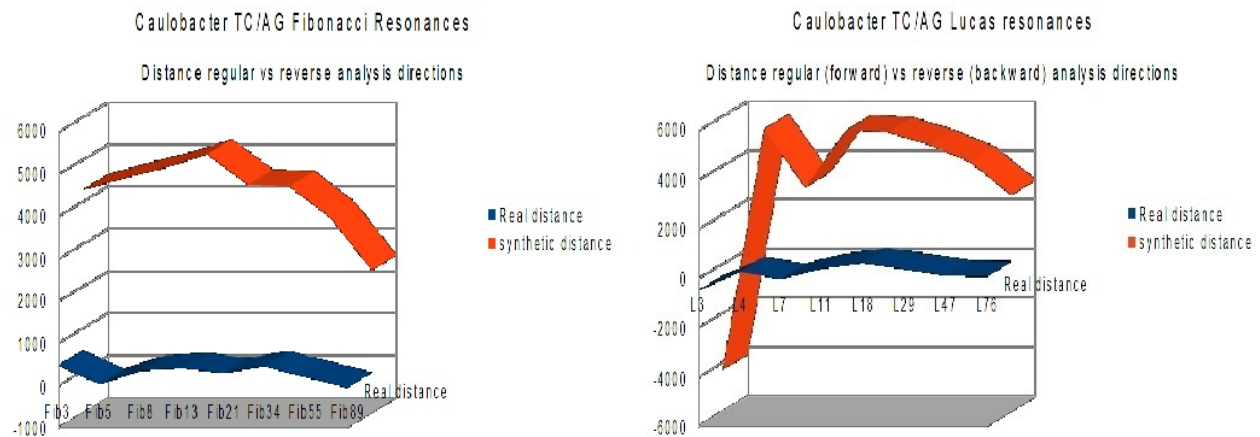
Figure 1: Comparing TC/AG Fibonacci and Lucas distances in real and synthetic Caulobacter genomes (regular-reverse distances weighted by the length of the genome, see detailed data in suplementary materials)

example of weighting by the length of the genome computing here the case of synthetic caulobacterium genome (case of the first Fibonacci resonance of length = 3 nucleotides):

regular - reverse distance = 290047-293417 = ⁻3370

weighting by the genome length : ⁻3370÷785750 = ⁻0.004288895959

normalization multiply by 1000000 = 1000000× ⁻0.004288895959 = ⁻4288.895959 = -4289

(see more details in suplementary materials page 2).

In the figure on the left, the average ratio of weighted distances by genome length between real genome and synthetic genome is 14.39 for TC/AG Fibonacci resonances (see supplementary materials).
For information, the same ratio related to Lucas TC/AG is = 14.345484

Computing details
real genome abs. Distances: 482   40   367   422   310   469   203   39
synthetic genome abs. Distances: 4289   4560   4810   5118   4403   4366   3619   2383
cumulating real genome abs. Distances: 2332

cumulating synthetic genome abs. Distances: 33548

Ratio synthetic genome abs. Distances / real genome abs. Distances = 14.38593482

TG/AC analysis

Table 2: TG/AC Fibonacci and Lucas analysis for real NA1000 genome and synthetic Caulobacter 2.0.

| TG/CA Real genome NA1000				TG/CA Synthetic genome CAULOBACTER 2.0			
Fibonacci		Lucas		Fibonacci		Lucas	
3	1607779 1614874 0.9956064684	3	1614874 1607779 1.00441292	3	296256 299993 0.9875430427	3	299993 296256 1.012614091
5	1349192 1358378 0.993237523	4	1011587 1022025 0.9897869426	5	244860 248873 0.9838753099	4	195156 199978 0.9758873476
8	915781 925976 0.9889899954	7	1174192 1180896 0.9943229548	8	169216 174299 0.9708374689	7	213326 216126 0.9870445944
13	642769 652548 0.9850141292	11	654997 665831 0.9837286038	13 120130 125243 959176		11	123819 128731 0.9618429127
21	384426 392873 0.9784994133	18	489376 497649 0.9833758332	21	73909 78870 0.9370990237	18	92670 96745 0.9578789602
34	209433 218299 0.9593859798	29	251102 259033 0.9693822795	34	42458 46179 0.9194222482	29	50145 53889 0.9305238546
55	91756 97158 0.9443998436	47	127067 133142 0.9543720239	55	20641 22427 0.9203638471	47	27305 29582 0.9230275167
89	31478 34303 0.917645687	76	45537 49636 0.9174188089	89	8188 9531 0.859091386	76	11334 12669 0.8946246744
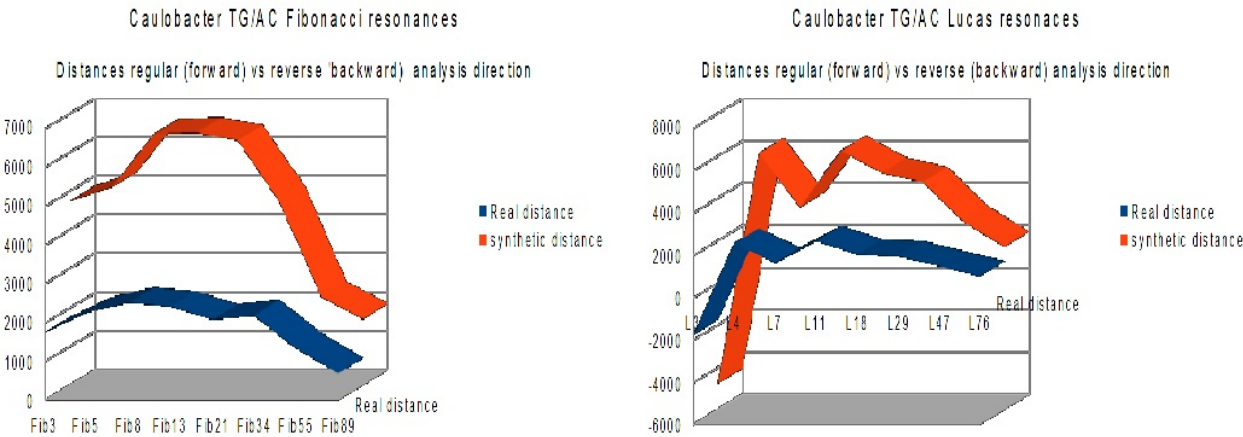
Figure 2: Comparing TG/AC Fibonacci and Lucas distances in real and synthetic Caulobacter genomes (regular-reverse distances weighted by the length of the genome, see detailed data in suplementary materials)

Part II: Mycoplasma Mycoides JCVI-syn1.0 (2010), JCVI-syn3.0 (2016) and JCVI-syn3A (2019)

In 2010, a 1079-kb genome based on the genome of *Mycoplasma mycoides* (JCV-syn1.0) was chemically synthesized and supported cell growth when transplanted into cytoplasm. (Gibson, 2010). In 2016, Hutchinson et al. design, build, and test cycle to reduce this *Mycoplasma mycoides* genome to 531 kb (473 genes). JCV-syn3.0 retains genes involved in key processes such as transcription and translation, but also contains 149 genes of unknown function. In the following section, we compare 6 (six) genomes : two reference real strain mycoplasma genomes including CAPRI strain, one transgenic building strain and the 3 strong JCV Labs ; synthetic genomes.

Table 3: Comparing TC/AG Fibonacci analysis for 6 real or synthetic Mycoplasm genomes

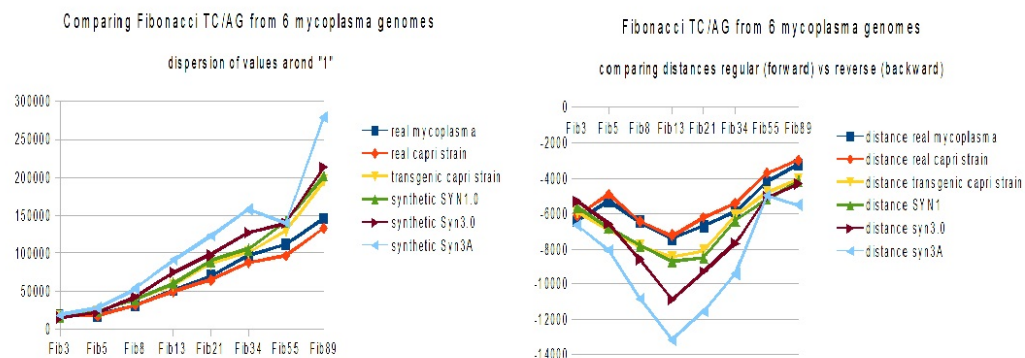| Mycoplasm REF real genomes			Synthetic Mycoplasm genomes		
Natural reference genomes		Transgenic genome	Synthetic genomes		
Reference real strain	Reference real strain CAPRI	Transgenic CAPRI strain	JCVI-Syn1.0	JCVI-Syn3.0	JCVI-Syn3A
3 425653 418009 1.01828 6688	3 431863 424453 1.017457 763	3 389604 383147 1.0168525 4	3 386328 380244 1.016000 252	3 191148 188330 1.014963 097	3 195794 192178 1.0188158 89
5 349081 342772 1.01840 5821	5 353999 348030 1.017150 82	5 322151 314644 1.0238587 1	5 319643 312321 1.023443 829	5 158678 155189 1.022482 264	5 162644 158262 1.0276882 64
8 249100 241395 1.03191 864	8 252787 244971 1.031905 817	8 228917 220387 1.0387046 42	8 227112 218687 1.038525 381	8 112973 108400 1.042186 347	8 116120 110250 1.0532426 3
13 182285 173428 1.05107 0185	13 184673 175894 1.049910 742	13 167319 158110 1.0582442 6	13 166119 156734 1.059878 52	13 83100 77329 1.074629 182	13 85559 78424 1.0909798 02
21 122345 114349 1.06992 6278	21 123850 116312 1.064808 446	21 111834 103016 1.0855983 54	21 110910 101748 1.090045 996	21 55296 50364 1.097927 091	21 57103 50837 1.1232566 83
34 80074 73025 1.09652 8586	34 81099 74515 1.088358 049	34 72890 66214 1.1008245 99	34 72247 65356 1.105437 909	34 36064 31989 1.127387 539	34 37460 32353 1.1578524 4
55 49907 44879 1.11203 4582	55 50665 46176 1.097215 003	55 45287 40085 1.1297742 3	55 44848 39272 1.141984 111	55 22000 19301 1.139837 314	55 22992 19368 1.1871127 63
89 30152 26319 1.14563 6232	89 30708 27108 1.132802 125	89 27153 22722 1.1950092 42	89 26742 22243 1.202265 881	89 13030 10741 1.213108 649	89 13749 10743 1.2798101 09

Figure 3: Left: Comparing TC/AG Fibonacci ratios from 6 mycoplasma genomes (relative values around 1), right: Comparing TC/AG Fibonacci distances from 6 mycoplasma genomes. (regular-reverse distances weighted by the length of the genome, see detailed data in suplementary materials)

In summary of this double analysis it seems obvious that synthetic genomes disturb and destroy a characteristic dimension of real genomes. This property could concern the mathematical topology of the genome (Rapoport, 2018) and probably its fractal, dynamic, evolution, and three-dimensional structures.

IV. Discussion

a) Comparing real E COLI Genome and synthetic changing TAG by TAA stop codons

In (Fredens et al., 2019), researchers published a synthetic genome of E COLI changing systematically genetic code equivalent codons. They replaced every occurrence of the serine codon TCG with AGC, every TCA (also serine amino acid) with AGT, and every TAG (stop codon) with TAA, for a total of 18,214 replacements. Here we run a sample comparison of TG

Fibonacci resonances changing stop codons TAG in TAA, then 7725 changes considering only TAG of the first codons reading frame ; In (Fredens et al., 2019), the sequences and genome design details used in this study are available in the Supplementary Data. Supplementary Data 1 provides the GenBank file of the *E. coli* MDS42 genome (NCBI accession number AP012306.1); Fredens's team systematically replaced every occurrence of the serine codon TCG with AGC, every TCA (also serine) with AGT, and every TAG (stop codon) with TAA, for a total of 18,214 replacements ; Not having access to the modified sequence of the synthetic genome yet, we simply changed all TAG codons to TAA codons, that is, 7,725 altered codons. We have limited this change to only the first reading frame codons.

Table 4: Comparing Fibonacci TG/AC from E-COLI real genome and E-Coli synthetic where all TAG codons are removed in TAA codons (1st codons reading frame only)

ECOLI reference wild type genome				ECOLI syn61 like where 7725 TAG ==> TAA			
3	1471002	1476399	0.9963444841	3	1458201	1484100	0.9825490196
5	1211718	1215554	0.9968442373	5	1204279	1221598	0.9858226683
8	852126	857586	0.9936332916	8	844110	865264	0.9755519703
13	612152	618151	0.9902952515	13	604631	625851	0.9660941662
21	385231	390203	0.9872579145	21	378106	397617	0.9509301665
34	222919	227478	0.9799585015	34	216935	233775	0.9279649235
55	107343	110152	0.9744988743	55	102252	114831	0.8904564099
89	43863	45199	0.9704418239	89	41531	47929	0.8665108807

Note, the TAG ==> TAA mutations (where G is mutated to A) does not affect the TC / AG structures, we have here to analyze the TG / AC structures.
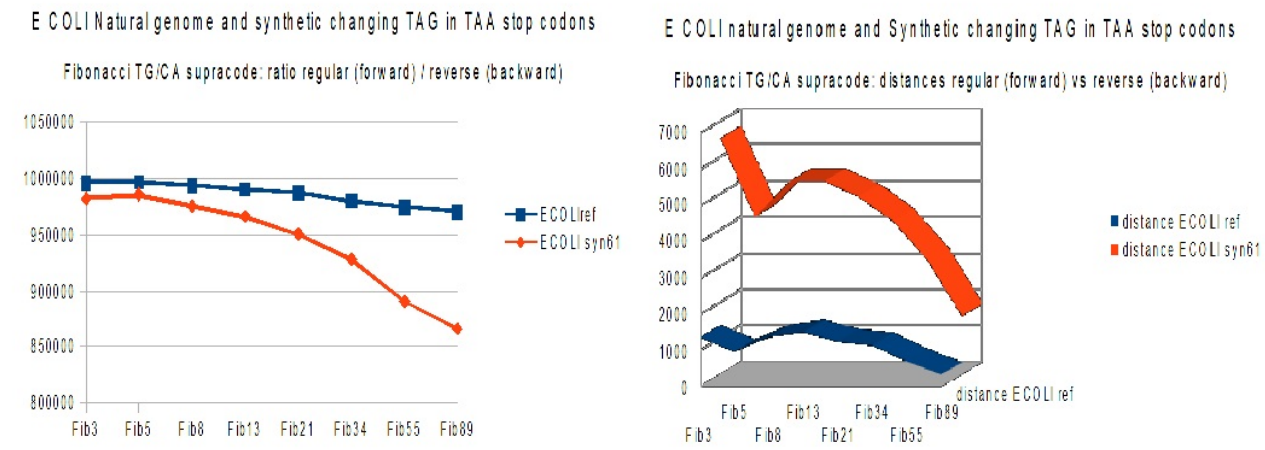

Figure 4: Left: Comparing TG/AC Fibonacci ratios in real and synthetic E-Coli genomes, Right: Comparing TG/AC Fibonacci distances in real and synthetic E-Coli genomes (regular-reverse distances weighted by the length of the genome, see detailed data in suplementary materials)

b) *Yeast Synthetic Genome, the case of the longer chromosome XII*

Since 2012 the Synthetic Yeast Genome Project (Sc2.0 http://syntheticyeast.org/sc2-0/) results from a worlwide partnership, « Sc2.0 International Consortium team », members spanning 4 continents to provide remote mentorship and solve challenges associated with synthetic individual chromosomes design features and assembly (Jee Loon Foo, 2018).

Sources

synthetic yeast project

http://syntheticyeast.org/

7 chromosomes now synthetised

http://syntheticyeast.org/sc2-0-data/

Consorsium has successfully synthesized seven chromosomes. Check the following links to learn about details related to each finished chromosomes:

- synII synIII synV synVI synIXR synX synXII

In (Weiming Zhang et al., 2017) process building the whole synthetic chromosome XII.

Having not yet obtained the synthetic genome from the authors, we have limited here our study to the concatenation of all wild type PCRTags on the one hand and synthetic ones on the other hand. For example:

Forward wild type PCRTag : TGCTTGAACTGCAAATACAGGCCCACTC

Forward synthetic PCRTag : AGCTTGGACAGCGAAAACTGGACCTGAT

They published particularly all the wild type and synthetic PCR Tags.

The full PCR Tags are available online:

http://syntheticyeast.org/wp-content/uploads/2016/10/synXII_PCRtag.txt

Details: PCRTags

« PCRTags are alterations incorporated into most open reading frames (ORFs) (on average one per ORF, as some ORFs are too small and others contain multiple PCRTags). These are made by recoding a ~20bp segments of the coding region of an ORF to a different DNA sequence encoding the same amino acid sequence. PCR primer pairs can then be designed that will selectively amplify only the synthetic or wild type sequences. In this way, transformants that have

incorporated a synthetic segment can be quickly scanned to ascertain that a complete substitution of the segment has occurred. PCRTags can also be used to monitor for the deletion of non-essential segments post-SCRaMbLE induction. » (from http://syntheticyeast.org/designs/alterations/pcrtags/).

We analysed 681 PCRTags of each 28 bp from wild YEAST XII and artificial SYN XII chromosomes. Then only resonances < 28 bp are to be considered in the following analysis.

We run 3 analysis :

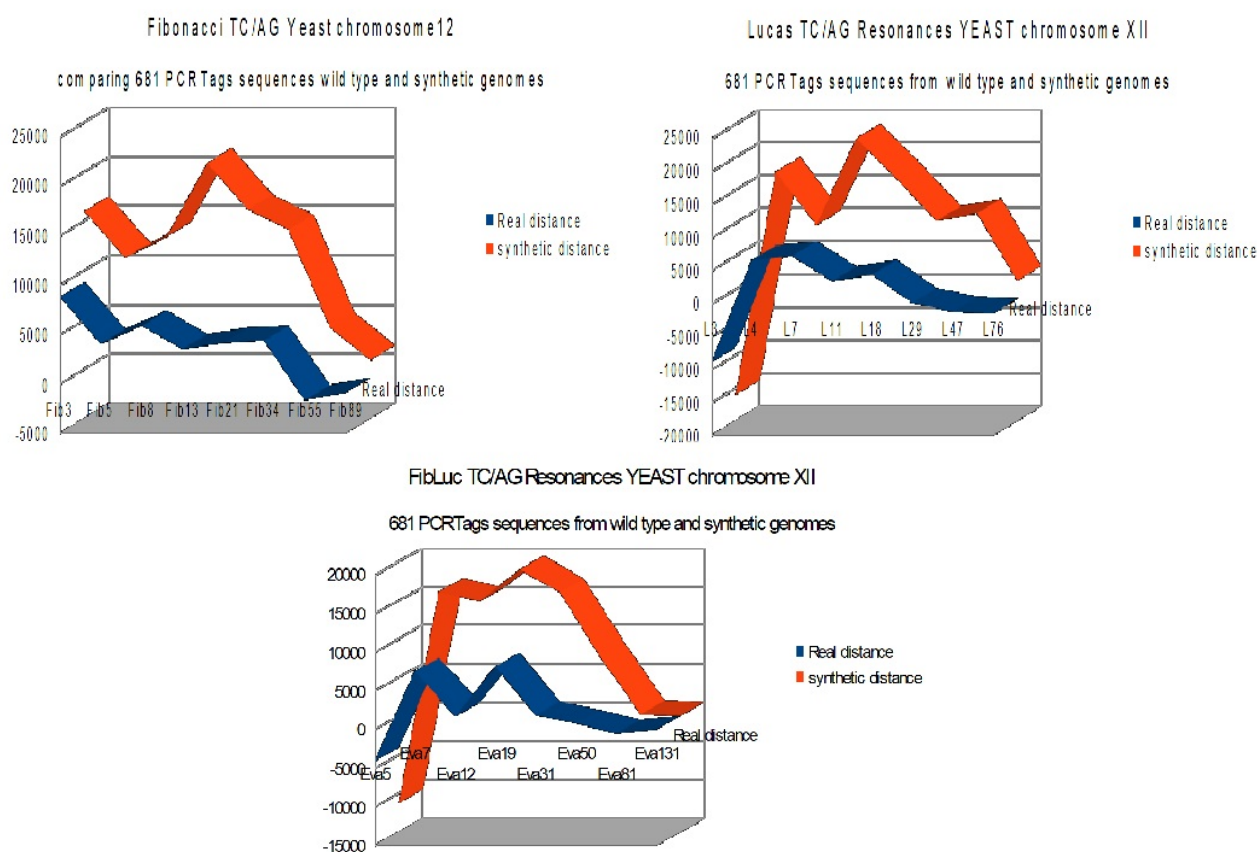Fibonacci sequence= 1 2 3 5 8 13 21 34 55 89

Lucas sequence= 1 3 4 7 11 18 29 47 76

FibLuc sequence= 5 7 12 19 31 50 81 131

Table 5: Comparing real and synthetic YEAST chromosome XII PCRTags with Fibonacci, Lucas and FibLuc resonances

YEAST XII real genome (681 wild type PCRTags)						Synthetic genome SYNXII (681 synthetic PCRTags)					
Fibonacci		Lucas		FibLuc		Fibonacci		Lucas		FibLuc	
3	6906 7073 0.976389 0853	3	7073 6906 1.024181 871	5	5726 5649 1.0136307 31	3	7133 7436 0.959252 2862	3	7436 7133 1.042478 62	5	6195 5982 1.0356068 2
5	5649 5726 0.986552 5672	4	4760 4886 0.974212 0344	7	3094 3237 0.9558232 932	5	5982 6195 0.965617 4334	4	4611 4948 0.931891 6734	7	3024 3330 0.9081081 081
8	4012 4124 0.972841 9011	7	4894 5032 0.972575 5167	12	3518 3552 0.9904279 279	8	4142 4396 0.942220 2002	7	5272 5456 0.966275 6598	12	3636 3918 0.9280245 023
13	2942 3009 0.977733 4663	11	2993 3058 0.978744 2773	19	1769 1924 0.9194386 694	13	2877 3272 0.879278 7286	11	2950 3383 0.872007 0943	19	1678 2038 0.8233562 316

21	1785 1864 0.957618 0258	18	2232 2317 0.963314 631	31	1236 1272 0.9716981 132	21	1749 2055 0.851094 8905	18	2193 2511 0.873357 2282	31	1057 1362 0.7760646 109
34	1038 1121 0.925958 9652	29	1261 1264 0.997626 5823	50	607 623 0.9743178 17	34	853 1121 0.760927 7431	29	1101 1298 0.848228 0431	50	419 566 0.7402826 855
55	538 507 1.061143 984	47	719 697 1.031563 845	81	221 212 1.0424528 3	55	358 438 0.817351 5982	47	491 710 0.691549 2958	81	172 177 0.9717514 124
89	195 179 1.089385 475	76	265 237 1.118143 46	131	37 38 0.9736842 105	89	105 122 0.860655 7377	76	188 212 0.886792 4528	131	17 19 0.8947368 421

Figure 5: Comparing TC/AG Fibonacci, Lucas and FibLuc distances in real and synthetic YEAST Chromosome XII PCRTags (regular-reverse distances weighted by the length of the genome, see detailed data in suplementary materials).

V. Conclusions

In all the cases analyzed here, we find that the real genomes or chromosomes have a property of coherence, consistency and unity that our method highlights. This property disappears in almost all (ALL) studied cases of synthetic genomes or chromosomes.

Transposons: a possible explanation of global harmonics structure

In (Weiming Zhang, 2017), authors write «RATIONALE: The synthetic yeast genome, designated Sc2.0, was designed according to a set of arbitrary rules, including the elimination of transposable elements and incorporation of specific DNA elements to facilitate further genome manipulation.»

In our article (Perez, 2010), https://www.ncbi.nlm.nih.gov/m/pubmed/20658335 we wrote:

« *Why and how could this ancient code be preserved and maintained in spite of the changes and mutations during millions of years of evolution of the human genome?"*

In the 1940's and 1950's, Nobel prize winner Barbara McClintock discovered a peculiar phenomenon in maize: certain regions of a chromosome moved, or transposed, to other positions. This was the discovery of TRANSPOSONS (Fedoroff, 1984): often called "jumping genes" because of their ability to "jump" to completely different regions within the chromosome and later "jump" back to their original positions. Meanwhile, "jumping genes" is a misleading term because transpositions are related to noncoding areas as well as coding areas. A particular class of transposons moves from one place to another. (Class II transposons consist of DNA sections that move directly from place to place).

Sometimes there is a palindrome-like swap of the transposon during this move.
Example, the original sequence:
5' TAAGGCTATGC 3'
3' ATTCCGATACG 5'

... Moves to another genome region and becomes reversed as follows:
5' GCATAGCCTTA 3'
3' CGTATCGGAAT 5'

We found the same process here. It joins a codon with its "mirror-codon". Perhaps DNA double strand topological reshaping processes could explain genesis of the reported facts (hairpin-like unfolding, Moebius-like ribbon, Class II transposons?)... »

These two observations about the role of transposons already partly explain the digital disharmony that we prove in this article. These famous transposons disrupt the functioning of synthetic genomes, so we delete them (!). On the contrary, we believe that these same transposons constitute a major piece of genome stability.

The creation by men of SYNTHETIC genomes leads to a paradox on which I invite you now to think about:

On the one hand, NATURAL DNA is a luxury of REDUNDANCY and SYMMETRY ...

On the other hand, SYNTHETIC DNA manipulation and synthesis technologies rely on and exploit the same luxury of REDUNDANCY and symmetries ... Thus; Sometimes the technology will try to EXPLOIT SYMMETRY and REDUNDANCY: this is the case of CRISPR technology based on DNA PALINDROMES, so on SYMMETRY and REDUNDANCY. Sometimes the technology will try to DESTROY symmetry and REDUNDANCY: Such is the case of mutations and alterations of transposons (Breuer, 2019)

in order to fight against these transposons which will alter the SYNTHETIC genome. This is also the case when one tries to reduce the REDUNDANCE of the universal genetic code by reducing it from 64 to 61 codons (Fredens, 2019). By our different research on the biomathematics of DNA, we have on the contrary demonstrated that this REDUNDANCY and this symmetry contribute to the UNITY and INTEGRITY of genes, chromosomes and genomes:

When a Meta-code unifies DNA, RNA and amino acids (Perez, 2009: Perez, 2011; Perez, 2015; Perez, 2018d);

When this master code unifies the genomic and proteomic meta structures of a gene (Perez, 2000; Perez, 2017e; Perez, 2017f; Perez, 2017g; Perez 2017h); When the multiple repetition of the same gene as DUF1220 is associated with mammalian brain properties via a kind of «FibLuc sequence» digital standing waves of its DNA (Sikela, 2006; Weiss, 2006; Parayon, 2011; Perez, 2017b);

When we prove the existence of a UNITY of Fibonacci sequences on the scale of an whole human chromosome such as chromosome4 (Perez, 2017c);

When we demonstrate how numerical proportions characterize the DNA of whole genomes of viruses, bacteria or Euchariotes (Perez 2013);

When we highlight the UNITY of the 3 billion base pairs of the entire human genome (Perez, 2010; Perez, 2017d);

When this whole human genome UNITY is destroyed by Cancer mutations (Perez, 2018a; Perez, 2018b; Perez, 2018c);

When there is an evidence that these numerical structures (Petoukhov, 2019) of the genomes, particularly SYMMETRY and REDUNDANCY, are of TOPOLOGICAL nature (Rapoport 2016). This topological unified hyper structure of whole genomes is based particularly on Fibonacci Numbers, Golden ratio (Friedman, 2018), and Klein bottle (Rapoport§Perez, 2018).

To conclude we will finally notice that the REAL genomes of bacteria analyzed obey two simultaneous numerical constraints of Phi and Phi * 2 (where Phi = 1.618 is the golden ratio and Phi * 2 = 2.618). For example, for a contiguous sequence of 21 TCAGs, we have simultaneously:

Regular (forward) 21 TCAG / 8 TC = Phi * 2

And

Reverse (backward) 21 TCAG / 13 TC = Phi.

This double strong constraint on REAL genomes almost disappears in the case of SYNTHETIC genomes.

We can not manipulate the genomes "no matter how". Thus, transposons certainly play a key role in the stability and epigenetics of genomes.

Manipulation technologies (CRISPR) and especially of artificial creation of genomes will have to respect these laws of nature.

In (Strecker et al., 2019) by using DNA sequences referred to as transposons, or "jumping genes" (genes that can change their position within the genome), a team from MIT led by NYSCF – Robertson Stem Cell Investigator Dr. Feng Zhang has created a new version of CRISPR (called CRISPR-associated transposase, or "CAST") that can insert functional DNA sequences into the genome without making cuts, which can often lead to unintended damage.

What about for the FUTURE? There are theoretical new background for Biology and Genetics, these tracks are MATHEMATICS (Perez § Montagnier L., 2021).

References Références Referencias

1. Blount B. A. et al, Construction of an Escherichia coli genome with fewer codons sets records, Nature 569, 492-494 (2019) doi: 10.1038/d41586-019-01584-x,

2. https://www.nature.com/articles/d41586-019-01584-x

3. Breuer M. et al., Essential metabolism for a minimal cell., Elife. 2019 Jan 18; 8 Published online 2019 Jan 18. doi: 10.7554/eLife.36842, https://www.ncbi.nlm.nih.gov/pubmed/30657448/

4. Callaway Ewen, 'Minimal' cell raises stakes in race to harness synthetic life, Nature 531, 557–558 (31 March 2016) doi: 10.1038/531557a, https://www.nature.com/news/minimal-cell-raises-stakes-in-race-to-harness-synthetic-life-1.19633

5. Fedoroff, N.V. (1984), Transposable genetic elements in maize. Scientific American 250, 84–98.

6. Fleaux Rachel, « La musique des genes ». Scientific journal Sciences et avenir , April 1995.

7. Fraser CM, Gocayne JD, White O, Adams MD, Clayton RA, Fleischmann RD, et al. (1995), The minimal gene complement of mycoplasma genitalium. Science. 1995;270:397–403. [PubMed] [Google Scholar] https://science.sciencemag.org/content/270/5235/397

8. Fredens J. et al., Total synthesis of *Escherichia coli* with a recoded genome, Nature, May 2019, DOI https://doi.org/10.1038/s41586-019-1192-5 , https://www.nature.com/articles/s41586-019-1192-5

9. Robert Friedman and Matthew Cross, (2018), Nature's Secret Nutrient: Golden Ratio Biomimicry for PEAK Health, Performance & Longevity, https://www.amazon.fr/Natures-Secret-Nutrient-Biomimicry-Performance-ebook/dp/B07N8LHDQ1

10. Hutchinson III et al, Design and synthesis of a minimal bacterial genome, *Science* 25 Mar 2016: Vol. 351, Issue 6280, aad6253 DOI: 10.1126/science.aad6253; https://science.sciencemag.org/node/676507.full

11. Gibson DG, Glass JI, Lartigue C, Noskov VN, Chuang RY, Algire MA, et al. (2010), Creation of a bacterial cell controlled by a chemically synthesized genome. Science. 2010;329:52–56. [PubMed] [Google Scholar]

12. Jee Loon Foo, Synthetic yeast genome reveals its versatility, *Nature* 557, 647-648 (2018), *doi: 10.1038/d41586-018-05164-3* https://www.nature.com/articles/d41586-018-05164-3/

13. Marcer P, Communications: Order and Chaos in DNA — the Denis Guichard Prizewinner: Jean-Claude Pérez, Kybernetes, Vol. 21, 1992, Issue: 2, pp.60 61, https://doi.org/10.1108/eb005922.

14. Orsini M. et al, Whole-Genome Sequencing of Mycoplasma mycoides subsp. mycoides Italian Strain 57/13, the Causative Agent of Contagious Bovine Pleuropneumonia.

15. Genome Announc. 2015 Mar 26;3(2). pii: e00197-15. doi: 10.1128/genomeA.00197-15.

16. https://www.ncbi.nlm.nih.gov/pubmed?LinkName=nuccore_pubmed&from_uid=731474337

17. Parayon Gabriel, (2011), On musical self-similarity, ISBN 978-952-10-6970-3 (PDF):

18. https://helda.helsinki.fi/bitstream/handle/10138/26236/onmusica.pdf

19. (Pellionisz et al ; 2012), Pellionisz A.J., Graham R., Pellionisz P.A., Perez JC. (2013) Recursive Genome Function of the Cerebellum: Geometric Unification of Neuroscience and Genomics. In: Manto M., Schmahmann J.D., Rossi F., Gruol D.L., Koibuchi N.

(eds) Handbook of the Cerebellum and Cerebellar Disorders. Springer, Dordrecht. https://doi.org/10.1007/978-94-007-1333-8_61

20. Pérez J.C., De nouvelles voies vers l'Intelligence Artificielle - 1988 - Ed. MASSON Paris - ISBN: 2-225-81326-4

21. Pérez JC (1989). De nouvelles voies vers l'Intelligence Artificielle. Ed. Masson Paris Second edition, ISBN: 2-225-81815-0

22. Pérez JC (1990a). La revolution des ordinateurs neuronaux. Hermes Ed. Paris, ISBN: 2-86601 -203-8. https://www.amazon.fr/R%C3%A9volution-ordi-nateurs-neuronaux-Jean-ClaudePérez/dp/2866012038

23. Pérez JC (1990b). Integer neural network systems (1.N.N.S) using resonance properties of a Fibonacci chaotic Golden Neuron, Published in: 1990 IJCNN International Joint Conference on Neural Networks, INSPEC Accession Number: 3926657 DOI: 10.1109/IJCNN.1990.137678 Publisher: IEEE Conference Location: San Diego, CA, USA, USA, https://ieeexplore.ieee.org/document/57266 38/

24. Pérez JC (1990c) Digital Holograms Computers: concepts and apllications. International conference on Neural Networks - LYON France March 1990 ; Springer Verlag. ISBN2-287-0005ISBN2- 287-00051-8 1-8.

25. Pérez JC, Chaos DNA and neurocomputers. https://www.researchgate.net/publication/258439719_JC_Pérez_1991_Chaos_DNA_and_Neurocomputers_A_Golden_Link_in_Speculations_in_Science_and_Technologyvol_14_no_4_ISSN_0155-7785

26. Perez j.C, (1997), « L'ADN décrypté » (Decyphered DNA), Resurgence publisher Liege, Belgium. https://www.amazon.co.uk/LADN-deciphered-security-language-experts/dp/2872110178/ref=asap_bc?ie=UTF8).

27. Pérez JC (2000). From Prions and Prions-like invariants to the self-assembly thesis. International symposium on Prion Diseases and Elated Processes. Annecy, France.

28. Pérez JC (2009) Codex Biogenesis. Resurgence, Liege Belgium. https://www.amazon.fr/Codex-Biogenesis13-codes-lADN/dp/2874340448

29. Perez J.C., Codon populations in single-stranded whole human genome DNA Are fractal and fine-tuned by the Golden Ratio 1.618., Interdiscip Sci. 2010 Sep;2(3):228-40. doi: 10.1007/s12539-010-0022-0. Epub 2010 Jul 25., https://www.ncbi.nlm.nih.gov/m/pubmed/20658335/

30. Pérez JC (2011), BIT Life Sciences' 3rd World Congress of Vaccine, Beijing, China" Decoding non-coding Dna Codes:Human Genome Meta Chromosomes Architecture " http://fr.scribd.com/doc/57828784/jcPérezB eijing032011

31. Pérez JC, Applied Mathematics, The "3 Genomic Numbers" Discovery: How Our Genome Single-Stranded DNA Sequence Is "Self-Designed" as a Numerical WholeVol.4 No.10B(2013), Article ID:37457,17 DOI: 10.4236/am.2013.410A2004. http://fr.scribd.com/doc/57828784/jcPérezB eijing032011

32. Perez JC (2015). Deciphering Hidden DNA Meta-Codes -The Great Unification & Master Code of Biology. J Glycomics Lipidomics 5:131. doi: 10.4172/2153- 0637.1000131 https://www.omics-online.org/openaccess/deciphering-hidden-dna-metacodesthe-great-unification--mastercode-of-biology-2153-0637- 1000131.php?aid=55261

33. Pérez JC (2017a). Sapiens Mitochondrial DNA Genome Circular Long Range Numerical Meta Structures are Highly Correlated with Cancers and Genetic Diseases mtDNA Mutations. J Cancer Sci Ther 2017, 9:512- 527. doi: 10.4172/1948-5956.1000469, https://www.omicsonline.org/open-access/sapiens-mitochondrial-dna-genomecircular-long-range-numericalmetastructures-are-highly-correlated-withcancers-and-genetic-disea-1948-5956- 1000469.php?aid=90737

34. Pérez JC (2017b). DUF1220 Homo Sapiens and Neanderthal fractal periods architectures breakthrough(2017) SDRP Journal of Cellular and Molecular Physiology https://www.siftdesk.org/articledetails/DUF1220-Homo-Sapiens-andNean-derthal--fractal-periods-architecturesbreakthrough/184

35. Pérez JC (2017c). The exceptional fractal meta organizations of whole chromosomes 4 of Sapiens, Neanderthal and superior primates, Bioinformatics and Computational BiologyLetters, 2017, 1 (1): 1-26, DOI: 10.24896/bcbl.2017112 https://bacbl.com/index.php/bcbl/article/vie w/2/2

36. Pérez J C (2017d). A proof of the unity, integrity and autopoietic autonomy of the whole human genome . Research in Biological Chemistry 2017, vol1, no. 1, https://www.rbchemistry.com/index.php/rbc/article/view/2

37. Pérez JC (2017e). The Master Code of Biology: from Prions and Prions-like Invariants to the Self-assembly Thesis. Biomed J Sci&Tech Res1(4)-2017. BJSTR.MS.ID.000369. DOI: 10.26717/BJSTR.2017.01.000369 https://biomedres.us/pdfs/BJSTR.MS.ID.00 0369.pdf

38. Pérez JC (2017f). Decyphering "the MASTER CODE ®" Structure and Discovery of a Periodic Invariant Unifying 160 HIV1/HIV2/SIV Isolates Genomes. Biomed J Sci & Tech Res 1(2)-2017. BJSTR.MS.ID.000209. http://biomedres.us/pdfs/BJSTR.MS.ID.000 209.pdf

39. Pérez JC (2017g). The "Master Code of DNA: Towards the Discovery of the SNPs Function (Single-Nucleotide Polymorphism). J Clin Epigenet. Vol. 3 No. 3:26, http://clinicalepigenetics.imedpub.com/the-master-codeof-dna-towards-the-discovery-of-the-snpsfunction-singlenucleotidepolymorphism.

pdf Pérez JC (2017e), Mannheim Germany 27 November 2017 Biomathematics conference in Mannheim Germany https://www.cammbio. hsm-annheim.de/kick-off-event.html … Dr. Jean-Claude Pérez, https://www.cammbio.hsmannheim.de/file-admin/user_upload/projek te/cammbio/events/20171127-kickoff/EPPérez.pdf

40. Pérez JC (2017h). The Master Code of Biology: Self-assembly of two identical Peptides beta A4 1-43 Amyloid In Alzheimer's Diseases. Biomed J Sci & Tech Res 1(4)- 2017. BJSTR.MS.ID.000394. DOI: 10.26717/BJSTR.2017.01.000394, https://biomedres .us/pdfs/BJSTR.MS.ID.00 0394.pdf

41. Pérez JC (2018a). Towards a Universal Law Controlling all Human Cancer Chromosome LOH Deletions, Perspectives in Prostate and Breast Cancers screening. Canc Therapy & Oncol Int J. 2018; 9(2): 555756. DOI: 10.19080/CTOIJ.2018. 09.555756. https://juniperpublishers.com/ctoij/pdf/ CTO IJ.MS.ID.555756.pdf

42. Pérez JC (2018b). Cancer, Quantum Computing and TP53 Tumor Suppressor Gene Mutations Prediction. Nov Appro in Can Study. 1(2). NACS.000507.2018. http://crimsonpublishers.com/ nacs/pdf/NA CS.000507.pdf

43. Pérez JC (2018c). Neuroblastoma and Glioblastoma Brain Cancers:«Human Genome Optimum» (HGO) a Global Genome Strategy controlling all Human Chromosome LOH Deletions. Nov Appro in Can Study. 1(3). NACS.000512.2018.http://crimson publishe rs.com/nacs/pdf/NACS.000512.pdf

44. Perez, J. C., & Montagnier, L. . (2021). Six Fractal Codes Of Life From Bioatoms Atomic Mass To Chromosomes Numerical Standing Waves: Three Breakthoughs In Astrobiology, Cancers And Artificial Intelligence. *International Journal of Research - GRANTHAALAYAH*, *9*(9), 133–191. https://doi.org/ 10.29121/granthaalayah.v9.i9.2021.4191

45. Perez J.C. (2019), 20219 paper to be published, « The Discovery of a "DNA SUPRACODE" Controlling Locally & Globally Humans Mitochondrial Genomes: Perspectives in Evolution and Cancers Early stage Diagnostic ».

46. (Perez§Montagnier, 2021), Perez and Montagnier (2021) - Perez, J. C., & Montagnier, L. . (2021). Six Fractal Codes Of Life From Bioatoms Atomic Mass To Chromosomes Numerical Standing Waves: Three Breakthoughs In Astrobiology, Cancers And Artificial Intelligence. International Journal of Research - Granthaalayah, 9(9), 133–191. DOI: https://doi.org/ 10.29121/granthaalayah.v9.i9.2021.4191

47. Sergey V. Petoukhov, Zhengbing Hu, Matthew He, Advances in Artificial Systems for Medicine and Education II, DOIhttps://doi.org/10.1007/978-3-030-12082-5, Springer Nature Switzerland AG 2020, https://link.springer.com/book/10.1007%2F978-3-030-12082-5#page=40

48. Rapoport D.L., Klein Bottle Logophysics, Self-reference, Heterarchies, Genomic Topologies, Harmonics and Evolution. Part III: The Klein Bottle Logic of Genomics and its Dynamics, Quantum Information, Complexity and Palindromic Repeats in Evolution Quantum Biosystems | November 2016 | Vol 7 | Issue 1 | Page 107-174 107 ISSN 1970-223X www.quantumbiosystems.org

49. Rapoport D. L. and Perez J.C. (2018) Golden ratio and Klein bottle Logophysics: the Keys of the Codes of Life and Cognition. *Quantum Biosystems. 9(2) 8-76.*Vol. 9 – n.2 – 2018

50. Sikela, J. M., The jewels of our genome: the search for the genomic changes underlying the evolutionarily unique capacities of the human brain. PLoS Genetics 2 (2006) doi.org/10.1371/journal. pgen.0020080

51. Sleator RD., The story of Mycoplasma mycoides JCVI-syn1.0: the forty million dollar microbe. *Bioeng Bugs*. 2010;1(4):229–230. doi:10.4161/bbug.1.4. 12465

52. Strecker J, Ladha A, Gardner Z, Schmid-Burgk JL, Makarova KS, Koonin EV, Zhang F., RNA-guided DNA insertion with CRISPR-associated trans-posases., *Science*. 2019 Jun 6. doi: 10.1126/ science.aax9181. https://doi.org/10.1126/science. aax9181

53. Venetz JE et al, Chemical synthesis rewriting of a bacterial genome to achieve design flexibility and biological functionality, PNAS 2019, doi: 10.1073/ pnas.1818259116

54. Weiming Zhang et al., Engineering the ribosomal DNA in a megabase synthetic chromosome *Science* 10 Mar 2017: Vol. 355, Issue 6329, eaaf3981 DOI: 10.1126/science.aaf3981

55. Weiss H and Weiss V., The golden mean as clock cycle of brain waves - Chaos, Solitons & Fractals, 2003.